# nature metabolism

Article

# Effects of different doses of exercise and diet-induced weight loss on beta-cell function in type 2 diabetes (DOSE-EX): a randomized clinical trial

Received: 18 January 2023

Accepted: 4 April 2023

Published online: 1 May 2023

Check for updates

A list of authors and their affiliations appears at the end of the paper

Diet-induced weight loss is associated with improved beta-cell function in people with type 2 diabetes (T2D) with remaining secretory capacity. It is unknown if adding exercise to diet-induced weight loss improves beta-cell function and if exercise volume is important for improving beta-cell function in this context. Here, we carried out a four-armed randomized trial with a total of 82 persons (35% females, mean age (s.d.) of 58.2 years (9.8)) with newly diagnosed T2D (<7 years). Participants were randomly allocated to standard care (n = 20), calorie restriction (25% energy reduction; n = 21), calorie restriction and exercise three times per week (n = 20), or calorie restriction and exercise six times per week (n = 21) for 16 weeks. The primary outcome was beta-cell function as indicated by the late-phase disposition index (insulin secretion multiplied by insulin sensitivity) at steady-state hyperglycemia during a hyperglycemic clamp. Secondary outcomes included glucose-stimulated insulin secretion and sensitivity as well as the disposition, insulin sensitivity, and secretion indices derived from a liquid mixed meal tolerance test. We show that the late-phase disposition index during the clamp increases more in all three intervention groups than in standard care (diet control group, 58%; 95% confidence interval (CI), 16 to 116; moderate exercise dose group, 105%; 95% CI, 49 to 182; high exercise dose group, 137%; 95% CI, 73 to 225) and follows a linear dose-response relationship (P > 0.001 for trend). We report three serious adverse events (two in the control group and one in the diet control group), as well as adverse events in two participants in the diet control group, and five participants each in the moderate and high exercise dose groups. Overall, adding an exercise intervention to diet-induced weight loss improves glucose-stimulated beta-cell function in people with newly diagnosed T2D in an exercise dose-dependent manner (NCT03769883).

As the progressive deterioration of normal beta-cell function is regarded as a determining factor for the onset and subsequent progression of T2D, re-establishing beta-cell function is considered pivotal to improving the pathogenesis of T2D<sup>1</sup>.

Although a substantial diet-induced weight loss is consistent with improved beta-cell function<sup>2-4</sup>, the effects of exercise on beta-cell function in T2D are not well understood<sup>5-9</sup>. Inconsistent findings may relate to differences in concomitant pharmacological therapy, the

e-mail: mathias.ried-larsen@regionh.dk

participants' pretrial insulin secretory capacity, or differences in exercise modality, intensity and/or volume<sup>10-14</sup>. The inconsistencies could also be related to a failure to correct for prevailing insulin sensitivity when assessing beta-cell function. As the normal physiological response to decreased insulin sensitivity is an increase in insulin secretion, the assessment of beta-cell function should incorporate both measures (that is, insulin sensitivity and secretion)<sup>15</sup>. A widely accepted measure of beta-cell function is the disposition index (DI), that is, the product of insulin sensitivity and insulin secretion<sup>15</sup>. Whereas there is evidence to suggest that exercise-induced improvements in DI are explained via improvements in insulin sensitivity and glucose disposal, the exercise-induced effects on insulin secretion in the context of prevailing insulin sensitivity remain to be clarified<sup>5,10,16</sup>.

Intensive structured weight management programs aiming for weight loss are recommended alongside pharmacological therapy to treat hyperglycemia<sup>17</sup>. Diet-induced weight loss is consistent with improvements in beta-cell function<sup>2,18</sup>, and glucose-lowering medications may increase insulin sensitivity, insulin secretion and incretin responses<sup>19–21</sup>. Therefore, potential interactions between these therapies and exercise should be considered when assessing the role of exercise on DI in people with T2D in a clinical setting. As such, there is a need to investigate the potential effects of exercise on DI in the context of standardized dietary weight loss and pharmacological therapy.

Accordingly, the primary objective of this study was to investigate the change in DI during the final 30 min of clamp-induced hyperglycemia (late-phase DI) after a 16-week intervention with different volumes of exercise in addition to diet-induced weight loss and algorithm-guided pharmacological management in people with newly diagnosed T2D. We hypothesized that late-phase DI would increase with increasing volumes of exercise in combination with diet-induced weight loss. Furthermore, we expected that both moderate and high volumes of exercise in combination with a diet-induced weight loss intervention would be superior to the control in improving late-phase DI<sup>22</sup>. The secondary objective was to assess the effects of the intervention on insulin sensitivity and secretion. Moreover, we aimed to explore the effects on cardiometabolic risk factors, postprandial glucose metabolism, glucose kinetics, glucagon-like peptide 1 (GLP-1) sensitivity and maximal insulin secretory capacity.

## Results

#### Trial population and adherence to the intervention

Eighty-two persons were included in the study (Fig. 1). Five participants were lost to follow-up: one was due to malignancy, one was dissatisfied with group allocation, one refrained from study testing due to COVID-19, and two were due to musculoskeletal injuries. The mean (s.d.) age was 58.2 years (9.8), body mass index (BMI) was 33 kg/m<sup>2</sup> (3.7), and glycated hemoglobin (HbA1c) was 50.2 mmol/mol (6.6). Thirty-five percent (35%) of participants were females, and the median (interquartile range (IQR)) T2D duration was 4.0 years (1.9 to 5.5). Baseline characteristics are presented in Table 1. Mean (s.d.) adherence to the prescribed diet intervention (~25-30% energy deficit per day) was 92% (11) for the diet control group (DCON), 91% (18) for the moderate exercise dose group (MED), and 88% (13) for the high exercise dose group (HED) (Supplementary Table 1). Mean (s.d.) adherence to the prescribed exercise protocol was 86% (28) and 93% (18) for HED and MED, respectively (Supplementary Tables 2-7). No compensatory decrease in total free-living physical activity was seen in the intervention groups during the intervention period (Supplementary Table 8). In-study adherence to the predefined pharmacological treatment was similar among all groups (Supplementary Table 9).

#### **Primary outcome**

The late-phase DI increased in all intervention groups from baseline to 16-week follow-up with no change in the control group (CON) (Fig. 2a) in the intention-to-treat (ITT) analysis; as such, all intervention groups increased more than CON (P < 0.005 for all comparisons; Table 2). Compared with DCON, both MED and HED increased the late-phase DI (MED versus DCON, 29% (95% CI, -5 to 77), P = 0.11; HED versus DCON, 50% (95% CI, 10 to 104), P = 0.01) (Table 2). The magnitude of increases across groups was consistent with a linear dose–response relationship (P for trend <0.001). The per-protocol (PP) analysis set consisted of CON n = 18 (90%), DCON n = 21 (100%), MED n = 19 (95%) and HED n = 18 (86%), and followed the pattern observed in ITT (Table 2). The distribution of absolute values at baseline and follow-up are presented in Extended Data Fig. 1.

#### Secondary outcomes

The dose–response relationship observed for the late-phase DI was also reflected in the late-phase glucose-stimulated insulin sensitivity index (ISI) (*P* for trend <0.001), where both MED and HED increased more than CON, although the difference between DCON and CON was less pronounced (Fig. 2b). HED was associated with a greater increase in late-phase ISI compared with DCON (55% (95% CI, 15 to 109), P = 0.004). No differences were observed in late-phase ISI between DCON and CON. Late-phase glucose-stimulated insulin secretion rate (ISR) increased more in all intervention groups than in CON (Fig. 2c, Table 2 and Supplementary Tables 10 and 11), but no differences were observed among the remaining groups.

DI derived from the mixed meal tolerance test (MMTT) (oral DI) increased more in all intervention groups than in CON (Fig. 2d). The MED and HED groups increased more than DCON (MED versus DCON, 25% (95% CI, -5 to 65), P = 0.12; HED versus DCON, 29% (95% CI, -2 to 70), P = 0.065) with no signs of additional increases in HED versus MED (4% (95% CI, -22 to 37), P = 0.81).

All groups increased oral ISI compared with CON (P < 0.001) (Fig. 2e), with more pronounced increases in HED than DCON (29% (95% CI, 3 to 62), P = 0.025) (Table 2). No differences were observed in the oral ISR between the groups (Fig. 2f).

#### Safety outcomes

Three serious adverse events were observed: one case of transient ischemic attack and one case of malignant melanoma in the CON group, and one case of prolactinoma in the DCON group (Table 3). Two participants in the DCON group, and five participants each in the MED and HED groups reported adverse events. Beyond musculoskeletal complaints and overuse injuries in MED and HED, the nature and frequency were similar between groups.

#### **Exploratory outcomes**

Supporting clamp-derived indices of beta-cell function. The first-phase (0-10 min of clamp-induced hyperglycemia) DI increased in all intervention groups for all comparisons with CON ( $P^{<}0.001$ ; Supplementary Table 11). In addition, both HED and MED increased the first-phase DI more than DCON (HED versus DCON, 37% (95% CI, 6 to 77), P = 0.001; MED versus DCON, 58% (95% CI, 22 to 105), P = 0.017). No difference was observed between HED and MED (Supplementary Table 11). Peak and mean ISR in response to GLP-1 and GLP-1 + arginine infusion increased more from baseline to follow-up in all intervention groups compared with CON (Fig. 3 and Supplementary Tables 10 and 11). Whereas HED did not increase ISR in response to GLP-1 compared with DCON, MED was associated with increased ISR in response to GLP-1 compared with DCON (peak ISR, 0.2 (pmol/kg/min) × mM<sup>-1</sup>  $(95\% \text{ CI}, 0.0 \text{ to } 0.3), P = 0.019; \text{ mean ISR}, 0.3 (pmol/kg/min) \times \text{mM}^{-1}$ (95% CI, 0.05 to 0.6), P = 0.045). All intervention groups increased ISR in response to arginine, but no consistent differences were observed among the intervention groups (Supplementary Table 11).

**Glucose kinetics.** The change in basal rate of glucose appearance  $(R_a)$  and disappearance  $(R_d)$ , and thus the basal endogenous glucose production (EGP), was increased only in HED compared with CON,



Fig. 1 | Flow of participants. CON, control group; DCON, diet control group; MED, moderate exercise dose group; HED, high exercise dose group; ITT, intention-to-treat; PP, per-protocol.

but no additional differences between the groups were observed. Late-phase  $R_d$  and  $R_a$  increased more in all intervention groups than in CON (P < 0.001 for all comparisons); HED increased more than DCON (difference in  $R_d$ , 0.8 (95% CI, 0.2 to 1.4), P = 0.012; difference in  $R_a$ , 0.7 (95% CI, 0.1 to 1.3), P = 0.022). Complete data on glucose infusion rate (GIR), EGP,  $R_d$  and  $R_a$  are presented in Extended Data Fig. 2 and Supplementary Tables 10 and 11.

**Postprandial glucose metabolism.** Postprandial plasma glucose and insulin decreased more in all intervention groups than in CON (total area under the curve (tAUC), t = 0-120 min, P < 0.001). No differences were observed between intervention groups (Supplementary Tables 10 and 11; incremental AUC (iAUC) is shown in Supplementary Tables 14 and 15). The tAUC for GLP-1 and gastric inhibitory polypeptide (GIP) secretion increased more in CON than in the intervention groups from baseline to follow-up (Supplementary Tables 10 and 11). No differences

were observed between the intervention groups (Supplementary Tables 10 and 11 and iAUC in Supplementary Tables 14 and 15). Postprandial responses are presented in Extended Data Figs. 3–8.

**Body weight.** Body weight decreased by 0%, 7%, 10% and 12% from baseline in CON, DCON, MED and HED, respectively, and decreased more in all intervention groups than in CON (Tables 4 and 5). Both MED and HED reduced the body weight by 3.2 kg (P = 0.043) and 4.5 kg (P = 0.004) more than DCON, respectively, with no difference in changes between the exercising groups. The same pattern was observed for BMI (Tables 4 and 5).

**Other cardiometabolic markers.** HbA1c decreased 0.6% (7 mmol/mol) more in all intervention groups than in CON (P < 0.001), but no differences were observed between intervention groups (Tables 4 and 5). The same pattern was observed for fasting glucose, fasting insulin,

#### Table 1 | Baseline characteristics

	CON	DCON	MED	HED	Total
	Mean (s.d.)	Mean (s.d.)	Mean (s.d.)	Mean (s.d.)	Mean (s.d.)
	n=20	n=21	n=20	n=21	n=82
Clinical and cardiometabolic measurement	S				
General					
Age, years	59.1 (9.2)	55.9 (10.0)	60.9 (7.6)	57.3 (11.8)	58.2 (9.8)
Sex, female, n (%)	7.0 (35.0)	7.0 (33.3)	7.0 (35.0)	8.0 (38.1)	29.0 (35.4)
T2D duration, median (IQR), years	3.5 (1.2; 5.4)	3.9 (2.9; 4.8)	4.1 (2.1; 5.9)	4.2 (3.4; 5.5)	4.0 (1.9; 5.5)
Glycemic control					
HbA1c, mmol/mol	52 (7)	49 (7)	51 (6)	49 (7)	50 (7)
HbA1c, %	6.9 (0.6)	6.7 (0.7)	6.8 (0.6)	6.7 (0.6)	6.7 (0.6)
Fasting glucose, median (IQR), mmol/l	9.1 (7.3; 10.3)	7.8 (7.3; 9.8)	7.7 (7.1; 9.6)	7.8 (7.0; 9.6)	7.8 (7.1; 10.0)
Fasting insulin, median (IQR), pmol/l	129 (95; 157)	149 (93; 198)	138 (88; 219)	127 (95; 166)	128 (91; 184)
Fasting C-peptide, pmol/l	1,206 (327)	1,247 (301)	1,387 (482)	1,248 (306)	1,271 (360)
Lipids					
LDL-C, mmol/l	3.2 (0.8)	2.9 (0.7)	2.9 (0.6)	2.6 (0.6)	2.9 (0.7)
Fasting triglycerides, mmol/l	1.5 (1.3)	1.5 (1.2)	1.8 (1.3)	1.4 (1.1)	1.5 (1.1)
Blood pressure					
Systolic, mmHg	128 (12)	127 (10.3*)	133 (11.2**)	129 (11)	129 (11)
Diastolic, mmHg	78 (6)	80 (6.7*)	81 (7.5**)	78 (7)	79 (7)
Glucose-lowering medication, n (%)					
None	5.0 (25.0)	8.0 (38.1)	5.0 (25.0)	5.0 (23.8)	23.0 (28.0)
Biguanide	9.0 (45.0)	7.0 (33.3)	11.0 (55.0)	9.0 (42.9)	36.0 (43.9)
Biguanide + SGLT2i or DPP4i	5.0 (25.0)	5.0 (23.8)	4.0 (20.0)	6.0 (28.6)	20.0 (24.4)
Biguanide + SGLT2i + DPP4i	1.0 (5.0)	1.0 (4.8)	0.0 (0.0)	1.0 (4.8)	3.0 (3.7)
Lipid-lowering medication, n (%)					
None	7.0 (35.0)	7.0 (33.3)	9.0 (45.0)	5.0 (23.8)	28.0 (34.1)
Statin	13.0 (65.0)	14.0 (66.7)	11.0 (55.0)	16.0 (76.2)	54.0 (65.9)
Blood pressure-lowering medication, n (%)					
None	11.0 (55.0)	9.0 (42.9)	11.0 (55.0)	6.0 (28.6)	37.0 (45.1)
ARB or ACEi	4.0 (20.0)	5.0 (23.8)	4.0 (20.0)	6.0 (28.6)	19.0 (23.2)
ARB or ACEi + thiazide or CCB	4.0 (20.0)	4.0 (19.0)	4.0 (20.0)	6.0 (28.6)	18.0 (22.0)
ARB or ACEi + thiazide + CCB	1.0 (5.0)	3.0 (14.3)	1.0 (5.0)	3.0 (14.3)	8.0 (9.8)
Physical fitness					
Absolute VO <sub>2max</sub> , ml/min	2,445.6 (413.2)	2,611.6 (618.2)	2,512.9 (634.5)	2,582.0 (714.4)	2,539.5 (599.2)
Relative VO <sub>2max</sub> , ml/kg/min	24.5 (3.7)	25.7 (3.6)	24.7 (4.6)	24.8 (4.3)	24.9 (4.0)
Watt max	192.8 (41.3)	204.8 (48.5)	189.7 (49.6)	204.8 (66.6)	198.2 (52.0)
1 RM chest press, median (IQR), kg	40.0 (35.0; 55.0**)	47.5 (35.0; 57.5)	45.0 (35.0; 57.5***)	52.5 (25.0; 65.0)	45.0 (35.0; 57.5)
1 RM leg extension, kg	68.0 (20.9*)	75.2 (24.4*)	63.6 (17.3)	68.0 (22.4)	68.7 (21.4)
Body anthropometrics					
Body weight, kg	100.3 (12.3)	100.8 (15.4)	101.6 (16.1)	102.8 (15.2)	101.4 (14.6)
BMI, kg/m <sup>2</sup>	32.4 (3.6)	33.2 (3.8)	33.2 (4.1)	33.4 (3.5)	33.1 (3.7)
Diet					
Energy intake, median (IQR), kcal/day	1,976.5 (1,772.0; 2,485.0)	1,990.0 (1,843.0; 2,247.0)	2,052.0 (1,800.0; 2,389.0**)	2,052.0 (1,571.0; 2,586.0)	1,995.0 (1,784.0; 2,465.0)
Hyperglycemic clamp					
Basal					
Mean ISR, median (IQR)	122 (72; 142)	110 (80; 175)	128 (82; 183)	100 (81; 167)	124 (78; 166)
Glucose R <sub>a</sub> , mg/kg/min	1.7 (0.2***)	1.7 (0.5**)	1.9 (0.5**)	1.8 (0.3*)	1.8 (0.4)

#### Table 1 (continued) | Baseline characteristics

	CON	DCON	MED	HED	Total
	Mean (s.d.)	Mean (s.d.)	Mean (s.d.)	Mean (s.d.)	Mean (s.d.)
Glucose R <sub>d</sub> , mg/kg/min	1.6 (0.3***)	1.6 (0.5**)	1.8 (0.5**)	1.7 (0.3*)	1.7 (1.0)
EGP, mg/kg/min	1.7 (0.2***)	1.7 (0.5**)	1.8 (0.5**)	1.7 (0.3*)	1.7 (0.4)
First-phase hyperglycemia (0–10 min)					
Mean GIR, mg/kg/min	10.7 (0.6)	10.6 (0.7)	10.6 (0.8)	10.6 (0.7)	10.6 (0.7)
Mean ISR, median (IQR), mmol/kg/min	106 (82; 143)	125 (77; 220)	123 (83; 190)	110 (83; 165)	116 (81; 173)
Peak ISR, median (IQR), mmol/kg/min	122 (92; 159)	132 (94; 229)	148 (103; 238)	136 (97; 173)	133 (94; 210)
Late-phase hyperglycemia (90–120 min)					
Late-phase DI, a.u.	1.6 (0.7)	1.6 (0.8)	1.5 (0.9)	1.9 (1.4)	1.7 (1.0)
Late-phase ISI, mmol/kg/min	3.0 (1.9; 5.1)	3.1 (2.3; 5.2)	2.7 (2.0; 3.9)	3.2 (2.2; 5.4)	3.0 (2.0; 4.8)
Late-phase ISR, median (IQR), pmol/kg/ min/mmol/l	239 (187; 459)	282 (185; 388)	317 (208; 532)	297 (211; 417)	282 (189; 446)
Mean GIR, mg/kg/min	2.5 (0.9)	2.4 (0.7)	2.6 (1.4)	2.8 (1.9)	2.6 (1.3)
Peak ISR, median (IQR)	253 (201; 525)	307 (198; 430)	355 (226; 563)	310 (234; 424)	309 (202; 503)
Glucose R <sub>a</sub> , mg/kg/min	4.1 (0.6***)	4.1 (0.7**)	4.4 (0.9**)	4.1 (1.1*)	4.2 (0.9)
Glucose R <sub>d</sub> , mg/kg/min	3.8 (0.7***)	4.3 (1.2**)	4.3 (0.9**)	4.7 (3.4*)	4.3 (1.9)
EGP, mg/kg/min	1.5 (0.8***)	1.6 (0.9**)	1.6 (1.2**)	1.4 (0.9*)	1.6 (0.9)
Hyperglycemia and GLP-1 (120–180 min)					
Mean GIR, median (IQR), mg/kg/min	4.4 (3.3; 5.4)	4.0 (3.5; 6.2)	5.1 (3.4; 7.9)	4.6 (3.2; 8.8)	4.5 (3.3; 6.9)
Mean ISR, median (IQR)	506 (312; 1,009)	647 (423; 887)	783 (408; 1,343)	609 (411; 1,043)	629 (376; 1,048)
Peak ISR, median (IQR)	680 (420; 1,627)	914 (577; 1,402)	1274 (624; 2,177)	1049 (600; 1,778)	943 (534; 1,817)
Hyperglycemia, GLP-1 and arginine (180–190 min)					
Mean ISR, median (IQR)	1,371 (1,027; 2,768.5***)	2,130 (1,504; 2,948.2***)	1,592 (1,228; 4,044.8****)	2,342 (1,151; 3,423.8***)	1,947 (1,163; 3,215)
Peak ISR, median (IQR)	1,937 (1,490; 3,579.0***)	3,227 (2,183; 3,775.0***)	2,256 (1,607; 5,048.0****)	3,011 (1,638; 4,746.0***)	2,818 (1,763; 4,050)
Mixed meal tolerance test					
0-30min					
tAUC glucose	5.2 (1.0)	5.1 (1.4)	4.8 (1.0)	4.9 (1.0)	5.0 (1.1)
tAUC C-peptide	838 (232)	880 (258)	868 (278)	888 (197)	869 (239)
tAUC insulin, median (IQR)	138 (110; 196)	171 (97; 210)	130 (84; 201)	177 (118; 209)	154 (92; 204)
tAUC GLP-1 total, median (IQR)	8 (7; 12)	8 (7; 11)	8 (7; 11)	9 (7; 11)	8 (7; 11)
tAUC GIP total	29 (10)	29 (9)	30 (12)	27 (10)	29 (10)
tAUC paracetamol, median (IQR)	0.0 (0.0; 0.0)	0.0 (0.0; 0.0)	0.0 (0.0; 0.0)	0.0 (0.0; 0.0)	0.0 (0.0; 0.0)
0-120 min					
Oral DI (Matsuda × IGI), median (IQR)	391.9 (201.3)	273.5 (148.5)	338.5 (177.7)	389.3 (231.1)	340.0 (178.2)
Oral DI (Matsuda × AUC insulin/glucose), median (IQR)	149.8 (88.1)	193.8 (93.7)	168.6 (109.6)	182.3 (99.1)	168.6 (94.3)
AUC (insulin/glucose), median (IQR)	36.7 (27.4; 71.1)	43.1 (34.6; 63.3)	60.0 (23.1; 71.3)	42.9 (36.7; 85.6)	43.0 (28.2; 73.6)
Oral ISI, median (IQR)	3.3 (2.9; 5.2)	2.6 (2.2; 5.0)	3.8 (2.3; 5.4)	3.7 (2.8; 4.6)	3.6 (2.5; 5.0)
tAUC glucose	27.9 (5.3)	28.6 (7.0)	26.1 (6.1)	26.2 (6.3)	27.2 (6.2)
tAUC C-peptide	5,607 (1,611)	6,206 (2,292)	5,636 (1,544)	6,371 (2,309)	5,963 (1,974)
tAUC insulin, median (IQR)	1,081 (872; 1,584)	1,374 (921; 1,699)	1,221 (677; 1,576)	1,155 (935; 1,987)	1,167 (862; 1,646)
tAUC GLP-1 total, median (IQR)	40 (32; 52)	41 (37; 45)	39 (33; 46)	42 (33; 48)	41 (33; 46)
tAUC GIP total, median (IQR)	156 (122; 186)	157 (137; 186)	148 (114; 184)	146 (125; 189)	150 (123; 186)
tAUC paracetamol, median (IQR)	0.1 (0.1; 0.1)	0.1 (0.1; 0.1)	0.1 (0.1; 0.1)	0.1 (0.1; 0.1)	0.1 (0.1; 0.1)

Data are presented as means and standard deviations or medians with IQR (25<sup>th</sup> to 75<sup>th</sup> percentile). Units for AUC glucose and paracetamol are presented in mmol/l×h, and units for AUC insulin, C-peptide, GLP-1 and GIP are presented in pmol/l×h. SGLT2i, sodium/glucose cotransporter 2 inhibitor; DPP4i, dipeptidyl peptidase 4 inhibitors; ARB, angiotensin ll receptor blocker; ACEi, angiotensin-converting enzyme inhibitor; CCB, calcium channel blocker; IGI, insulinogenic. \*n=20; \*\*n=19; \*\*\*n=16.

HED

HED

HED

MED

MED



**Fig. 2** | **Changes from constrained baseline to 16-week follow-up in the primary and secondary outcomes. a**–**f**, The bars represent estimated mean change from baseline for each intervention group from the *n* = 82 persons included in the study. Error bars represent 95% confidence intervals. Data were log(e)-transformed and back-transformed, and the results are presented as relative (percentage term) changes based on the ratio of geometric mean change from baseline to follow-up. Results were adjusted for sex. Data were analyzed

fasting C-peptide, fasting triglycerides and systolic blood pressure (Tables 4 and 5). All intervention groups had reduced diastolic blood pressure, and the reduction was greater in HED and MED than in DCON. No reductions in low-density lipoprotein cholesterol (LDL-C) were observed. Physical fitness defined as maximum oxygen consumption in ml O<sub>2</sub>/min (VO<sub>2max</sub>) increased in MED and HED compared with CON and DCON, and HED improved more than MED. VO<sub>2max</sub> relative to body weight defined as ml O<sub>2</sub>/min/kg (relative VO<sub>2max</sub>) changed by -3%, 8%, 23% and 39% in CON, DCON, MED and HED, respectively (Table 4). HED improved absolute and relative-to-body-weight 1 repetition maximum (RM) chest press compared with all of the other groups, whereas 1 RM

using a constrained baseline longitudinal model. The dots represent the relative (percentage term) individual changes from baseline to follow-up. Left panel: Data are based on the final 30 min of the hyperglycemic clamp (stage 1). Right panel: Data are from 0 to 120 min of the MMTT. **a**, Change in late-phase DI by group. **b**, Change in late-phase ISI by group. **c**, Change in late-phase ISR by group. **d**, Change in oral DI of the MMTT by group. **e**, Change in oral ISI by group. **f**, Change in oral ISR by group. CON, *n* = 20; DCON, *n* = 21; MED, *n* = 20; HED, *n* = 21.

MED

leg extensions relative to body weight improved in HED only when compared with CON and DCON (Table 5).

#### Sensitivity analyses

The multiple imputation analyses on the primary and secondary outcomes agreed with the primary analyses (Supplementary Table 13).

#### Post hoc analyses

As a post hoc outcome, the need for medication, after completion of follow-up testing, was calculated based on the prespecified algorithm. Reductions of glucose-lowering medication were 11%, 92%, 81% and 89%

#### Table 2 | Pairwise comparisons of the change in the primary outcome and secondary outcomes

	HED vs. CON		MED vs. CON		DCON vs. CON		HED vs. DCON		MED vs. DCO	N	HED vs. MED		Global P
	MD (95% CI)	Р	MD (95% CI)	Р	MD (95% CI)	Р	MD (95% CI)	Р	MD (95% CI)	Р	MD (95% CI)	Р	
Primary outcome													
Late-phase DI (ITT)	137 (73; 225)	<sup>&lt;</sup> 0.001	105 (49; 182)	<sup>&lt;</sup> 0.001	58 (16; 116)	0.004	50 (10; 104)	0.01	29 (-5; 77)	0.11	16 (-16; 59)	0.34	<sup>&lt;</sup> 0.001
Late-phase DI (PP)	126 (62; 214)	<0.001	115 (54; 202)	<0.001	66 (21; 128)	0,002	36 (-1; 86)	0.058	30 (-7; 79)	0.12	5 (-25; 47)	0.79	<sup>&lt;</sup> 0.001
Secondary outcome													
ISI	83 (35; 149)	<sup>&lt;</sup> 0.001	50 (10; 104)	0.011	18 (–13; 60)	0.28	55 (15; 109)	0.004	27 (-7; 71)	0.13	23 (-10; 66)	0.20	<sup>&lt;</sup> 0.001
ISR	28 (13; 45)	<sup>&lt;</sup> 0.001	38 (22; 57)	<sup>&lt;</sup> 0.001	33 (18; 50)	<sup>&lt;</sup> 0.001	-4 (-15; 9)	0.56	4 (-8; 18)	0.51	-8 (-18; 5)	0.22	<sup>&lt;</sup> 0.001
Oral DI	141 (80; 223)	<sup>&lt;</sup> 0.001	133 (73; 213)	<sup>&lt;</sup> 0.001	87 (40; 148)	<sup>&lt;</sup> 0.001	29 (-2; 70)	0.065	25 (-5; 65)	0.12	4 (-22; 37)	0.81	<sup>&lt;</sup> 0.001
Oral ISI	127 (78; 188)	<sup>&lt;</sup> 0.001	110 (65; 168)	<sup>&lt;</sup> 0.001	75 (38; 122)	<sup>&lt;</sup> 0.001	29 (3; 62)	0.025	20 (-4; 51)	0.12	8 (–15; 36)	0.53	<sup>&lt;</sup> 0.001
Oral ISR	5 (-16; 32)		13 (-10; 42)		7 (-14; 33)		-2 (-20; 22)		6 (-14; 31)		-7 (-25; 16)		0.76

Data were log(e)-transformed and back-transformed, and the results are presented as relative changes based on the ratio of estimated geometric mean (95% CI) change from baseline in one group versus the other. Results are adjusted for sex. Data were analyzed using a constrained baseline longitudinal model. *P* values are two-sided. No corrections for multiple comparisons were performed. If Global *P* is >0.1, *P* values are not reported for between-group comparisons. MD, mean difference.

#### Table 3 | Adverse events after randomization

	CON, n (%)	DCON, n (%)	MED, n (%)	HED, n (%)	All, n (%)
Participants with $\ge 1$ SAE or AE <sup>a</sup>	2 (10.0)	3 (14.3)	5 (25.0)	5 (23.8)	15 (18.3)
SAE	2 (10.0)	1 (4.8)	0 (0.0)	0 (0.0)	3 (3.7)
Infections (COVID-19)	0 (0.0)	0 (0.0)	2 (10.0)	0 (0.0)	2 (2.4)
Musculoskeletal pain and discomfort					
Back pain	0 (0.0)	0 (0.0)	0 (0.0)	1 (4.8)	1 (1.2)
Lower extremities	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)
Upper extremities	0 (0.0)	0 (0.0)	0 (0.0)	1 (4.8)	1 (1.2)
Other	0 (0.0)	0 (0.0)	0 (0.0)	2 (9.5)	2 (2.4)
Musculoskeletal injury					
Back pain	0 (0.0)	0 (0.0)	0 (0.0)	1 (4.8)	1 (1.2)
Lower extremities	0 (0.0)	0 (0.0)	1 (5.0)	1 (4.8)	2 (2.4)
Fatigue	0 (0.0)	0 (0.0)	0 (0.0)	1 (4.8)	1 (1.2)
Other	0 (0.0)	0 (0.0)	0 (0.0)	1 (4.8)	1 (1.2)
Complications associated with clinical or experimental procedures					
Allergic reactions to bandages and wound plasters	0 (0.0)	0 (0.0)	1 (5.0)	2 (9.5)	3 (3.7)
Felt uncomfortable during hyperglycemic clamp	0 (0.0)	1 (4.8)	1 (5.0)	0 (0.0)	2 (2.4)
Pain from muscle biopsy	0 (0.0)	1 (4.8)	1 (5.0)	0 (0.0)	2 (2.4)
Peripheral intravenous catheter went subcutaneous during hyperglycemic clamp	0 (0.0)	1 (4.8)	0 (0.0)	0 (0.0)	1 (1.2)
Nutrition	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)

<sup>a</sup>(%)The numerator is the number of participants with at least one adverse event in each group, and the denominator is the total number of participants in the group. SAE, serious adverse event; AE, adverse event. CON, *n*=20; DCON, *n*=21; MED, *n*=20; HED, *n*=21. SAEs included one case of transient ischemic attack and one case of malignant melanoma in the CON group, and one case of prolactinoma in the DCON group. Musculoskeletal pain and discomfort allows for modification of an exercise so that the prescribed exercise intervention can be performed. Musculoskeletal injury is defined as pain or discomfort to an extent that precludes participating in at least one protocol-prescribed exercise for >7 days. Nutrition is defined as events of increased or decreased hunger or satiety.

in CON, DCON, MED and HED, respectively (Table 4). The corresponding numbers for discontinuations of glucose-lowering medication were 28%, 39%, 69% and 83%, respectively (Table 4). The odds of reductions and discontinuations were higher in all intervention groups than in CON (P < 0.05). No differences were observed in the odds of reductions of glucose-lowering medication between the intervention groups, but the odds of discontinuations were higher for HED than for CON (odds ratio (OR), 2.7 (95% CI, 1.1; 7.9); Table 5). Although the odds of discontinuation were higher in MED than in DCON (OR, 3.4 (95% CI, 0.6; 21.6)), it did not reach statistical significance (P = 0.2; Table 5). The odds of discontinuations were similar between HED and MED, and no differences were observed in any group comparison for other medications (Table 5). The role of weight loss on the primary and secondary outcomes was explored in a post hoc statistical mediation analysis (Supplementary Table 16). It revealed that the treatment effect mediated by weight loss on late-phase DI was similar across the exercising groups and accounted for around 50–60% of the total effect, whereas 70% of the treatment effect was mediated by weight loss in DCON. Regarding late-phase ISI, the pattern was similar for the exercising groups, but for DCON, the weight loss was entirely responsible for the treatment effect. Weight loss did not explain the increase in late-phase ISR.

#### Discussion

One of our main findings is that all intervention groups improved beta-cell function, as expressed by late-phase DI, more than



Fig. 3 | Insulin secretion rate across a three-phase hyperglycemic clamp. Data are represented as the marginal means with 95% confidence intervals. Results are adjusted for sex. Data were analyzed using a longitudinal mixed model. The black dashed line represents baseline values, and the red solid line represents the 16-week follow-up values.

standard care. Furthermore, adding exercise to diet-induced weight loss improved beta-cell function more than diet-induced weight loss or standard care alone. This seemed to be achieved by additional increases in insulin sensitivity induced by exercise in a dose-dependent manner. However, the secondary and exploratory outcomes did not uniformly support the linear dose-response relationship observed for the primary outcome.

There is a paucity of studies investigating the role of exercise and exercise volume in conjunction with diet-induced weight loss, but our observations are in line with previous findings suggesting that high volumes of exercise without a concomitant dietary intervention improve first-phase and/or late-phase DI in people with prediabetes and T2D<sup>5,12,23</sup>. In line with other studies, our data support that the exercise component increases DI due to increases in insulin sensitivity rather than increased insulin secretion<sup>5,12,23,24</sup>. In contrast, other studies have shown that exercise may increase insulin secretion and not insulin sensitivity in people with dysglycemia<sup>6,8</sup>. One reason for the discrepancy may be the higher glucose clamp levels (25 mmol/l) used in ref. 6 compared with the -13 mmol/l plasma glucose used in our study. At clamp levels closer to our target (-13 mmol/l plasma glucose), the researchers in ref. 6 also did not observe an increase in insulin secretion. The differences could also relate to the exercise intensities in the studies wherein, for a given increase in insulin sensitivity, it has been shown that high-intensity exercise results in a larger reduction in insulin secretion than low-intensity to moderate-intensity exercise<sup>12</sup>.

Although we did not observe a difference in late-phase ISR with increased exercise volume compared with diet-induced weight loss alone, all intervention groups exhibited similar increases in late-phase ISR. Therefore, it could be speculated that diet-induced weight loss alone might explain this observation and that a weight loss of ~7.5% body weight may be sufficient to re-establish late-phase ISR in this study population. Supporting this, a previous study from our group, using the same intervention protocol as for HED, also increased DI. Consistent with our current findings, only the improvement in insulin sensitivity index, and not the small increase in insulin secretion, was associated with exercise volume in that study<sup>10</sup>. Taken together, these findings suggest that changes in insulin sensitivity are more exercise driven, whereas changes in insulin secretion are primarily driven by weight loss. In further support of this, previous studies have shown an increased insulin secretory capacity after caloric restriction with no or marginal improvement in peripheral insulin sensitivity; thus, first-phase, late-phase and total-phase DI are explained mainly by increases in insulin secretion<sup>3,18,25</sup>. The increased first-phase ISR following diet-induced weight loss is consistent with findings of other studies<sup>3,26</sup> and may relate to a decrease in fasting plasma glucose (which was comparable between the intervention groups in our study)<sup>27</sup>. However, it was surprising that exercise was associated with an attenuation of the increase in first-phase ISR induced by diet-induced weight loss. When first-phase DI was calculated (correcting the ISR for insulin sensitivity), the increase was larger in the exercising groups than with diet-induced weight loss alone. Although there seems to be a consensus that regaining first-phase insulin secretion is characteristic of T2D remission, we found a reduction when exercise was added to a diet-induced weight loss intervention. An explanation may relate to both improved insulin sensitivity and glucose effectiveness observed with exercise<sup>5,16</sup>, or could be ascribed to a blunted insulin secretion during hyperglycemia, GIP stimulation and arginine stimulation after high volumes of exercise<sup>28-30</sup>. The HED group may have experienced an exercise-induced blunting of insulin secretion whereas MED did not, which could also explain why there were no further increases in ISR, first-phase DI, GLP-1 stimulation and arginine stimulation despite the largest late-phase ISI being in HED. The metabolic consequences of a slightly blunted insulin secretion in people with T2D are unknown. Nevertheless, exercise-induced decreases in insulin secretion during concomitant increases in insulin sensitivity are consistently found in individuals with prediabetes and/ or obesity, as well as in healthy people<sup>12,23,28</sup>. Given that insulin secretion physiologically counterbalances insulin sensitivity as a homeostatic response<sup>15,31</sup>, an exercise-induced increase in insulin sensitivity added to diet-induced weight loss may reduce the demand on beta cells, offering beta-cell rest and therefore preserving beta-cell health.

In this study, both late-phase DI and late-phase ISI increased linearly with increasing treatment intensity. However, a similar relationship was not observed during the MMTT nor in first-phase DI or the GLP-1 and arginine stimulations. Although speculative, this may relate to the route of glucose administration. Recently, it was described that

#### Table 4 | Within-group changes (0-16 weeks) in cardiometabolic outcomes

Change (95% Cl)Change (95% Cl)Change (95% Cl)Change (95% Cl)Change (95% Cl)Glycemic controlHbA1c, mmol/mol2 (0; 4)-5 (-6; -3)-5 (-7; -3)-5 (-7; -3)HbA1c, %0.1 (0.1; 0.1)0.1 (-1.1; 0.1)0.1 (-1.1; 0.1)0.1 (-1.1; 0.1)Fasting glucose, mmol/l0.1 (0.1; 1.1)-1.1 (-2.1; -1.1)-2.1 (-3.1; -1.1)-2.1 (-3.1; -1.1)Fasting insulin, pmol/l1 (-23; 25)-46 (-67; -26)-60 (-82; -38)-64 (-86; -43)Fasting C-peptide, pmol/l35 (-83; 153)-216 (-319; -112)-321 (-432; -209)-363 (-472; -254)
Glycemic control           HbA1c, mmol/mol         2 (0; 4)         -5 (-6; -3)         -5 (-7; -3)         -5 (-7; -3)           HbA1c, %         0.1 (0.1; 0.1)         0.1 (-1.1; 0.1)         0.1 (-1.1; 0.1)         0.1 (-1.1; 0.1)           Fasting glucose, mmol/l         0.1 (0.1; 1.1)         -1.1 (-2.1; -1.1)         -2.1 (-3.1; -1.1)         -2.1 (-3.1; -1.1)           Fasting insulin, pmol/l         1 (-23; 25)         -46 (-67; -26)         -60 (-82; -38)         -64 (-86; -43)           Fasting C-peptide, pmol/l         35 (-83; 153)         -216 (-319; -112)         -321 (-432; -209)         -363 (-472; -254)
HbA1c, mmol/mol         2 (0; 4)         -5 (-6; -3)         -5 (-7; -3)         -5 (-7; -3)           HbA1c, %         0.1 (0.1; 0.1)         0.1 (-1.1; 0.1)         0.1 (-1.1; 0.1)         0.1 (-1.1; 0.1)           Fasting glucose, mmol/l         0.1 (0.1; 1.1)         -1.1 (-2.1; -1.1)         -2.1 (-3.1; -1.1)         -2.1 (-3.1; -1.1)           Fasting insulin, pmol/l         1 (-23; 25)         -46 (-67; -26)         -60 (-82; -38)         -64 (-86; -43)           Fasting C-peptide, pmol/l         35 (-83; 153)         -216 (-319; -112)         -321 (-432; -209)         -363 (-472; -254)
HbA1c, %         0.1 (0.1; 0.1)         0.1 (-1.1; 0.1)         0.1 (-1.1; 0.1)         0.1 (-1.1; 0.1)           Fasting glucose, mmol/l         0.1 (0.1; 1.1)         -1.1 (-2.1; -1.1)         -2.1 (-3.1; -1.1)         -2.1 (-3.1; -1.1)           Fasting insulin, pmol/l         1 (-23; 25)         -46 (-67; -26)         -60 (-82; -38)         -64 (-86; -43)           Fasting C-peptide, pmol/l         35 (-83; 153)         -216 (-319; -112)         -321 (-432; -209)         -363 (-472; -254)
Fasting glucose, mmol/l         0.1 (0.1; 1.1)         -1.1 (-2.1; -1.1)         -2.1 (-3.1; -1.1)         -2.1 (-3.1; -1.1)           Fasting insulin, pmol/l         1 (-23; 25)         -46 (-67; -26)         -60 (-82; -38)         -64 (-86; -43)           Fasting C-peptide, pmol/l         35 (-83; 153)         -216 (-319; -112)         -321 (-432; -209)         -363 (-472; -254)
Fasting insulin, pmol/l         1 (-23; 25)         -46 (-67; -26)         -60 (-82; -38)         -64 (-86; -43)           Fasting C-peptide, pmol/l         35 (-83; 153)         -216 (-319; -112)         -321 (-432; -209)         -363 (-472; -254)
Fasting C-peptide, pmol/l         35 (-83; 153)         -216 (-319; -112)         -321 (-432; -209)         -363 (-472; -254)
Glucose-lowering medication
Reduction, no (%), n=65 <sup>a</sup> 2.0 (11.1; 0.0)         12.0 (92.3; 0.0)         13.0 (81.3; 0.0)         16.0 (88.9; 0.0)
Discontinuation, no (%), n=65 <sup>a</sup> 5.0 (27.8; 0.0)         5.0 (38.5; 0.0)         11.0 (68.8; 0.0)         15.0 (83.3; 0.0)
Intensification, no (%), n=82 <sup>b</sup> 9.0 (45.0; 0.0)         0.0 (0.0; 0.0)         2.0 (10.0; 0.0)         2.0 (9.5; 0.0)
Lipid-lowering medication
Intensification, no (%), n=82 <sup>b</sup> 5.0 (25.0; 0.0)         1.0 (4.8; 0.0)         1.0 (5.0; 0.0)         0.0 (0.0; 0.0)
Blood pressure-lowering medication
Reduction, no (%), n=47 <sup>a</sup> 1.0 (9.1; 0.0)         3.0 (27.3; 0.0)         1.0 (10.0; 0.0)         2.0 (13.3; 0.0)
Discontinuation, no (%), n=47 <sup>a</sup> 0.0 (0.0; 0.0)         1.0 (9.1; 0.0)         0.0 (0.0; 0.0)         0.0 (0.0; 0.0)
Intensification, no (%), n=82 <sup>b</sup> 1.0 (5.0; 0.0)         1.0 (4.8; 0.0)         0.0 (0.0; 0.0)         1.0 (4.8; 0.0)
Lipids
LDL cholesterol, mmol/l -1(-1; 0) -1(-1; 0) -1(-1; 0)
Fasting triglycerides, % change from baseline         -17 (-28; -6)         -41 (-48; -33)         -37 (-45; -28)         -38 (-46; -30)
Blood pressure
Systolic, mmHg -3 (-6; 0) -7 (-9; -4) -9 (-12; -6) -7 (-10; -5)
Diastolic, mmHg         -4 (-5; -2)         -4 (-6; -3)         -7 (-8; -5)         -7 (-8; -5)
Physical fitness
Absolute VO <sub>2max</sub> , ml/min, n=78         -109.1 (-248.7; 30.7)         -25.1 (-138.1; 87.9)         209.7 (81.3; 338.0)         540.9 (411.6; 670.1)
Relative VO <sub>2max</sub> , ml/kg/min, n=78         -0.8 (-2.3; 0.7)         2.0 (0.8; 3.2)         5.6 (4.2; 6.9)         9.5 (8.2; 10.9)
Watt max, W/kg, n=78         -0.1 (-0.2; 0.1)         0.2 (0.1; 0.3)         0.7 (0.5; 0.8)         0.9 (0.8; 1.1)
1 RM chest press, kg, n=79       0.2 (-3.4; 3.9)       -1.5 (-4.4; 1.4)       -1.7 (-4.6; 1.2)       5.5 (2.7; 8.4)
1 RM chest press, kg/kg body weight, n=79         0.0 (0.0; 0.1)         0.0 (0.0; 0.1)         0.0 (0.0; 0.1)         0.1 (0.1; 0.2)
1 RM leg extension, kg, n=81       4.2 (0.1; 8.3)       -1.2 (-4.8; 2.3)       2.1 (-1.6; 5.9)       2.1 (-1.6; 5.7)
1 RM leg extension, kg/kg body weight, n=81         0.1 (0.0; 0.1)         0.1 (0.0; 0.1)         0.1 (0.1; 0.1)         0.1 (0.1; 0.2)
Body anthropometrics
Body weight, kg         -0.3 (-2.6; 2.0)         -7.4 (-9.5; -5.3)         -10.6 (-12.8; -8.3)         -11.9 (-14.1; -9.7)
BMI, kg/m <sup>2</sup> -0.1 (-0.8; 0.6) -2.4 (-3.1; -1.7) -3.4 (-4.1; -2.7) -3.8 (-4.5; -3.1)

Data are estimated means with 95% confidence intervals. Data were analyzed using a constrained baseline longitudinal model (two-sided). No corrections for multiple comparisons were performed. Reduction is defined as at least one step down on the predefined algorithm. Discontinuation is defined as discontinuation of all drugs when therapeutic target was met. Intensification is defined as at least one step up on the predefined algorithm. <sup>a</sup>Participants on medication at baseline (denominator). <sup>b</sup>All participants (denominator).

the increased insulin secretion observed after oral administration of glucose compared with intravenous administration might be accompanied by a compensatory decrease in insulin sensitivity<sup>32</sup>. Furthermore, the incretin response during the MMTT might activate a GIP-induced vasoconstriction in the microvasculature of skeletal muscles<sup>33</sup>, which could dampen a difference in insulin action between the exercise groups achieved by exercise-induced skeletal muscle capillarization<sup>34</sup>.

These findings suggest that there is only a limited effect of increasing exercise volume from three to six sessions weekly in the context of diet-induced weight loss. Likewise, although weight loss was larger in both exercise groups compared with diet-induced weight loss alone, there was no apparent additional weight loss when doubling the exercise volume from three to six sessions per week. In contrast, the increase in both absolute and relative  $VO_{2max}$  was positively associated with exercise dose, with only marginal differences in maximal strength. Although a weight loss of  $\geq 5\%$  may increase beta-cell function slightly, a weight loss of  $\geq 11\%$  may be necessary to maximize an increase in peripheral insulin sensitivity in people with obesity<sup>2</sup>. However, diet-induced weight losses of  $\geq 15$  kg resulting in significant improvements in beta-cell function have been reported in people with T2D without concomitant increases in peripheral insulin sensitivity<sup>3</sup>.

Still, diet-induced weight loss may primarily improve hepatic (central) insulin sensitivity and beta-cell insulin secretory capacity through reductions in visceral and ectopic fat (that is, in liver and pancreas) that confer a dose-dependent increase in beta-cell function<sup>3,4</sup>. This might explain why the post hoc statistical mediation analysis suggested that the role of weight loss on the intervention effect mediated the entire effect of late-phase ISI in DCON. In contrast, exercise mainly improves peripheral insulin sensitivity<sup>35</sup> and may explain why only 50–60% of the intervention effect was mediated by weight loss on late-phase

			MED ve CON <sup>a</sup>		DCON VE CON <sup>a</sup>		HED ve DCON <sup>a</sup>				HED VE MED <sup>a</sup>		Global D
	MD (95% CI)	٩	MD (95% CI)	٩	MD (95% CI)	٩	MD (95% CI)	٩	MD (95% CI)	٩	MD (95% CI)	٩	
Glycemic control													
HbA1c, mmol/mol	-7 (-9; -5)	<0.001	-7 (-9; -5)	<0.001	-7 (-9; -5)	<0.001	0 (-3; 2)	0.75	0 (-2; 2)	0.93	0 (-2; 2)	0.82	<0.001
HbA1c, %	-1.1 (-1.1; 0.1)	<0.001	-1.1 (-1.1; 0.1)	<0.001	-1.1 (-1.1; 0.1)	<0.001	0.1 (0.1; 0.1)	0.76	0.1 (0.1; 0.1)	0.97	0.1 (0.1; 0.1)	0.79	<0.001
Fasting glucose, mmol/l	-2.1 (-3.1; -1.1)	<0.001	-2.1 (-3.1; -1.1)	<0.001	-2.1 (-3.1; -1.1)	<0.001	0.1 (-1.1; 0.1)	0.30	0.1 (-1.1; 0.1)	0.31	0.1 (-1.1; 1.1)	1.00	<0.001
Fasting insulin, pmol/l	-65 (-97; -34)	<0.001	-61 (-93; -29)	<0.001	-47 (-78; -17)	0.003	-18 (-47; 11)	0.23	-13 (-43; 16)	0.38	-5 (-35; 26)	0.77	<0.001
Fasting C-peptide, pmol/l	-398 (-555; -240)	<0.001	-356 (-515; -196)	<0.001	-250 (-404; -96)	0.001	-147 (-295; 0)	0.050	-105 (-255; 44)	0.17	-42 (-195; 111)	0.59	<0.001
Glucose-lowering medication													
Reduction, OR	3.7 (1.9; 9.4)	<0.001	5.4 (2.0; 20.1)	<0.001	71.5 (6.2; 4341.4)	<0.001	0.8 (0.1; 3.8)	1.000	0.4 (0.0; 5.4)	0.77	1.8 (0.2; 24.8)	0.88	<0.001
Discontinuation, OR	2.3 (1.3; 4.5)	0.002	2.3 (1.0; 5.7)	0.039	1.6 (0.3; 9.6)	0.81	2.7 (1.1; 7.9)	0.028	3.4 (0.6; 21.6)	0.21	2.2 (0.3; 17.4)	0.55	0.003
Intensification, OR	0.5 (0.2; 0.9)	0.025	0.4 (0.1; 0.9)	0.031	0.1 (0.0; 0.34 <sup>b</sup> )	0.001	1.6 (0.4; ∞ <sup>b</sup> )	0.49	2.6 (0.2; ∞ <sup>b</sup> )	0.46	1.0 (0.1; 14.4)	1.000	0.001
Lipid-lowering medication													
Intensification, OR	0.5 (0.0; 0.98 <sup>b</sup> )	0.041	0.4 (0.1; 1.3)	0.18	0.2 (0.0; 1.6)	0.16	1.0 (0.0; 6.24 <sup>b</sup> )	1.00	1.1 (0.0; 86.7)	1.00	1.0 (0.0; 37.14 <sup>b</sup> )	0.98	0.031
Blood pressure-lowering medication													
Reduction, OR	1.2 (0.4; 4.6)		1.1 (0.1; 9.8)		3.5 (0.2; 216.2)		0.7 (0.2; 2.1)		1.1 (0.1; 9.8)		1.4 (0.1; 90.6)		0.70
Discontinuation, OR	NE (0.0; 0.0)	NE	NE	NE	NE	NE	NE	NE	NE	NE	NE	NE	NE
Intensification, OR	1.0 (0.2; 4.3)		1.0 (0.0; 6.24 <sup>b</sup> )		1.0 (0.0; 78.5)		1.0 (0.1; 9.1)		1.1 (0.0; 40.95 <sup>b</sup> )		1.0 (0.0; ∞ <sup>b</sup> )		1.00
Lipids													
LDL cholesterol, mmol/l	0 (0; 0)		0 (0; 0)		0 (0; 0)		0 (0; 0)		0 (0; 0)		0 (0; 0)		0.99
Fasting triglycerides, % change from baseline	-25 (-37; -12)	0.001	-24 (-35; -10)	0.002	-29 (-39; -16)	<0.001	5 (-11; 23)	0.41	7 (-9; 25)	0.57	-2 (-17; 15)	0.80	<0.001
Blood pressure													
Systolic, mmHg	-4 (-7; -1)	0.021	-6 (-9; -2)	0.001	-4 (-7; 0)	0.039	0 (-4; 3)	0.79	-2 (-6; 1)	0.18	2 (-2; 5)	0.29	0.010
Diastolic, mmHg	-3 (-5; -1)	0.011	-3 (-5; -1)	0.011	-1 (-3; 1)	0.46	-2 (-4; 0)	0.063	-2 (-4; 0)	0.059	0 (-2; 2)	0.95	0.017
Physical fitness													
Absolute VO <sub>2max</sub> , ml/min, $n=78$	649.9 (461.2; 838.7)	<0.001	318.7 (130.7; 506.7)	0.001	84.0 (-94.3; 262.2)	0.36	566.0 (396.0; 736.0)	<0.001	234.7 (65.6; 403.9)	0.007	331.2 (151.0; 511.4)	<0.001	<0.001
Relative VO <sub>2max</sub> , ml/kg/min, <i>n</i> =78	10.3 (8.3; 12.3)	<0.001	6.4 (4.4; 8.4)	<0.001	2.8 (0.9; 4.7)	0.004	7.5 (5.7; 9.3)	<0.001	3.6 (1.8; 5.3)	<0.001	4.0 (2.1; 5.9)	<0.001	<0.001
Watt max, W/kg, $n = 78$	1.0 (0.8; 1.2)	<0.001	0.7 (0.5; 0.9)	<0.001	0.2 (0.1; 0.4)	0.008	0.8 (0.6; 0.9)	<0.001	0.5 (0.3; 0.6)	<0.001	0.3 (0.1; 0.5)	0.002	<0.001
1 RM chest press, kg, $n=79$	5.3 (0.7; 9.9)	0.025	-2.0 (-6.6; 2.7)	0.41	-1.7 (-6.4; 2.9)	0.46	7.0 (3.0; 11.1)	0.001	-0.2 (-4.3; 3.8)	0.91	7.3 (3.2; 11.3)	<0.001	0.001
1 RM chest press, kg/kg body weight, $n=79$	0.1 (0.1; 0.2)	<0.001	0.0 (0.0; 0.1)	0.31	0.0 (0.0; 0.1)	0.53	0.1 (0.1; 0.2)	<0.001	0.0 (0.0; 0.1)	0.66	0.1 (0.1; 0.1)	<0.001	<0.001
1 RM leg extension, kg, $n=81$	-2.1 (-7.6; 3.3)		-2.1 (-7.6; 3.4)		-5.5 (-10.9; -0.1)		3.3 (-1.7; 8.4)		3.4 (-1.8; 8.5)		0.0 (-5.2; 5.2)		0.241
1 RM leg extension, kg/kg body weight, $n=81$	0.1 (0.0; 0.1)	0.028	0.0 (0.0; 0.1)	0.12	0.0 (-0.1; 0.1)	0.91	0.1 (0.0; 0.1)	0.013	0.1 (0.0; 0.1)	0.074	0.0 (0.0; 0.1)	0.52	0.035
Body anthropometrics													
Body weight, kg	-11.6 (-14.8; -8.4)	<0.001	-10.3 (-13.5; -7.1)	<0.001	-7.1 (-10.2; -4.0)	<0.001	-4.5 (-7.5; -1.4)	0.004	-3.2 (-6.2; -0.1)	0.043	-1.3 (-4.5; 1.8)	0.40	<0.001
BMI, kg/m <sup>2</sup>	-3.7 (-4.7; -2.7)	<0.001	-3.3 (-4.3; -2.3)	<0.001	-2.3 (-3.3; -1.3)	<0.001	-1.4 (-2.4; -0.5)	0.004	-1.0 (-1.9; 0.0)	0.044	-0.4 (-1.4; 0.5)	0.38	<0.001
Data are estimated means with 95% confidence inte Continuous data were analyzed using a constrained in medication changes. "Reference category for the	ervals unless stated othe d baseline longitudinal n e looistic regression anal	erwise. Red nodel, and lyses. Wher	uction, Discontinuation ordinal variables were orzero-event data were	n and Inter analyzed u observed	nsification are defined using exact logistic rec a continuity correctio	as in Table gression. Pv n was used	<ol> <li>NE, not estimable alues are two-sided that was inversely r</li> </ol>	. If Global I No correc	<sup>2</sup> is >0.1, <i>P</i> values are tions for multiple cc 1 to the relative size.	mot repoi	rted for between-gro ns were performed b posite aroup	oup compa oecause of	ırisons. sparse data

Table 5 | Pairwise comparisons of the change in cardiometabolic outcomes

Nature Metabolism | Volume 5 | May 2023 | 880-895

ISI when exercise was added to the diet. As such, the small additional weight loss observed in the exercise groups compared with DCON most likely does not alone explain the add-on effect on late-phase ISI. These findings support that exercise may improve beta-cell function by increasing peripheral insulin sensitivity beyond the effects of weight reduction alone<sup>34–36</sup>.

Interestingly, weight loss completely mediated the intervention effects in the oral DI and oral ISI, suggesting that weight loss becomes the most important signal in the context of an oral mixed meal and normal homeostatic postprandial regulation. Although speculative, this may relate not only to the complex neuronal and endocrine organ crosstalk, but also to the meal composition wherein certain amino acids and fatty acids regulate insulin secretion, as well as skeletal muscle microvascular blood flow.

#### Limitations

Our findings must be interpreted in the context of the limitations of the study. First, the sample size was based on a previous study including up to five aerobic exercise sessions per week. Thus, the lack of differences between MED and HED or DCON and MED could be a type 2 error due to low statistical power. However, given the consistent signal across most beta-cell indices, the results can be interpreted with confidence. Second, we assessed beta-cell DI with a hyperglycemic clamp. Although the hyperglycemic clamp is the gold standard for beta-cell function<sup>15</sup>, it is an unphysiological assessment that limits physiological translation. However, we clamped the glucose level at only 5.4 mmol/l above the fasting glucose level, attempting to mimic postprandial glucose levels. Furthermore, we assessed beta-cell indices during an MMTT to compare the supraphysiological hyperglycemic clamp (glucose levels ~13 mmol/l at baseline and follow-up) to the physiological conditions during the MMTT (peak glucose levels ~16 mmol/l during MMTT at baseline). Because we observed a consistent pattern between the hyperglycemic clamp and MMTT, this allows for translating the findings from the hyperglycemic clamp to a physiological context. Third, we applied pharmacological constraints, and the participants were all relatively newly diagnosed and pharmacologically well regulated before randomization as well as throughout the study. Therefore, we cannot directly translate the results to people with longer T2D duration, treated with other pharmacological agents, or who have poor glycemic control. However, as our findings are consistent with pharmacological weight-loss trials<sup>37</sup>, they may still have clinical implications. Fourth, the intervention was only 16 weeks. Diminished benefits concerning beta-cell indices when going from three to six exercise sessions per week could be due to ceiling effects for the time course of the intervention. Hence, three exercise sessions per week combined with a 25% energy deficit may almost fully saturate the rate of adaptation for the mechanisms influencing beta-cell function. Furthermore, organ-specific changes in response to chronic exercise may occur on different time courses and will also reflect individual responses to exercise<sup>35,38</sup>. Thus, 16 weeks may have been too short to see significant deviations between MED and HED or even from DCON. Fifth, we did not use the gold standard hyperinsulinemic-euglycemic clamp to assess insulin sensitivity; however, the hyperglycemic clamp provides a reliable measure of glucose disposal<sup>15,39</sup>. Moreover, we observed that late-phase EGP did not change while  $R_d$  increased compared with CON, suggesting an increased peripheral insulin sensitivity and not hepatic insulin sensitivity. Sixth, we assessed dietary adherence using self-reporting, which may include information bias<sup>40</sup>. Seventh, although DI is considered the most accurate assessment of beta-cell function<sup>15</sup>, the relationship between insulin secretion and insulin sensitivity is not consistently hyperbolic across levels of glucose tolerance, BMI or measurements of DI<sup>31</sup>. Moreover, an increased beta-cell DI does not necessarily imply improved beta-cell health. This is because an increase in DI via an increased demand of insulin secretion to compensate for decreased insulin sensitivity (that is, higher allostatic load) has been associated with deterioration of beta-cell function compared with increasing DI via improved insulin sensitivity<sup>31,41,42</sup>. Therefore, we evaluated both insulin secretion and insulin sensitivity for calculating beta-cell DI, and our results are in line with preclinical and clinical studies suggesting that increasing beta-cell DI through insulin sensitivity is beneficial for beta-cell health.

#### **Conclusion and perspectives**

Among adults with T2D within 7 years of diagnosis, exercise in addition to diet-induced weight loss increases late-phase DI across a 16-week intervention. The most pronounced benefits were observed with exercise six times per week.

The direction of the exercise effects on the oral DI and oral ISI are consistent with an additional benefit when added to diet-induced weight loss. In contrast to the linear dose-response relationship observed with glucose stimulation only, the oral DI and oral ISI displayed a curve-linear relationship with diminished returns when comparing three and six exercise sessions per week. Hence, data from the meal stimulation suggest that increasing the exercise dose beyond three times per week may be redundant to gain additional benefits of exercise on beta-cell function when performed in conjunction with diet-induced weight loss. Further research is needed to confirm this.

## Methods

#### Study design

The study was a 16-week, parallel-group, four-arm, assessor-blinded, randomized clinical trial conducted between February 2019 and October 2021 at the Centre for Physical Activity Research (CFAS), Rigshospitalet, Copenhagen, Denmark. The study was preregistered at ClinicalTrials.gov (NCT03769883) and was approved by the Scientific Ethical Committee of the Capital Region of Denmark (approval number H-18038298) before the commencement of any study procedures. Guidelines from the Helsinki Declaration were followed, and the data are reported following the CONSORT guideline for multi-arm trials<sup>43</sup> and the REPORT standards<sup>43</sup>. The study protocol for this clinical trial is available in the Supplementary Information and has been published previously<sup>22</sup>. The prespecified full statistical analysis plan (SAP) was completed and uploaded to our website before commencing any statistical analyses (https://aktivsundhed.dk/images/docs/SAP\_doseex\_nov21.pdf).

#### Participants and eligibility criteria

Participants were recruited through the media, municipalities and the Danish Health Data Authorities. The potential participants contacted the study nurse and completed the screening process before the medical examination. The main inclusion criteria were (1) men and women aged 18–80 years, (2) diagnosed with T2D within <7 years, (3) no current treatment with insulin and (4) BMI > 27 kg/m<sup>2</sup> and <40 kg/m<sup>2</sup>. All participants provided written and oral informed consent before any testing.

#### Interventions

CON received standard care and was encouraged to maintain habitual physical activity and dietary habits throughout the study. DCON received standard care and dietary intervention. MED received standard care, dietary intervention and an exercise intervention with two aerobic training sessions per week and one combined aerobic and resistance training session per week, totaling 150–165 min of exercise training per week. HED received the standard care and dietary interventions as described above but had twice as much exercise as MED, with a total of four aerobic training sessions per week, totaling 300–330 min of exercise training per week.

**Standard care component.** Standard care included pharmacological management of blood glucose, blood lipids and blood pressure according to a prespecified algorithm and was managed by an endocrinologist

who was blinded for participant allocation<sup>22</sup>. To minimize an influence on the findings of poor glucose control upon study entry, medical standardization was introduced according to the prespecified treat-to-target algorithm for 6 weeks before the baseline measurements. Furthermore, the pharmacological treatment was evaluated according to the algorithm following baseline measurements and at week 12 of the intervention. The treatment targets were in line with current guidelines. In adjunct to the algorithm, pharmacological treatment was adapted to mitigate subjective signs of hypotension or hypoglycemia. Blood lipids, blood pressure and blood glucose were measured before the intervention and 4, 12 and 16 weeks into the intervention. In case of any adverse events, the participants were advised to contact the study nurse. At each visit, the study nurse interviewed all participants about potential adverse events. The adverse events definition followed ICH E2A guidelines<sup>44</sup>.

**Dietary component.** Daily energy requirements were estimated using the age-adjusted Oxford equation<sup>45</sup>. The dietary intervention aimed at -25–30% energy deficit per day with a macronutrient distribution within the range of 45–60 energy percent (E%) carbohydrate, 15–20E% protein and 20–35E% fat (<7E% saturated fat). The intervention consisted of individualized recommendations and recipes. A clinical dietician implemented the plan at three sessions during the intervention, and adjustments were performed based on self-reported, 3-day food records.

**Exercise component.** The exercise intervention consisted of both aerobic and resistance training, and the first 2 weeks served as a familiarization period. The aerobic training sessions of 30-min duration had a target intensity of 60-100% of maximal heart rate (HR<sub>max</sub>). Throughout the intervention, the relative time spent exercising in intensity zone 80-100% of HR<sub>max</sub> was increased, and the relative time spent in the intensity zone 60-79% of HR<sub>max</sub> was reduced accordingly. Resistance training was added in combined sessions with 30 min of aerobic training and 30-45 min of resistance training. The resistance training consisted of three sets in the main muscle groups, for example, chest press, leg press, back row, and leg extension. The 8–12 repetitions aimed at a resistance consistent with 0-3 repetitions in reserve<sup>46</sup>. All heart-rate profiles were recorded during the exercise interventions (Polar V800), and all training sessions were supervised by educated trainers.

#### **Experimental days**

Two experimental days were conducted at baseline and repeated at 16-week follow-up. Forty-eight hours before the experimental days, the participants were instructed to discontinue glucose-lowering medication use and refrain from any exercise. Moreover, no alcohol or caffeine was permitted 24 h before the visits, and the participants were instructed to maintain their habitual diet. The participants arrived at the testing facilities at 07:30 am after an overnight fast ( $\geq 10$  h fasting). Experimental days 1 and 2 were planned to be separated by 1 week.

**Experimental day 1.** The participants completed a 3-h MMTT. The liquid meal was prepared using 400 ml of Nestlé Resource with an additional 36 g of dextrose (total energy content, 735 kcal; E%, 64/24/12 carbohydrate/fat/protein). Paracetamol (1.5 g) was added to assess gastric emptying. Body weight was measured with an electronic scale, and height was measured with a Holtain stadiometer according to standard procedures.  $VO_{2max}$  was assessed using indirect calorimetry (Quark CPET, Cosmed) on a Monark LC4 bicycle (Monark Exercise). The test was performed with a 5-min warm-up followed by increases of 20 watts/ min until exhaustion. Maximum muscle strength was assessed by two exercises performed in resistance training machines (chest press, leg extension) via estimating the maximum weight (kg) that could be lifted once with a full range of motion with proper form (that is, 1 RM).

Experimental day 2. A three-stage hyperglycemic clamp was performed. After baseline blood sampling, a priming bolus of  $[6.6^{-2}H_{2}]$ glucose was injected intravenously and a continuous tracer infusion was initiated. The bolus dose and infusion rate of the tracer depended on the participant's fasting glucose level and body weight as described elsewhere<sup>5</sup>. After 2 h of tracer infusion, hyperglycemia was introduced by clamping glucose at 5.4 mM above fasting glucose (whereas the absolute postintervention clamp glucose level was equal to the preintervention clamp level). An initial increase in blood glucose was brought about by a square-wave glucose infusion lasting 15 min. After this, the glucose concentration was kept constant by adjusting GIRs based on blood glucose measurements (ABL 8 series, Radiometer) performed every 5 min according to an automated algorithm<sup>5</sup>. After 2 h of hyperglycemia, a continuous GLP-1 infusion was initiated at a rate of 0.5 pmol/kg/min, and after 1 h of hyperglycemia + GLP-1 infusion, an intravenous bolus of arginine hydrochloride (5 g given over 30 s) was administered to provide a maximal stimulus to the beta cells, leading to secretion of remaining intracellular vesicles of insulin. Before baseline sampling, the participant voided. Every time the participant voided during the clamp, the urine was accumulated, and urinary glucose concentration was measured at the end of the procedure.

**Free-living measurements.** Assessments of free-living physical activity and blood pressure were recorded by the participants between the 2 study days. Physical activity was also assessed with physical activity monitors (AX3, Axivity) for 7 consecutive days. Blood pressure was assessed with home-based resting measurements across 3 days, including three measurements morning and evening. Furthermore, a 3-day record of total dietary intake was completed at baseline, during the intervention period (at weeks 4 and 12), and during the 3 days leading up to follow-up testing.

**Blood sample analyses.** Blood samples (plasma insulin, C-peptide, glucose, HbA1c, LDL-C, triglycerides and paracetamol) were analyzed at the Department of Clinical Biochemistry, Rigshospitalet, using standard procedures. GLP-1 and GIP were analyzed using in-house carboxy-terminal radioimmunoassays. The total GLP-1 assay (codename 89390) is based on the amidated COOH terminus and therefore measures GLP-1(7–36)NH<sub>2</sub> and GLP-1(9–36) NH<sub>2</sub>. The assay results, therefore, reflect the secretion rate of GLP-1 (refs. 47,48). The total GIP assay (codename 80867) reacts fully with intact GIP and amino-terminally truncated forms<sup>49</sup>. The glucose tracer [6,6<sup>-2</sup>H<sub>2</sub>]glucose was used for whole-body measurements of  $R_a$  and  $R_d$  of glucose during steady-state hyperglycemia and was calculated using non-steady-state equations<sup>50</sup> adapted for stable isotopes<sup>51,52</sup>.

#### Participant compensation

All participants received up to DKK 6,000 (€800) in total to cover lost earnings, transport and discomfort. The transaction was completed upon completion of the study (all four full laboratory days (V1, V2, V6 and V7) or upon withdrawal). For every completed day of laboratory testing, participants received DKK 1,000. Moreover, DKK 500 in compensation was added per biopsy (up to four in total). To prevent loss to follow-up in the CON group, we offered three supervised training sessions and a free 16-week membership in a fitness center following final testing.

#### Outcomes

**Primary outcome.** The primary outcome was the change in late-phase DI from baseline to the 16-week follow-up, reflecting the beta-cell response during the last 30 min of the hyperglycemic stage<sup>15</sup>. DI was calculated as the product of late-phase ISR and late-phase ISI (designated secondary outcomes, see below).

Secondary outcomes. Secondary outcomes were prespecified in the SAP (designated 'Major secondary outcomes' in the SAP) and included the late-phase ISR, late-phase ISI derived during the last 30 min of the hyperglycemic stage, and the oral DI, oral ISI and oral ISR derived from the MMTT<sup>53</sup>. Late-phase ISR was calculated from the deconvoluted C-peptide measurements<sup>54</sup> and subsequently normalized to ambient blood glucose concentrations. Late-phase ISI was calculated as the GIR divided by the product of insulin and glucose<sup>39</sup>. Oral DI was calculated as the product of oral ISI and oral ISR. Oral ISI (the Matsuda index) was calculated as 10,000/ $\checkmark$ (fasting glucose × fasting insulin) × (mean glucose<sub>0-120min</sub> × mean insulin<sub>0-120min</sub>), and oral ISR was calculated as the tAUC for glucose divided by the tAUC for insulin from time 0 to 120 min during the MMTT<sup>53</sup>.

**Exploratory outcomes.** The exploratory outcomes (designated 'Other secondary outcomes' in the SAP) included the change (baseline to 16-week follow-up) in first-phase ISR, EGP, first-phase DI, ISI and ISR, as well as HbA1c, LDL-C, fasting glucose, fasting insulin, fasting C-peptide, fasting triglycerides, systolic blood pressure, diastolic blood pressure, body weight, absolute  $VO2_{max}$ , relative  $VO2_{max}$ , 1 RM for chest press and leg extension (both absolute and relative to body weight), and tAUC and iAUC in glucose, insulin, C-peptide, GLP-1, GIP and paracetamol from the MMTT. AUCs for the different time periods were calculated using the trapezoidal rule.  $R_a$  and  $R_d$  were calculated from glucose tracers during clamp-induced steady-state hyperglycemia. Adverse events were self-reported.

**Post hoc outcomes.** Post hoc outcomes included intensification (yes or no), reduction (yes or no) and discontinuation (yes or no) for glucose-lowering and blood pressure-lowering medications. Due to restrictions in our pharmacological treatment algorithm regarding lipid-lowering medications, only intensifications were assessed for this outcome.

#### **Randomization and blinding**

The participants were randomly allocated to the four intervention arms upon successful completion of the baseline measurements. An independent statistician (author R.C.) prepared a computer-generated randomization schedule in a ratio of 1:1:1:1, stratified by sex. To ensure concealment, the (permuted) block sizes were not disclosed. The schedule was forwarded to a secretary who was not involved in any study procedures and stored on a password-protected computer. Sequentially numbered, opaque, sealed envelopes were prepared and stored in a locked cabinet before commencing the recruitment. The envelopes were lined with aluminum foil to render the envelope impermeable to intense light. Following the conclusion of the hyperglycemic clamp, the appropriate envelope was opened by a study nurse, and the participant was informed about the allocation stated on the card inside the envelope. The participant received the allocation in a closed room. As such, the participants were blinded for treatment allocation until after the completion of the hyperglycemic clamp. Following the baseline assessment, blinding of the participants was no longer possible. Both study personnel involved with the data collection and the study endocrinologist managing pharmacological treatment and safety were blinded to allocation. The clinical results used for pharmacological management and safety assessment were presented to the endocrinologist by the study nurse without disclosing participant allocation.

#### Sample size and power considerations

We expected that an exercise intervention would increase the late-phase DI by 1.5 arbitrary units (a.u.) more than the control group, with a standard deviation of 1.5 a.u. of the change in the exercise and 1.0 a.u. in the control group<sup>5</sup>. For a contrast in a one-way analysis of variance (ANOVA) with four means (1.5, 1.0, 0.5, 0.0) and contrast coefficients (1, 0, 0, -1) using a two-sided significance level of 0.05, assuming an

error standard deviation of 1.5 and a balanced design, a total sample size of 80 participants in the PP population (approximately 20 participants in each group) would yield statistical power of 87.7%.

#### Statistical analysis

According to the protocol and the SAP, the analysis of the primary outcome was based on the as-observed population (missing data were not imputed in the primary analysis)<sup>55,56</sup>, as well as the PP population. The 'Full Analysis Set' for the ITT population included all randomized participants irrespective of their compliance with the interventions. The PP population criteria included (1) completion of the primary outcome assessment (all groups), (2) compliance with the diet protocol defined as being within  $\pm 30\%$  of the prescribed energy intake (DCON, MED and HED), and (3) compliance with the exercise training protocol defined as completing  $\geq$ 70% of the prescribed exercise volume across the intervention period (from weeks 2 to 16) (MED and HED). Missing data were assumed to be missing at random. Continuous data, including the primary, secondary and exploratory outcomes, were analyzed using constrained baseline longitudinal analysis via a linear mixed model<sup>57</sup>. As the baseline value is a part of the outcome vector, all participants with at least one measurement (baseline or follow-up) were included in the analyses<sup>57</sup>. The model included fixed effects for time (two levels), treatment (coded 0 for all groups at baseline and coded 0, 1, 2 or 3 at follow-up for CON, DCON, MED and HED, respectively) and sex (two levels), as well as the unique patient identifier as a random effect. The potentially biased PP population analysis was further adjusted for putative confounders: diabetes duration and baseline maximal oxygen consumption (ml O<sub>2</sub>/kg/min). Data are presented as the difference in the mean changes with 95% confidence intervals unless stated otherwise. The adequacy of the models was investigated via the predicted values and residuals. If the model assumptions were violated, the analyses were conducted using the log-transformed data and subsequently exponentiated for interpretation. Back-transformed data were expressed as the ratio of the geometric mean and interpreted as either percent change from baseline (within group) or difference in change between groups. A linear trend (interpreted as a linear doseresponse relationship) was examined by treating each treatment category as a continuous variable in the main model and tested using a Wald test (Pvalue reported). Linearity was inspected visually, and the P for trend was calculated only for the primary and secondary outcomes to the extent that the relationship was linear (that is, for late-phase DI and late-phase ISI). Sensitivity analyses were performed using multiple linear imputation procedures with the change in outcomes (post-pre values)55. The model included all covariates included in the main model, and beta coefficient and standard errors were based on 30 imputed data sets and adjusted for between-imputation variability58. Dichotomous outcomes were analyzed using logistic regression analyses. As sparsity of dichotomous outcomes (as expected for medications) invalidates the confidence intervals, exact logistic regression (exlogistic in Stata) was used when cases were  $<5^{59,60}$ . A post hoc statistical mediation analysis was performed to examine the extent to which the observed treatment effect (in the intervention groups) on the primary and secondary outcomes was mediated by the change in body weight. An exploratory statistical mediation analysis was performed in R<sup>61</sup> to examine the extent to which the observed treatment effect (in the intervention groups) on the primary and key secondary outcomes was mediated by the change in body weight. The lme4 package was used to construct the linear mixed models for the analysis<sup>62</sup>. This simple mediation analysis partitions the total causal effect into average direct effects (ADE) and average causal mediation effects (ACME; otherwise known as indirect effects). Bias-corrected and accelerated 95% confidence intervals were generated via nonparametric bootstrap analysis (2,000 resamples with replacement).

All non-hypothesis-based comparisons (that is, on the secondary and exploratory outcomes) are per definition considered exploratory and supportive of the interpretation of the primary outcome. If the global test of significance indicated between-group differences  $(P < 0.1)^{63}$ , all outcomes (primary, secondary, exploratory and post hoc) on pairwise comparisons were explored. Although no corrections for multiplicity were performed, family-wise type 1 error rate on the primary outcome was retained by using a hierarchical analytic approach<sup>63</sup>. In accordance with our prespecified SAP, the six prespecified hierarchical hypotheses (based on a superiority assumption) were tested using the prespecified sequence: (1) CON versus HED, (2) CON versus MED, (3) CON versus DCON, (4) DCON versus HED, (5) DCON versus MED, (6) MED versus HED. If we failed to progress from any of the prior between-group comparisons (P > 0.05), the subsequent P values and confidence intervals were regarded as indicators of associations rather than causality. The statistical significance level (for superiority) was set at  $\alpha < 0.05$  (two-sided). The statistical analyses were performed using Stata/SE (StataCorp), version 17.1.

#### **Reporting summary**

Further information on research design is available in the Nature Portfolio Reporting Summary linked to this article.

#### Data availability

Data are not available for download owing to privacy and ethical restrictions under the European Union's General Data Protection Regulation (EUGDPR). Specific requests for access to the trial and individual-level and unique biological data included in this article may be sent to mathias.ried-larsen@regionh.dk. Based on the request, access may be provided to a named individual in agreement with the rules and regulations of the Danish Data Protection Agency and the National Committee on Health Research Ethics. Requests will be considered from the date of publication of this article.

## **Code availability**

The syntax files (Stata) are available upon request to mathias. ried-larsen@regionh.dk.

#### References

- Schwartz, S. S. et al. A unified pathophysiological construct of diabetes and its complications. *Trends Endocrinol. Metab.* 28, 645–655 (2017).
- 2. Magkos, F. et al. Effects of moderate and subsequent progressive weight loss on metabolic function and adipose tissue biology in humans with obesity. *Cell Metab.* **23**, 591–601 (2016).
- 3. Lim, E. L. et al. Reversal of type 2 diabetes: normalisation of beta cell function in association with decreased pancreas and liver triacylglycerol. *Diabetologia* **54**, 2506–2514 (2011).
- 4. Taylor, R. et al. Remission of human type 2 diabetes requires decrease in liver and pancreas fat content but is dependent upon capacity for  $\beta$  cell recovery. *Cell Metab.* **28**, 547–556.e3 (2018).
- Karstoft, K. et al. Mechanisms behind the superior effects of interval vs continuous training on glycaemic control in individuals with type 2 diabetes: a randomised controlled trial. *Diabetologia* 57, 2081–2093 (2014).
- Dela, F., von Linstow, M. E., Mikines, K. J. & Galbo, H. Physical training may enhance beta-cell function in type 2 diabetes. *Am. J. Physiol. Endocrinol. Metab.* 287, E1024–E1031 (2004).
- 7. Rogers, M. A. et al. Improvement in glucose tolerance after 1 wk of exercise in patients with mild NIDDM. *Diabetes Care* **11**, 613–618 (1988).
- Krotkiewski, M. et al. The effects of physical training on insulin secretion and effectiveness and on glucose metabolism in obesity and type 2 (non-insulin-dependent) diabetes mellitus. *Diabetologia* 28, 881–890 (1985).
- Eriksen, L., Dahl-Petersen, I., Haugaard, S. B. & Dela, F. Comparison of the effect of multiple short-duration with single long-duration exercise sessions on glucose homeostasis in type 2 diabetes mellitus. *Diabetologia* 50, 2245–2253 (2007).

- 10. Johansen, M. Y. et al. Effects of an intensive lifestyle intervention on the underlying mechanisms of improved glycaemic control in individuals with type 2 diabetes: a secondary analysis of a randomised clinical trial. *Diabetologia* **63**, 2410–2422 (2020).
- Curran, M. et al. The benefits of physical exercise for the health of the pancreatic β-cell: a review of the evidence. *Exp. Physiol.* 105, 579–589 (2020).
- 12. Slentz, C. A. et al. Effects of exercise training intensity on pancreatic beta-cell function. *Diabetes Care* **32**, 1807–1811 (2009).
- 13. Zhang, S., Wei, Y. & Wang, C. Impacts of an exercise intervention on the health of pancreatic beta-cells: a review. *Int. J. Environ. Res. Public Health* **19**, 7229 (2022).
- AbouAssi, H. et al. The effects of aerobic, resistance, and combination training on insulin sensitivity and secretion in overweight adults from STRRIDE AT/RT: a randomized trial. J. Appl. Physiol. (1985) 118, 1474–1482 (2015).
- Hannon, T. S. et al. Review of methods for measuring β-cell function: design considerations from the Restoring Insulin Secretion (RISE) Consortium. *Diabetes Obes. Metab.* **20**, 14–24 (2018).
- 16. Karstoft, K. et al. Glucose effectiveness, but not insulin sensitivity, is improved after short-term interval training in individuals with type 2 diabetes mellitus: a controlled, randomised, crossover trial. *Diabetologia* **60**, 2432–2442 (2017).
- Davies, M. J. et al. Management of Hyperglycemia in Type 2 Diabetes, 2022. A Consensus Report by the American Diabetes Association (ADA) and the European Association for the Study of Diabetes (EASD). *Diabetes Care* 45, 2753–2786 (2022).
- 18. Sathananthan, M. et al. Six and 12 weeks of caloric restriction increases  $\beta$  cell function and lowers fasting and postprandial glucose concentrations in people with type 2 diabetes. *J. Nutr.* **145**, 2046–2051 (2015).
- Goto, Y. et al. Improvement of skeletal muscle insulin sensitivity by 1 week of SGLT2 inhibitor use. *Endocr. Connect.* 9, 599–606 (2020).
- 20. Omar, B. & Ahrén, B. Pleiotropic mechanisms for the glucose-lowering action of DPP-4 inhibitors. *Diabetes* **63**, 2196–2202 (2014).
- 21. Bahne, E. et al. Metformin-induced glucagon-like peptide-1 secretion contributes to the actions of metformin in type 2 diabetes. *JCI Insight* **3**, e93936 (2018).
- 22. Lyngbaek, M. P. P. et al. The effects of different doses of exercise on pancreatic  $\beta$ -cell function in patients with newly diagnosed type 2 diabetes: study protocol for and rationale behind the "DOSE-EX" multi-arm parallel-group randomised clinical trial. *Trials* **22**, 244 (2021).
- Malin, S. K. et al. Pancreatic β-cell function increases in a linear dose-response manner following exercise training in adults with prediabetes. *Am. J. Physiol. Endocrinol. Metab.* **305**, E1248–E1254 (2013).
- 24. Madsen, S. M., Thorup, A. C., Overgaard, K. & Jeppesen, P. B. High intensity interval training improves glycaemic control and pancreatic  $\beta$  cell function of type 2 diabetes patients. *PLoS ONE* **10**, e0133286 (2015).
- 25. Malandrucco, I. et al. Very-low-calorie diet: a quick therapeutic tool to improve  $\beta$  cell function in morbidly obese patients with type 2 diabetes. *Am. J. Clin. Nutr.* **95**, 609–613 (2012).
- 26. Lean, M. E. J. et al. Primary care-led weight management for remission of type 2 diabetes (DiRECT): an open-label, cluster-randomised trial. *Lancet* **391**, 541–551 (2018).
- 27. Kanat, M. et al. Impaired early- but not late-phase insulin secretion in subjects with impaired fasting glucose. *Acta Diabetol.* **48**, 209–217 (2011).
- 28. King, D. S. et al. Insulin secretory capacity in endurance-trained and untrained young men. *Am. J. Physiol. Endocrinol. Metab.* **259**, E155–E161 (1990).

#### Article

- 29. Dela, F. Functional adaptation of the human  $\beta$ -cells after frequent exposure to noradrenaline. *J. Physiol.* **593**, 3199–3206 (2015).
- Shima, K., Hirota, M., Sato, M., Iwami, T. & Oshima, I. Effect of exercise training on insulin and glucagon release from perfused rat pancreas. *Horm. Metab. Res* 19, 395–399 (1987).
- Vazquez Arreola, E., Hanson, R. L., Bogardus, C. & Knowler, W. C. Relationship between insulin secretion and insulin sensitivity and its role in development of type 2 diabetes: beyond the disposition index. *Diabetes* 71, 128–141 (2022).
- 32. Mingrone, G. et al. Insulin sensitivity depends on the route of glucose administration. *Diabetologia* **63**, 1382–1395 (2020).
- Roberts-Thomson, K. M. et al. Oral and intravenous glucose administration elicit opposing microvascular blood flow responses in skeletal muscle of healthy people: role of incretins. *J. Physiol.* 600, 1667–1681 (2022).
- Akerstrom, T. et al. Increased skeletal muscle capillarization enhances insulin sensitivity. *Am. J. Physiol. Endocrinol. Metab.* 307, E1105–E1116 (2014).
- Sylow, L. & Richter, E. A. Current advances in our understanding of exercise as medicine in metabolic disease. *Curr. Opin. Physiol.* 12, 12–19 (2019).
- Dubé, J. J. et al. Effects of weight loss and exercise on insulin resistance, and intramyocellular triacylglycerol, diacylglycerol and ceramide. *Diabetologia* 54, 1147–1156 (2011).
- Frías, J. P. et al. Tirzepatide versus semaglutide once weekly in patients with type 2 diabetes. *N. Engl. J. Med.* 385, 503–515 (2021).
- Lundby, C., Montero, D. & Joyner, M. Biology of VO<sub>2</sub> max: looking under the physiology lamp. *Acta Physiol.* (*Oxf.*) **220**, 218–228 (2017).
- Meneilly, G. S. & Elliott, T. Assessment of insulin sensitivity in older adults using the hyperglycemic clamp technique. *J. Am. Geriatr.* Soc. 46, 88–91 (1998).
- Trabulsi, J. & Schoeller, D. A. Evaluation of dietary assessment instruments against doubly labeled water, a biomarker of habitual energy intake. *Am. J. Physiol. Endocrinol. Metab.* 281, E891–E899 (2001).
- 41. Kahn, S. E. et al. Effects of rosiglitazone, glyburide, and metformin on  $\beta$ -cell function and insulin sensitivity in ADOPT. *Diabetes* **60**, 1552–1560 (2011).
- Boland, B. B. et al. Pancreatic β-cell rest replenishes insulin secretory capacity and attenuates diabetes in an extreme model of obese type 2 diabetes. *Diabetes* 68, 131–140 (2019).
- Atlas Collaboration. Reconstruction of hadronic decay products of tau leptons with the ATLAS experiment. *Eur. Phys. J. C* 76, 295 (2016).
- ICH E2A Clinical safety data management: definitions and standards for expedited reporting (European Medicines Agency, 1995).
- Henry, C. J. K. Basal metabolic rate studies in humans: measurement and development of new equations. *Public Health Nutr.* 8, 1133–1152 (2005).
- Helms, E. R., Cronin, J., Storey, A. & Zourdos, M. C. Application of the repetitions in reserve-based rating of perceived exertion scale for resistance training. *Strength Cond. J.* 38, 42–49 (2016).
- Ørskov, C., Rabenhøj, L., Wettergren, A., Kofod, H. & Holst, J. J. Tissue and plasma concentrations of amidated and glycine-extended glucagon-like peptide I in humans. *Diabetes* 43, 535–539 (1994).
- Wewer Albrechtsen, N. J. et al. Stability of glucagon-like peptide 1 and glucagon in human plasma. *Endocr. Connect.* 4, 50–57 (2015).
- Lindgren, O. et al. Incretin hormone and insulin responses to oral versus intravenous lipid administration in humans. J. Clin. Endocrinol. Metab. 96, 2519–2524 (2011).

- 50. Steele, R. Influences of glucose loading and of injected insulin on hepatic glucose output. *Ann. N. Y. Acad. Sci.* **82**, 420–430 (1959).
- Plomgaard, P. et al. Tumor necrosis factor-alpha induces skeletal muscle insulin resistance in healthy human subjects via inhibition of Akt substrate 160 phosphorylation. *Diabetes* 54, 2939–2945 (2005).
- Matthews, D. E. Radioactive and Stable Isotope Tracers in Biomedicine: Principles and Practice of Kinetic Analysis: by Robert R Wolfe, 1992, 471 pages, hardcover, \$89.95. John Wiley & Sons, Inc, Somerset, NJ. Am. J. Clin. Nutr. 58, 452 (1993).
- Maki, K. C., McKenney, J. M., Farmer, M. V., Reeves, M. S. & Dicklin, M. R. Indices of insulin sensitivity and secretion from a standard liquid meal test in subjects with type 2 diabetes, impaired or normal fasting glucose. *Nutr. J.* 8, 22 (2009).
- 54. Van Cauter, E., Mestrez, F., Sturis, J. & Polonsky, K. S. Estimation of insulin secretion rates from C-peptide levels: comparison of individual and standard kinetic parameters for C-peptide clearance. *Diabetes* **41**, 368–377 (1992).
- 55. White, I. R., Horton, N. J., Carpenter, J. & Pocock, S. J. Strategy for intention to treat analysis in randomised trials with missing outcome data. *BMJ* **342**, d40 (2011).
- Detry, M. A. & Lewis, R. J. The intention-to-treat principle: how to assess the true effect of choosing a medical treatment. *JAMA* **312**, 85–86 (2014).
- 57. Coffman, C. J., Edelman, D. & Woolson, R. F. To condition or not condition? Analysing 'change' in longitudinal randomised controlled trials. *BMJ Open* **6**, e013096 (2016).
- Rubin, D. B. Multiple imputation after 18+ years. J. Am. Stat. Assoc. 91, 473–489 (1996).
- 59. Greenland, S., Mansournia, M. A. & Altman, D. G. Sparse data bias: a problem hiding in plain sight. *BMJ* **352**, i1981 (2016).
- 60. Kirk, S., Scott, B. J. & Daniels, S. R. Pediatric obesity epidemic: treatment options. J. Am. Diet. Assoc. **105**, S44–S51 (2005).
- 61. R: a language and environment for statistical computing. (R Foundation for Statistical Computing, 2018).
- 62. Bates, D., Machler, M., Bolker, B. & Walker, S. lme4: linear mixed-effects models using Eigen and S4; https://github.com/ lme4/lme4 (2022).
- 63. Dmitrienko, A. & D'Agostino, R. B.Sr. Multiplicity considerations in clinical trials. *N. Engl. J. Med.* **378**, 2115–2122 (2018).

#### Acknowledgements

We thank the study participants for their time and engagement in this study, as well as current and former staff at CFAS, Rigshospitalet. We also thank the current and former staff at the Centre for Diabetes Research at the Municipality of Copenhagen for their support in recruitment and intervention delivery. The project was supported by a grant from TrygFonden and Svend Andersen Fonden. CFAS is supported by TrygFonden (grants ID 101390, ID 20045 and ID 125132). R.C. is from the Section for Biostatistics and Evidence-Based Research, the Parker Institute, Bispebjerg and Frederiksberg Hospital, which is supported by a core grant from the Oak Foundation (OCAY-18-774-OFIL). M.P.P.L. was supported by a research grant from the Danish Diabetes Academy (grant no. NNF17SA0031406), which is funded by the Novo Nordisk Foundation.

#### **Author contributions**

B.K.P., K.K., T.P.A. and M.R.-L. conceived the study. M.P.P.L., G.E.L. and M.R.-L. wrote the first draft. M.R.-L. is the principal investigator. M.R.-L., M.P.P.L. and G.E.L. had full access to the data in the study, verified the data, and had full responsibility for the decision to submit and publish. M.P.P.L., G.E.L., M.R.-L., T.P.A., K.K., B.K.P., T.P.J.S., R.C., G.V.H. and J.J.H. contributed to protocol development and study design. G.E.L., M.P.P.L., S.L.B., C.S.F., N.S.N., B.L., U.N., M.Ø., K.T. and B.T. performed the experiments and collected the data. T.P.A. and K.T. performed pharmacological management. B.H., J.J.H. and G.V.H. performed biochemical analyses of incretins or stable isotopes. J.C.B. performed the accelerometer analyses. M.R.-L., K.T., N.S.N., G.E.L. and M.P.P.L. integrated and quality-checked the data. M.R.-L. and C.G.D. performed the statistical analyses. G.E.L., M.P.P.L., R.C. and M.R.-L. wrote the SAP. All authors read the SAP, critically revised it for important intellectual content and approved the final version. All authors read the manuscript, critically revised it for important intellectual content and approved the final version.

#### **Competing interests**

J.J.H. is a member of advisory boards for Novo Nordisk. All other authors declare that they have no competing interests.

#### **Additional information**

**Extended data** is available for this paper at https://doi.org/10.1038/s42255-023-00799-7.

**Supplementary information** The online version contains supplementary material available at https://doi.org/10.1038/s42255-023-00799-7.

**Correspondence and requests for materials** should be addressed to Mathias Ried-Larsen.

**Peer review information** *Nature Metabolism* thanks the anonymous reviewers for their contribution to the peer review of this work. Primary handling editor: Isabella Samuelson, in collaboration with the *Nature Metabolism* team.

**Reprints and permissions information** is available at www.nature.com/reprints.

**Publisher's note** Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

**Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons license and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this license, visit http://creativecommons. org/licenses/by/4.0/.

© The Author(s) 2023

Grit E. Legaard<sup>1,12</sup>, Mark P. P. Lyngbæk @<sup>1,12</sup>, Thomas P. Almdal @<sup>2,3</sup>, Kristian Karstoft @<sup>1,4</sup>, Sebastian L. Bennetsen @<sup>1</sup>, Camilla S. Feineis<sup>1</sup>, Nina S. Nielsen @<sup>1</sup>, Cody G. Durrer<sup>1</sup>, Benedikte Liebetrau @<sup>1</sup>, Ulrikke Nystrup<sup>1</sup>, Martin Østergaard<sup>1</sup>, Katja Thomsen @<sup>1</sup>, Beckey Trinh @<sup>1</sup>, Thomas P. J. Solomon @<sup>5</sup>, Gerrit Van Hall<sup>6,7</sup>, Jan Christian Brønd @<sup>8</sup>, Jens J. Holst @<sup>9</sup>, Bolette Hartmann<sup>9</sup>, Robin Christensen @<sup>10,11</sup>, Bente K. Pedersen @<sup>1</sup> & Mathias Ried-Larsen @<sup>1,8</sup> 🖂

<sup>1</sup>Centre for Physical Activity Research, Rigshospitalet, Copenhagen, Denmark. <sup>2</sup>Department of Endocrinology PE, Rigshospitalet, University of Copenhagen, Copenhagen, Denmark. <sup>3</sup>Department of Immunology & Microbiology, University of Copenhagen, Copenhagen, Denmark. <sup>4</sup>Department of Clinical Pharmacology, Bispebjerg-Frederiksberg Hospital, University of Copenhagen, Copenhagen, Denmark. <sup>5</sup>Blazon Scientific, London, UK. <sup>6</sup>Biomedical Sciences, Faculty of Health & Medical Science, University of Copenhagen, Rigshospitalet, Copenhagen, Denmark. <sup>7</sup>Clinical Metabolomics Core Facility, Clinical Biochemistry, University of Copenhagen, Rigshospitalet, Copenhagen, Denmark. <sup>7</sup>Clinical Metabolomics Core Facility, Clinical Biochemistry, University of Copenhagen, Rigshospitalet, Copenhagen, Denmark. <sup>8</sup>Department of Sports Science and Clinical Biomechanics, University of Southern Denmark. <sup>9</sup>Department of Biomedical Sciences and the Novo Nordisk Foundation Center for Basic Metabolic Research, University of Copenhagen, Denmark. <sup>10</sup>Section for Biostatistics and Evidence-Based Research, the Parker Institute, Bispebjerg and Frederiksberg Hospital, Copenhagen, Denmark. <sup>11</sup>Research Unit of Rheumatology, Department of Clinical Research, University of Southern Denmark, Odense, Denmark. <sup>12</sup>These authors contributed equally: Grit E. Legaard, Mark P.P. Lyngbæk. *Cenail: mathias.ried-larsen@regionh.dk* 

# Distribution of observed values



Extended Data Fig. 1|Box-plots for baseline values and by group at follow-up.

As the primary analyses are performed using a constrained baseline model, where all groups are assumed to be similar a baseline, the baseline values are not depicted by group. Circles denotes the individual participant values. Center line is the median values, light grey area is the lower inter quartile range, dark grey area is the upper interquartile range, the whiskers show + /-1.5 x the interquartile range, CON: control group (N = 20 independent samples), DCON: Dietary control group (N = 21 independent samples), MED: Moderate volume exercise (N = 20 independent samples), HED: High volume exercise (N = 21 independent samples), DI: Disposition index, ISI: Insulin sensitivity index, ISR: Insulin secretion rate, pmol: pico mol, mmol: milli mol, a.u.: arbitrary units, kg: kilograms.



Extended Data Fig. 2 | See next page for caption.

**Extended Data Fig. 2** | **The black circles represent baseline values, and the red circles represents the 16-week follow-up values.** Data are represented as the estimated means. Error bars are 95% confidence intervals. Results are adjusted for sex. Time 0-120 minutes is the hyperglycemic phase. Time 120-180 minutes is the hyperglycemic - and GLP-1 stimulation phase. Time 180-190 minutes is the hyperglycemic, GLP-1, and Arginine HCl stimulation phase. CON: control group (N = 20 independent samples), DCON: Dietary control group (N = 21 independent samples), MED: Moderate volume exercise (N = 20 independent samples), HED: High volume exercise (N = 21 independent samples).

## MMTT Glucose CON DCON 20 18 16 14 12 10 8 6 4 2 0 mmol/L MED HED 20 18 16 14 12 10 8 6 4 2 0 , so 6 °. <u>ر</u>ې ્રંડ , <sup>65</sup> ,<sub>8</sub>0 , so , 65 , 80 Time (minutes) Baseline 16 week follow-up

**Extended Data Fig. 3** | **The black circles represent baseline values, and the red circles represents the 16-week follow-up values.** Data are represented as the estimated means. Error bars are 95% confidence intervals. Results are adjusted for sex. CON: control group (N = 20 independent samples), DCON: Dietary

control group (N = 21 independent samples), MED: Moderate volume exercise (N = 20 independent samples), HED: High volume exercise (N = 21 independent samples).



**Extended Data Fig. 4** | **The black circles represent baseline values, and the red circles represents the 16-week follow-up values.** Data are represented as the estimated means. Error bars are 95% confidence intervals. Results are adjusted for sex. CON: control group (N = 20 independent samples), DCON: Dietary

control group (N = 21 independent samples), MED: Moderate volume exercise (N = 20 independent samples), HED: High volume exercise (N = 21 independent samples).



**Extended Data Fig. 5** | **The black circles represent baseline values, and the red circles represents the 16-week follow-up values.** Data are represented as the estimated means. Error bars are 95% confidence intervals. Results are adjusted for sex. CON: control group (N = 20 independent samples), DCON: Dietary

control group (N = 21 independent samples), MED: Moderate volume exercise (N = 20 independent samples), HED: High volume exercise (N = 21 independent samples).



**Extended Data Fig. 6** | **Represented as the estimated means.** Error bars are 95% confidence intervals. Results are adjusted for sex. CON: control group (N = 20 independent samples), DCON: Dietary control group (N = 21 independent samples), MED: Moderate volume exercise (N = 20 independent samples), HED: High volume exercise (N = 21 independent samples).





**Extended Data Fig. 7** | **The black circles represent baseline values, and the red circles represents the 16-week follow-up values.** Data are represented as the estimated means. Error bars are 95% confidence intervals. Results are adjusted for sex. CON: control group (N = 20 independent samples), DCON:

Dietary control group (N = 21 independent samples), MED: Moderate volume exercise (N = 20 independent samples), HED: High volume exercise (N = 21 independent samples).





Dietary control group (N = 21 independent samples), MED: Moderate volume exercise (N = 20 independent samples), HED: High volume exercise (N = 21 independent samples).

# nature portfolio

Corresponding author(s): Mathias Ried-Larsen

Last updated by author(s): 2023-27-03

# **Reporting Summary**

Nature Portfolio wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Portfolio policies, see our <u>Editorial Policies</u> and the <u>Editorial Policy Checklist</u>.

#### **Statistics**

For	all st	atistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.
n/a	Cor	firmed
	$\boxtimes$	The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement
	$\boxtimes$	A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
		The statistical test(s) used AND whether they are one- or two-sided Only common tests should be described solely by name; describe more complex techniques in the Methods section.
	$\boxtimes$	A description of all covariates tested
	$\boxtimes$	A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
		A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
		For null hypothesis testing, the test statistic (e.g. <i>F</i> , <i>t</i> , <i>r</i> ) with confidence intervals, effect sizes, degrees of freedom and <i>P</i> value noted <i>Give P values as exact values whenever suitable</i> .
$\boxtimes$		For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings

For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes

Estimates of effect sizes (e.g. Cohen's *d*, Pearson's *r*), indicating how they were calculated

Our web collection on statistics for biologists contains articles on many of the points above.

## Software and code

Policy information a	about <u>availability of computer code</u>
Data collection	No software was used
Data analysis	Data was analyzed usign STATA/SE (StataCorp, College Station, Texas), version 17.1.

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Portfolio guidelines for submitting code & software for further information.

#### Data

Policy information about availability of data

All manuscripts must include a data availability statement. This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A description of any restrictions on data availability
- For clinical datasets or third party data, please ensure that the statement adheres to our policy

Data are not available for download due to privacy/ethical restrictions under the EU GDPR. Specific requests for access to the trial and unique biological data as well as code may be sent to mathias.ried-larsen@regionh.dk. Based on the request access may be provided to a named individual in agreement with the rules and regulations of the Danish Data Protection Agency and the National Committee on Health Research Ethics.

## Human research participants

Policy information about studies involving human research participants and Sex and Gender in Research.

Reporting on sex and gender	Randomization was stratified by sex to ensure a balanced design. Accordingly the analyses were adjusted for sex.
Population characteristics	Adult (age 58 years, 35% female) persons with type 2 diabetes (4 years from diagnosis)
Recruitment	Participants were recruited through the media, municipalities, and the Danish Health Data Authorities. As participants self-referred to the study, this may have caused a selection bias towards healthier persons. Consequently, the study results may not be generalized to the entire population living with type 2 diabetes
Ethics oversight	Scientific Ethical Committee of the Capital Region of Denmark (approval number H-18038298)

Note that full information on the approval of the study protocol must also be provided in the manuscript.

# Field-specific reporting

Please select the one below that is the best fit for your research. If you are not sure, read the appropriate sections before making your selection.

Life sciences

🔀 Behavioural & social sciences 🛛 🗌 Ecological, evolutionary & environmental sciences

For a reference copy of the document with all sections, see <u>nature.com/documents/nr-reporting-summary-flat.pdf</u>

# Behavioural & social sciences study design

All studies must disclose on these points even when the disclosure is negative.

Study description	The study was a 16-week parallel-group, 4-arm, assessor-blinded, randomized clinical trial using quantitative data
Research sample	Danish Men and women aged 18–80 years, diagnosed with T2D within <7 years, no current treatment with insulin, BMI >27 kg/m2 and <40 kg/m2. The sample was recruited due to increased risk of morbidities where diet and exercise may alleviate these. Prior studies have indicated that to reestablish beta-cell function, the diabetes should be of a relative short duration and remaining beta-cell function should be present. Thus short diabetes duration was considered and we included persons without the need of exogenous insulin. The sample is likely not representative of the entire population living with type 2 diabetes
Sampling strategy	This was a convenience sample and the final sample size was determined by the N participants needed to reach a statistical power of >80%. Sample size was calculated based on a previous exercise study in persons with type 2 diabetes (DOI: 10.1007/ s00125-014-3334-5). For a contrast in a one-way ANOVA with four means (1.5, 1.0, 0.5, 0.0) and contrast coefficients (1, 0, 0, -1) using a two-sided significance level of 0.05, assuming an error standard deviation of 1.5 and a balanced design, a total sample size of 80 participants in the per-protocol population (approximately 20 participants in each group) would yield statistical power of 87.7%.
Data collection	The web-based Clinical Trial Management System EasyTrial was used for data entry and management (EasyTrial ApS). EasyTrial has been approved by the Danish Data Protection Board. Electronic case report forms (eCRF) and questionnaires will be generated by the sponsor in EasyTrial. Fields have been programmed with acceptable ranges for data entry. All paper material (CRF, blood screen results, questionnaires and dietary records) was collected and stored in a locked cabinet at CFAS, Rigshospitalet Denmark. All information from the paper material was entered twice by in non-consecutive order into the electronic back-end system. In case of discrepancies between the entries, the original paper record was consulted. Upon completion of the study, all paper material have been scanned and stored on the secured hospital server in an electronic form. Data management will be performed using appropriate statistical software Excel and STATA. To enable pseudonymised data, all participants was ascribed a unique participant identification (ID) number. The identification key (ID number to personal information) was stored on a password-protected computer, separate from the unique ID number and the database. Printed data was kept in a separate locked area with limited access. All patient-related information obtained during the study was handled in accordance with the Danish law for protection of personal data ("lov om behandling af personolysninger") and the Danish health law ("sundhedsloven"). The blood samples were registered from the hospital blood sample portal (Labka) and para-clinical observations will be obtained through "Sundhedsportaler". The study was reported to the Danish Data Protection Agency ("Region H's paraplyanmeldelse") VD-2018-516/ I-suite no. 6768. The researchers were blinded for allocation. No other persons were present during data collection other than the participants and the researchers.
Timing	February 2019 and October 2021
Data exclusions	Pre-defined exclusion criteria were defined. No participants were excluded after randomization.
Non-participation	Five participants were lost-to-follow-up; one due to malignancy, one was dissatisfied with group allocation, one refrained from study testing due to COVID19, and two due to musculoskeletal injuries.

The participants were randomly allocated to the 4 intervention arms upon successful completion of the baseline measurements. An independent statistician prepared a computer-generated randomization schedule in a ratio of 1:1:1:1, stratified by sex. To ensure concealment, the (permuted) block sizes were not disclosed. The schedule was forwarded to a secretary not involved in any study procedures and stored on a password-protected computer. Sequentially numbered opaque, sealed envelopes were prepared and stored in a locked cabinet prior to commencing the recruitment. The envelopes were lined with aluminum foil to render the envelope impermeable to intense light. Following the conclusion of the hyperglycemic clamp, the appropriate envelope was opened by a study nurse and the participant was informed about the allocation stated on the card inside the envelope. The participant received the allocation in a closed room. As such, the participants were blinded for treatment allocation until after the completion of the hyperglycemic clamp.

# Reporting for specific materials, systems and methods

Methods

We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.

MRI-based neuroimaging

#### Materials & experimental systems

n/a	Involved in the study	n/a	Involved in the study
$\times$	Antibodies	$\boxtimes$	ChIP-seq
$\boxtimes$	Eukaryotic cell lines	$\boxtimes$	Flow cytometry
$\boxtimes$	Palaeontology and archaeology	$\boxtimes$	MRI-based neuroim
$\boxtimes$	Animals and other organisms		
	🔀 Clinical data		
$\boxtimes$	Dual use research of concern		

# Clinical data

Policy information about <u>clinical studies</u> All manuscripts should comply with the ICMJE guidelines for publication of clinical research and a completed CONSORT checklist must be included with all submissions.

Clinical trial registration	NCT03769883
Study protocol	Submitted with the manuscript
Data collection	Conducted between February 2019 and October 2021 at the Centre for Physical Activity Research (CFAS), Rigshospitalet, Copenhagen, Denmark
Outcomes	The primary outcome was assessed using a hyperglycemic clamp method. The secondary outcomes where assessed using a hyperglycemic clamp method and a mixed meal tolerance test. Blood samples were drawn as outlined in the protocol and manuscript by blinded research staff. The samples were sent do a central lab at Rigshospitalet, where they were analyzed by staff unaware of allocation.