

REVIEW ARTICLE OPEN Circadian rhythm regulates the function of immune cells and participates in the development of tumors

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Circadian rhythms are present in almost all cells and play a crucial role in regulating various biological processes. Maintaining a stable circadian rhythm is essential for overall health. Disruption of this rhythm can alter the expression of clock genes and cancerrelated genes, and affect many metabolic pathways and factors, thereby affecting the function of the immune system and contributing to the occurrence and progression of tumors. This paper aims to elucidate the regulatory effects of BMAL1, clock and other clock genes on immune cells, and reveal the molecular mechanism of circadian rhythm's involvement in tumor and its microenvironment regulation. A deeper understanding of circadian rhythms has the potential to provide new strategies for the treatment of cancer and other immune-related diseases.

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FACTS

- Circadian rhythms profoundly affect various physiological processes in the body.
- Circadian rhythms influence the maturation, localization, and function of immune cells in a variety of ways (including genes, adhesion molecules, cytokines, etc.).
- The immune system can respond by influencing circadian rhythms.
- Circadian rhythms are involved in cell cycle regulation.
- The components related to circadian rhythm in tumor cells and their microenvironment are altered, and there is an epidemiological link between cancer risk and circadian rhythm gene expression.

OPEN QUESTIONS

- Can circadian rhythm considerations be used to optimize clinical administration, nutrition, and surgical timing to optimize patient outcomes?
- Can circadian rhythms be integrated into drug use and surgical timing for better outcomes?
- Can we predict cancer risk by detecting the expression status of circadian rhythm genes in combination with other markers?

INTRODUCTION

The 24-h cycle generated by the earth's rotation has caused the evolution of circadian rhythms of almost all life forms on the earth [1]. This evolutionarily conservative time maintenance mechanism enables organisms to adapt to the ever-changing environment [2, 3]. In essence, the circadian clock constitutes an automatic regulatory sequence for the expression, accumulation and degradation of clock gene products, forming an autonomous molecular oscillator [3]. The molecular clock in the animals controls the expression of genes that are output throughout the body, thereby controlling the activity and function of different cells and organs over time [4]. Therefore, mutations in the clock gene can lead to changes in various biological rhythmic behaviors [5].

Almost all physiological activities of mammals are regulated by the body's biological clock, including sleep-wake, eating-fasting, and activity-rest cycle [2]. Its master clock is located in the suprachiasmatic nucleus (SCN) of the hypothalamus, which can process light input signals and transmit timing information to other parts of the body [1]. Peripheral clocks are located in other tissues and organs, some are strictly dependent on the regulation of the central clock (such as the liver), and some remain relatively independent [6]. SCN neurons maintain synchronization and are coupled to each other through synaptic connections [7]. Synchronization signals are transmitted from the SCN clock to the peripheral clock through neural and hormonal stimulation to achieve a coherent rhythm throughout the organism [8, 9].

In humans, the physiological and behavioral circadian rhythms have evolved to form the biological clock that supports the occurrence of wakefulness during the daytime and promotes sleep during the night. Various aspects such as body temperature, metabolism, immune response, etc., exhibit circadian rhythms, and sleep is also a rhythmic behavior. Therefore, disruption of the circadian rhythm can directly impact sleep, and research has shown that light exposure can directly promote wakefulness and reduce sleep propensity [10]. Disturbances in the circadian rhythm can lead to various negative consequences, one of which is Circadian Rhythm Sleep-Wake Disorders (CRSWD). CRSWD is a distinct category of sleep disorders (sleep disorders mainly include

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insomnia, circadian rhythm disorders, sleep-disordered breathing, hypersomnia/narcolepsy, parasomnias, and restless legs syndrome/periodic limb movement disorder [11]). According to the classification criteria of ICSD-3, CRSWDs can generally be divided into two categories: one resulting from changes in the endogenous circadian rhythm system and the other due to a mismatch between sleep-wake time and internal circadian rhythm caused by environmental changes [12]. Melatonin, as an endogenous circadian factor that promotes sleep, when used at the appropriate time for treatment, can regulate the sleep-wake cycle [13]. In a placebo-controlled double-blind crossover trial involving 30 patients, it was observed that when patients took 5 mg of melatonin, their endogenous melatonin secretion occurred 1.5 h earlier. PSG analysis indicated that these patients showed advances and shortened sleep onset latency during the treatment period, without significant changes in sleep architecture. Furthermore, compared to before treatment, patients exhibited increased alertness upon waking in the morning and significant improvements in guality of life [14].

The core clock of mammalian circadian rhythms consists of a group of conservative transcription factors, in which the core is the key regulatory transcription activator circadian locomotor output cycles protein kaput (CLOCK), brain and muscle ARNT-like 1 (BMAL1) and neuronal PAS domain-containing protein 2 (neuronal PAS2) [1]. They form heterodimer complex CLOCK-BMAL1 and neuron PAS2-BMAL2 and they act on E-box element on target genes [1]. These target genes include circadian rhythm transcription factors period circadian protein homologue 1 (PER1) and PER2, cryptochrome 1 (CRY1) and CRY2, REV-ERB α (NR1D2), etc., all of which act as an inhibitor to the core clock [1].

Transcription of *Per* and *Cry* genes is induced by the CLOCK/ BMAL1 complex. After their proteins being synthesized, they accumulate in the cytoplasm and form heterodimers that shuttle between the nucleus and cytoplasm [15]. When the level of PER/ CRY complexes in the nucleus is sufficiently high, they inhibit the transcription of E-box genes (including their own genes) via

blocking CLOCK/BMAL1-mediated transcription [3]. Gene mutations that change the phosphorylation state, mobility, or degradation of PER and CRY proteins may affect the clock speed [16, 17]. PER proteins are subjected to successive phosphorylation events of multiple residues by CK1 δ and CK1 ϵ and by CK2 [17–25], leading to the change of their susceptibility to proteasomal degradation mediated by E3 ubiquitin ligase β -TrCP [26, 27]. Whether phosphorylation promotes protein stability or degradation depends not only on the modified residues, but also on the subtle differences in the kinases responsible [3]. For example, phosphorylation of CK16 selective splicing variants has an inverse effect on the stability of PER2 and the length of the circadian rhythm cycle [28]. In addition, the length of the circadian rhythm cycle is closely related to CRY protein abundance [29-36]. Phosphorylated CRY1 and CRY2 become targets for E3 ubiquitin ligases FBXL3 and FBXL21, while JMJD5 can promote CRY1-FBXL3 complexes to target proteasomes, thereby regulating the length of the circadian rhythm cycle via affecting the degradation of CRY proteins [37-42].

Nuclear receptors REV-ERB α and REV-ERB β are not only key components of the molecular circadian clock, but also major regulators of metabolism and mood [43–45]. The retinoid-related orphan receptor (ROR) nuclear receptor drives the expression of BMAL1 in the feedforward loop [1]. REV-ERB α inhibits *Bmal1* gene expression by binding to the ROR response elements (RRE) located in these gene promoters, and to a lesser extent inhibits *Clock* gene expression [46–49]. ROR α shares DNA binding sites with REV-ERBs [1], and when REV-ERB α inhibits the transcription of *Bmal1*, the ROR α activates it [50]. Pharmacological activation of ROR α and ROR γ , which are the antagonists of REV-ERB α and REV-ERB β in the molecular clock, can enhance rhythmic oscillations [51, 52], thereby demonstrating therapeutic potential in mouse models of Alzheimer's disease, Parkinson's disease, depression, and obesity [51, 53–55] (Fig. 1).

Circadian rhythms are involved in regulating almost every kind of physiological activity, including mood regulation, cell cycle control, metabolism, and immune responses. In some pathological



Fig. 1 Circadian rhythm mechanism. The master clock affecting the body's circadian rhythm is located in the hypothalamic suprachiasmatic nucleus (SCN), and the synchronization signal is transmitted from the SCN clock to the peripheral clock. The heterodimer complex CLOCK/ BMAL1 acts on E-box elements on target genes. After synthesis, PER and CRY accumulate in the cytoplasm and form heterodimers that shuttle between the nucleus and cytoplasm. After phosphorylation under the action of CK1 δ and CK1 ε , they inhibit the transcription of E-box genes by blocking CLOCK/BMAL1-mediated transcription; REV-ERB α inhibits the expression of *Bmal1* by binding to the ROR responsive element (RRE) in the promoter, while ROR α has the opposite activation effect.

conditions, the expression of clock genes also changed. Disruption of circadian rhythm may lead to abnormal cell proliferation, increased gene mutation, and resistance to apoptosis [56, 57], thus participating in the occurrence and development of a variety of diseases, including metabolic, cardiovascular, psychiatric diseases, and cancer. This paper systematically summarizes the molecular mechanism of circadian rhythm. The effects of relevant molecules on the immune system and cancer development; it will help open up new ideas for the treatment of certain diseases from the perspective of circadian rhythm.

EFFECT OF CIRCADIAN RHYTHMS ON IMMUNE CELLS Macrophages

Macrophages are one of the important innate immune cells, which induce a series of signal cascade reactions through Toll-like receptors (TLRs) on the cell surface to convert cells into immune active state [58]. A study found that the death caused by *E. coli* endotoxin changed with the injection time, indicating that the TLR4 signal was under the control of circadian rhythms [59]. In addition, in the TLR9-dependent septicemia model, stronger lethality was observed on the day when TLR9 expression was highest in spleen macrophages and B cells [60]. The expression of *Tlr2* and *Tlr6* mRNA in spleen macrophages also showed rhythm [60]. Therefore, the circadian rhythms of TLR and its downstream signal pathways may be one of the important mechanisms guiding circadian inflammation.

Macrophages can polarize into pro-inflammatory M1 phenotype with lipopolysaccharide (LPS) or anti-inflammatory M2 phenotype with IL-4, altering glycolysis and oxidative phosphorylation [61, 62]. BMAL1 inhibits septicemia development by affecting glycolysis metabolism. Loss of BMAL1 in macrophages reduces glycolysis control and increases infective shock mediated by PD-L1/T cells [63]. Macrophages lacking clock genes Per1 and Per2 exhibit a pro-inflammatory phenotype similar to M1 [64], while the hormone melatonin promotes M2 phenotype production via RORa and metabolite signals [65, 66]. Proteins Nfil3 and Dbp from circadian rhythm genes competitively bind to the II12b promoter, inhibiting and enhancing expression, respectively [63, 67]. The oscillation of these circadian rhythm proteins provides a rhythmic response to LPS [63].

Macrophages secrete cytokines and chemokines to participate in inflammation. REV-ERBa inhibits macrophage inflammation through various mechanisms [63]. The lack of REV-ERBa increases inflammation in alveolar macrophages [68], while REV-ERBa agonists inhibit inflammatory gene expression in macrophages [69]. REV-ERBa also regulates inflammatory markers in microglia [70] and colitis models [70, 71]. It directly regulates NIrp3 mRNA in a hepatitis model [72]. The PER/CRY complex is another mechanism for inhibiting inflammatory mediators in macrophages. Deletion of Per1 increases Ccr2 expression and migration ability [73], while deletion of Per2 increases TNFa and IL-12 production [74]. CRY negatively regulates the cAMP-PKA-NF-KB pathway [75]. Its deficiency leads to upregulation of II6, Tnfa, and inos. Studies targeting REV-ERBa found that GSK4112 inhibited NLRP3 expression and IL-1ß production in an LPS-induced inflammation model [76]. Disruption of circadian rhythm and KLF4 downregulation affect macrophage function in aging mice [77], highlighting the regulatory role of circadian rhythm.

Circadian rhythm can affect the activation of toll-like receptor (TLRs) signaling pathway on the surface of macrophages, regulate the immune activity and inflammatory response of macrophages, and also affect the polarization state and metabolic pathway selection of macrophages, thereby regulating their ability to participate in inflammatory response and immune function. This interaction may be one of the important mechanisms guiding circadian-related inflammation.

Dendritic cells

Dendritic cells (DCs) are antigen-presenting cells whose activity is influenced by circadian rhythm. In Rev-erba KO or Rev-erb β KO mouse bone marrow, DCs extracted show increased expression of MHCII and CD86, indicating maturity [78]. Conversely, the presence of REV-ERB agonist leads to the opposite effect. Defective migration of Bmal1 KO DCs into the spleen was observed, resulting in reduced CD8 T cell activation [79]. In addition, DCs exhibit stronger migration from skin to lymphatic vessels during the day compared to nighttime, and the deletion of Bmal1 gene eliminates this rhythmic migration pattern [80].

DCs also play a vital role in anti-tumor immunity. RNA sequencing analysis reveals that clock genes and clock control genes are expressed rhythmically in CD11c⁺MHCll^{hi} migrating DC subsets collected after tumor transplantation or under sham conditions [81]. The rhythmic production of DC costimulatory factors may contribute to the rhythmic activation of CD8⁺ T cells. The expression of CD80 in different CD11c⁺ subgroups is timedependent and controlled by the circadian clock mechanism. Deletion of BMAL1 eliminates the time difference in CD80 expression [81]. The proliferation of OT-I CD8⁺ T cells is dependent on the rhythmic phase of DCs. BMDCs with BMAL1 deficiency fail to induce rhythmic proliferation of OT-I CD8⁺ T cells. Treatment with anti-CD80 antibody eliminates the time difference in OT-I CD8⁺ T cell proliferation, confirming the correlation between CD80 and the rhythmic response of CD8⁺ T cells [81]. Chromatin Immunoprecipitation (ChIP) confirms the rhythmic binding of BMAL1 to the promoter region of the CD80 gene, indicating that CD80 is directly controlled by the clock gene BMAL1 [81].

Circadian rhythms influence the activity and function of dendritic cells (DCs), including maturity, antigen presentation, and migration. Circadian mechanisms also regulate the role of DCs in anti-tumor immunity, including the rhythmic expression of clock genes and related factors and the production of factors associated with the activation of CD8⁺ T cells. CD80 plays an important role in the circadian rhythm controlled CD8⁺ T cell response and is directly transcriptionally controlled by the clock.

B lymphocytes

The relationship between biological clock genes and lymphocyte development has been well established. Deficiency of BMAL1 negatively affects B cell development, leading to a decrease in their number in the blood and spleen of animals with knocked out BMAL1 gene [82]. However, adoptive transfer experiments have shown that it is not the internal clock of B cells but the loss of Bmal1 in the bone marrow microenvironment that hinders B cell differentiation [82]. On the other hand, mice with B-cell-specific Bmal1 deficiency do not exhibit any changes in B cell differentiation or function [83], suggesting that the influence of clock genes on B cell development is dependent on the microenvironment.

Some research has found that lymphocytes, including B cells, in mouse lymph nodes exhibit circadian rhythm oscillation. The highest number of lymphocytes is observed at the beginning of the active period (nighttime for mice). This rhythmic oscillation is due to the rhythmic homing of cells from blood into lymph nodes and their rhythmic egress from lymph nodes to efferent lymph vessels [84-86]. C-C-chemokine receptor 7 (CCR7) is a key regulatory factor that shows diurnal oscillation in both T cells and B cells [84]. In addition, the migration of B cells to lymph nodes also relies on other receptors, such as C-X-C-chemokine receptor 4 (CXCR4) and CXCR5 [87, 88]. The expression levels of CCR7 and CCL21 peak before and after the onset of nocturnal disease, leading to the recruitment of B cells. This process involves the microenvironment and lymphocytes [84]. Sphingosine 1-phosphate receptor 1 (S1P1) plays a regulatory role in the rhythmic export of cells to efferent lymph vessels [84]. Its rhythmic expression promotes the egress of lymphocytes from lymph

nodes in response to the chemotactic lipid sphingosine 1-phosphate [89].

Circadian rhythm disorders from irregular work and rest can damage B cell function, affecting the immune system. A study discovered that day-night shift rotation (DNSR) reduces nurses' IL-10 B cell (B10 cell) immunosuppressive ability and impairs Tr1 production in B10 cells [90]. Following DNSR, although B10 cell numbers remain unchanged, IL-10 levels decrease. RNA sequencing reveals CLOCK as one of B10 cells' most highly expressed genes post-DNSR. CLOCK levels correlate negatively with IL-10 mRNA levels. In addition, bB10 cells (pre-DNSR B10 cells) inhibit CD4⁺ T cell proliferation and induce Tr1 cells, but this immunoregulatory ability weakens after DNSR [90].

At present, the research on the connection between circadian rhythm and B lymphocytes is still little, and the regulation and influence of circadian rhythm on B cells need to be further explored.

T lymphocytes

 $CD8^+$ T cells. $CD8^+$ T cells are one of the important cells that participate in adaptive immune response, and they eliminate intracellular infection, control chronic infection, and eliminate tumor by producing cytokines (such as IL-2, IFN- γ , and TNF- α) and cytotoxic molecules (perforin and granzyme) [91]. A study confirmed the expression of *Per1* in CD8⁺ T cells of mouse spleen and the continuous rhythmic expression of PER2 protein in vitro-cultured CD8⁺ T cells [79, 83], indicating that CD8⁺ T cells have endogenous and cellular autonomic circadian clock.

T cells exhibit rhythmic expression of chemokine receptors and surface molecules, leading to migratory patterns and daily oscillations in organs. Cortisol regulates CXCR4 expression on T cells, impacting migration to CXCL2-expressing sites [92, 93], thus regulating Naive T cell levels in human blood. Naive T cells peak at night and are low during the day. Human effector CD8⁺ T cells peak during the day under catecholamine control [93]. In mice, $CD4^+$ and $CD8^+$ T cells are highest at the start of the day (ZT4-5) and lowest at the start of the night (ZT16) [94, 95]. Lymph nodes show an opposite rhythm, indicating T cell redistribution from blood to lymph nodes at night [84, 86]. This rhythmic migration is due to rhythmic CCR7 expression in CD8⁺ T cells and CCL21 (CCR7 ligand) in lymph nodes [91]. In addition, CD127 expression in CD8⁺ T cells induces CXCL4 (CXCL12 receptor) expression via IL-7, mediating daily oscillation in lymph organs [86].

Circadian rhythm mainly influences T cell proliferation. Stimulation of CD8⁺ T cells collected from mouse lymph nodes during a 24-h dark/dark cycle at night results in faster proliferation [96]. However, the proliferation rhythm is eliminated in T cells from mice with negative transcription factor CLOCK and deletion of the essential clock gene Bmal1 in mature CD8⁺T cells [79, 96]. A study using high-throughput RNA sequencing data from the Cancer Genome Atlas discovered that circadian rhythm disruption in tumors is associated with increased tumor-associated T cell dysfunction in human cancers. The study analyzed 716 samples from 29 different types of cancer. In most cancer types, the expression of a negative regulator of the circadian clock was downregulated, while a positive regulator, ARNTL2, was upregulated. This indicates that tumors lose the oscillating mRNA expression of the circadian clock gene. In addition, high expression of the clock gene correlated positively with levels of T cell dysfunction markers such as PD-1 and CTLA-4 in tumor samples. These findings suggest a connection between dysregulated clock gene expression and T cell dysfunction in aging and cancer, implying that circadian rhythms influence adaptive immune cell aging [97].

Disruption of circadian rhythms in tumors may lead to impaired function of tumor-associated T cells. In addition, T cells exhibit different chemotaxis and migration patterns under circadian rhythms, and the rhythmic production of cortisol and catechol amines regulates the migration and localization of T cells. The proliferation of T cells is also influenced by circadian rhythms, showing a faster rate of proliferation at night. These research findings suggest that circadian rhythms have important effects on the activity, migration, and proliferation of T cells, and disruption of circadian rhythms may impact immune responses, including tumor immune responses.

Helper T cells. Some studies have found that the specific circadian rhythm gene, rather than the main circadian rhythm regulator (BMAL1/CLOCK), regulates the specific CD4 Tn response [98]. In CD4 Tn, the circadian rhythm gene Per1 suppresses the expression of Th1 cytokines by inhibiting Akt/mTORC1 signal transduction [99], shifting the balance of helper T cell subsets towards Th2 and increasing the level of secreted antibodies during stress response [98].

Among T cells, Th17 is influenced by circadian rhythms. Th17 is a group of CD4⁺ helper T cells that play a vital role in immune response and are related to inflammatory reactions and autoimmune diseases. It secretes cytokines such as IL-17 and IL-22 and its differentiation and function depend on the transcription factor RORyt [100]. Genes *Nfil3* and *Rorc* play a key role in Th17 differentiation [101]. NFIL3 expression is regulated by REV-ERBa and REV-ERB β [102]. Expression levels of NFIL3 and RORyt were measured in a study, showing opposite patterns during day and night [101]. The study also found increased Th17 cells in the intestine and spleen of NFIL3 knockout mice (Nfil3^{-/-}) and inhibition of Th17 differentiation with NFIL3 overexpression [101].

Circadian rhythms have a significant impact on Th17 cells beyond CD4 Tn cells. A study found that circadian rhythm disorder (CRD) contributes to the development of allergic rhinitis (AR) by affecting Th2 and Th17 immune responses [103]. Levels of IL-4, IL-6, IL-13, and IL-17A significantly increased in CRD⁺AR patients, while IFN- γ levels decreased [103]. In addition, levels of Th2 and Th17 cells were upregulated, while Th1 and Treg cells were downregulated [103]. These findings suggest that CRD enhances the immune response mediated by Th2 and Th17 in AR by regulating Th1/Th2 and Th17/Treg immune balance [103]. Therefore, changes in clock activity can increase susceptibility to allergic diseases.

In CD4 T cells, the circadian rhythm gene Per1 inhibits Akt/ mTORC1 signaling, thereby suppressing the expression of Th1 cell cytokines and promoting the differentiation of CD4 T cells towards Th2 cells. This results in a tendency for CD4 T cells to produce more Th2 cells at specific time points and an increase in antibody levels under stress stimuli. In addition, the circadian rhythm also plays a significant role in Th17 cells in helper T cells. The circadian rhythm genes Nfil3 and Rorc play crucial roles in the differentiation of Th17 cells, and the quantity and functionality of Th17 cells may vary at different time points. Circadian rhythm disruption (CRD) can also affect the immune response of Th2 and Th17 cells, leading to excessive activation of these cells and increased susceptibility to allergic diseases. In summary, the circadian rhythm regulates the differentiation, function, and immune response of T cells by modulating specific circadian rhythm genes and signaling pathways, thus influencing the balance and function of the immune system.

NK cells

Natural killer (NK) cells are innate immune cells that directly eliminate tumor cells without activation. They release granzyme B, perforin, and pro-inflammatory cytokines, such as TNF- α [104, 105]. NK cells also regulate other white blood cells, including macrophages, T cells, and dendritic cells (DCs), through cytokine release [106]. Circadian rhythms affect NK cell function, including cytokine release and cytolytic factors [107], but their impact on NK cell development and function remains unclear.

A recent study investigated the impairment of NK cell function and potential mechanisms due to circadian rhythm disorders [108]. Using a chronic shift-lag mouse model, researchers observed decreased NK cell numbers in the spleen and lungs. They also found an increase in aging NK cells (CD27⁻CD11b⁺) and a decrease in functional NK cells (CD27⁺CD11b⁺) in the bone marrow and spleen. These findings suggest that circadian rhythm disorders promote NK cell apoptosis and aging, affecting their quantity and function [108]. In addition, disrupted circadian rhythms led to changes in NK cell receptor expression, with increased immature receptors (CD117 and CD127) and decreased functional receptors (Ly49D, Ly49G2, and Ly49H) [108].

Circadian rhythm disorder significantly reduces the secretion of CD107a and IFN-y by NK cells, impairing their ability to clear MHC-I deficient tumor cells and B16 melanoma cells [108]. IL-15 signal transduction is crucial for NK cell survival and homeostasis in the peripheral immune environment [109]. CD122 plays a key role in maintaining IL-15 stimulation [110]. In NK cells with circadian rhythm disorders, the expression of Eomes and T-bet, important transcription factors that regulate NK cell development and function, is altered [108]. Eomes affects NK cell development and function by transcriptionally regulating CD122, and its expression is significantly reduced in NK cells from mice with chronic shift-lag. Blocking CD122 abolishes the inhibitory effect of chronic shift-lag on CD107a and IFN-y secretion by NK cells [108], indicating that circadian rhythms indirectly affect IL-15 levels through the regulation of Eomes and CD122 expression, thereby impacting NK cell immune monitoring ability (Fig. 2).

In general, disruption of the circadian rhythm has a negative impact on NK cell function. Research has found that circadian disruption leads to a decrease in the number of NK cells, an increase in aging NK cells, and affects the expression of NK cell receptors. In addition, circadian disruption reduces the immune surveillance ability of NK cells and their ability to clear tumor cells. These effects may be achieved through the regulation of key transcription factors and signaling pathways. Further research is helpful in gaining a deeper understanding of the relationship between the circadian rhythm and the development and function of NK cells.

THE SLEEP-WAKE CYCLE IS INVOLVED IN INFLUENCING IMMUNE SYSTEM FUNCTION AND DNA DAMAGE REPAIR, AND IS REGULATED BY INFLAMMATION AND CYTOKINE FEEDBACK

As one of the three major cycles that affect mammalian physiological activity and function, the sleep-wake cycle plays an important role in maintaining normal physiological functions and ensuring normal gene expression. Especially for the immune system, sleep, as a pathway for specifically regulating the endocrine and autonomic nervous systems, should systematically control immune functions [111, 112]. Human studies have confirmed that regular sleep has a promoting effect on the primary immune response [113, 114], so maintaining a regular sleep-wake cycle can promote the formation of a good immune system in the body.

Sleep affects the production of cytokines and participates in the regulation of inflammation

Sleep affects immune status by regulating the secretion of cytokines by T cells. The cytokine balance between helper T cell 1 (Th1) and helper T cell 2 (Th2) is crucial for immune response. Th1 cells release interferon- γ (IFN- γ), interleukin-2 (IL-2), and tumor necrosis factor- α (TNF- α), promoting cellular (type 1) reactions against intracellular pathogens. Th2 immunity (IL-4, IL-5, IL-10, and IL-13) stimulates humoral (type 2) defense against extracellular pathogens [115]. Nocturnal sleep has been reported to benefit the transition to Th1-mediated immune defense. IFN- γ and IL-10 production in vitro exhibit a circadian rhythm, with a peak at around 03:00 during nighttime sleep [116, 117]. Sleep, particularly in the early stages dominated by slow-wave sleep (SWS), inhibits glucocorticoid release and promotes growth hormone (GH) and prolactin release [118-120]. These factors shift the Th1/Th2 balance towards Th1 advantage, promoting cell-mediated immune responses against intracellular pathogens [121, 122]. Another study confirmed that nocturnal sleep inhibits Th2 cell production compared to wakefulness, while the effect on Th1 cells remains unclear. Thus, sleep induces a shift towards Th1 dominance [115]. Inhibition of cortisol release and increased melatonin levels at night may contribute to Th1 dominance [116, 123]. However, this effect is limited to early sleep, mainly SWS, and reversed during late sleep dominated by rapid eye movement (REM) sleep, with lower CD4⁺ cell ratio of IFN-y/IL-4 during sleep compared to wakefulness [115]. Elderly individuals with sleep and SWS disorders show reduced GH and prolactin release, leading to Th2 dominance [124, 125].

Besides helper T cells, sleep also decreases TNF- α production by T suppressor/cytotoxic (CD8⁺) cells [115]. This decrease may be related to effector CD8⁺ cells and memory T cells rather than initial CD8⁺ cells, reflecting changes in migration to extravascular tissues [126, 127]. However, the role of cell recycling in sleep-mediated immune function remains unclear.

The impact of sleep on the immune system is mainly reflected in cytokine balance and the number of immune cells. Sleep can promote the activation of Th1 cells, increase the production of IFN-y and other related cytokines, thereby enhancing the ability to combat intracellular viruses and bacteria. At the same time, sleep can inhibit the release of cortisol, promote the release of growth hormone and prolactin. These factors work together to tilt the Th1/Th2 cytokine balance towards Th1 dominance, thereby enhancing cell-mediated immune responses. In addition, sleep can also suppress the production of Th2 cells, reduce the production of IL-4 and other related cytokines by CD4⁺ cells, further strengthening the Th1 dominance. Furthermore, sleep can also reduce the production of TNF- α by CD8⁺ cells. Although the specific mechanisms are not fully understood, the effects of sleep on the immune system primarily occur during the early stages of sleep (dominated by slow-wave sleep), while rapid eye movement (REM) sleep may have opposite effects. In elderly individuals, sleep and slow-wave sleep disorders lead to a reduction in the release of GH and prolactin, and the cytokine balance tilts towards Th2 dominance. In conclusion, sufficient sleep helps maintain the normal function of the immune system, especially enhancing cellmediated immune responses. However, further research is still needed to understand the impact of sleep on immune function.

Peripheral blood and intestinal inflammation are regulated by clock genes. Inflammation is linked to sleep, with various inflammatory cytokines involved in the development of cardiovascular diseases, diabetes, and other conditions. Sleep has an impact on the activities of these cytokines [128]. Epidemiological data suggest that insufficient sleep can increase the risk and mortality of chronic diseases due to its influence on the inflammatory mechanism [129]. Experimental sleep deprivation studies have reported alterations in immune response, leading to elevated levels of inflammatory markers such as IL-6, TNF-α, and C-reactive protein [130–132]. The cellular origin of pro-inflammatory cytokine activity remains unclear, although monocytes are considered the main contributors in peripheral blood [128]. TLR4-mediated lipopolysaccharide ligation has been shown to significantly increase IL-6 and TNF-α production in peripheral blood monocytes following sleep deprivation [128]. Moreover, insufficient sleep can induce changes in the expression of pro-inflammatory cytokines, the circulating clock gene PER1, immediate early response genes, as well as signal transduction mediators and growth factor-related genes. Structural functional bioinformatics analysis has revealed



Fig. 2 The effect of circadian rhythm on immune cells. Circadian rhythm can influence immune cell function and response by modulating specific genes and signaling pathways, thereby affecting the balance and functionality of the entire immune system. In macrophages, circadian rhythm can affect Toll-like receptor activation and downstream signaling pathways, regulating immune activity and inflammatory response. Additionally, circadian rhythm can impact macrophage polarization, metabolic pathway selection, and other factors, thus affecting their ability to participate in inflammation and immune function. For dendritic cells, circadian rhythm can affect their maturation, antigen presentation, and migration functions. As for B cells, their development and migration can be influenced by circadian rhythm, but the underlying mechanisms require further research. In T cells, circadian rhythm can influence the expression of key genes such as NFIL3 and ROR_Yt, affecting Th17 differentiation and function. Finally, circadian rhythm can also affect the number and function of NK cells, thus influencing their ability to clear tumor cells.

the involvement of multiple signaling pathways, including the nuclear factor κB inflammatory signaling system and classic hormone and growth factor response pathways, in the transcriptional response of white blood cells to sleep deprivation [128].

The biological clock plays a crucial role in regulating intestinal immunity and maintaining intestinal barrier function. For instance, knockout of BMAL1 disrupts the homeostasis of gut group 3 innate lymphoid cells (ILC3), leading to impaired intestinal epithelial reactivity, microbiome imbalance, increased susceptibility to intestinal infection, and lipid metabolism disorders [133]. BMAL1 knockout mice also exhibit reduced small intestinal crypts and impaired intestinal regeneration after radiation-induced gastrointestinal syndrome [134], highlighting the importance of BMAL1 in intestinal cell proliferation. Overexpression of CRY1 inhibits vascular inflammation induced by sleep deprivation (SD), possibly through the NF- κ B and cyclic adenosine monophosphate/ protein kinase A (cAMP/PKA) pathways in vivo [135]. Acute sleep deprivation (ASD) commonly affects shift workers, causing lethargy and persistent fatigue. ASD contributes to inflammation by influencing the intestinal microbiota. In elderly individuals with insomnia, the impact of physical activity on sleep disorders can be regulated by the gut microbiota's primary metabolites. The genus Monoglobus shows a negative correlation with physical activity and sleep latency, while exhibiting a positive correlation with sleep

efficiency [136]. Disruption of circadian rhythm leads to decreased abundance of specific bacteria (e.g., Lactobacillus and Ruminococcus) along with downregulation of BMAL1 [137]. Increased Candidatus Arthromitus in the SD group may contribute to colitis, intestinal barrier dysfunction, and systemic inflammation [138]. Thus, circadian rhythm and microbiota composition play crucial roles in maintaining intestinal barrier function. Adequate sleep and self-regulation of the microbiota might have protective effects against disease and help restore the intestinal epithelial barrier, thereby improving inflammation [138].

In summary, it can be found that sleep regulates cytokine secretion and intestinal microflora through various pathways, affecting the inflammatory state of the body, thereby participating in the development of various inflammatory diseases. Breaking the regular sleep-wake cycle may increase the secretion of proinflammatory cytokines in the peripheral blood, thus increasing the risk of cardiovascular disease, diabetes, etc. In addition, the downregulation of specific flora abundance and clock gene expression caused by acute sleep deprivation may lead to colitis, intestinal barrier dysfunction, and other diseases.

Sleep has a significant impact on the immune system and inflammatory state. Lack of sleep can lead to increased inflammatory response, increased risk of chronic diseases, and mortality rates. Experimental sleep deprivation can alter immune responses and increase circulating levels of inflammatory markers. Sleep deprivation also affects changes in the expression of proinflammatory cytokines and signaling pathways. The biological clock plays a crucial role in intestinal immunity and barrier function. Knocking out the BMAL1 gene disrupts the homeostasis of intestinal group 3 innate lymphoid cells, leading to impaired intestinal responsiveness and dysbiosis. Adequate sleep and selfregulation of the gut microbiota can protect the immune system, improve inflammatory status, and restore intestinal barrier function. Therefore, sleep has a significant impact on the body's inflammatory state and the development of inflammatory diseases, and disrupting the circadian rhythm may increase the risk of various diseases.

Sleep deprivation mediates the occurrence of cancer via affecting DNA damage repair and oxidative stress

Circadian rhythm disorders caused by chronic imbalances between behavioral rhythms and external light/dark cycles are common [139] and have been proven to be causes of various cancers [140] and chronic health problems [141]. Increasing evidence suggests that circadian rhythms affect multiple pathways regulating cancer outcomes, including cell cycle checkpoints, apoptosis, cell proliferation, DNA repair, and inflammation [142–150].

Circadian rhythm disorders are prevalent among night shift workers [151], leading to severe disruption of human biomolecular pathways during and after night shifts [152–155], thus increasing the cancer risk [156]. Rodent studies confirm this view, showing that changes in circadian rhythms can disrupt cell cycles, alter metabolism, promote tumor development, induce tumor cell growth, and trigger tumor-related immune cell remodeling under simulated shift work and chronic jet lag conditions [157–160].

A study analyzed volunteers subjected to simulated day or night shift schedules, which resulted in circadian rhythm disorders, and collected blood samples to study their impact on endogenous biological processes [139]. The findings revealed impaired effectiveness of repairing DNA damage caused by endogenous sources and exogenous irradiation in white blood cells obtained under night shift conditions. Moreover, compared to the simulated night shift group, night infrared (IR) exposure led to more DNA damage in the white blood cells of the simulated night shift group [139]. In addition, it reports for the first time the circadian rhythm imbalance of cell cycle and DNA repair regulatory factors in human leukocytes after night shifts. Key genes involved in cell cycle checkpoint activation and DNA repair were identified, including ATR, CDK4, CDKN1C, TP53, WEE1, ERCC6, TP53, RPA3, XPA, H2AFX, PARP1, and RAD50. Disruption of the circadian rhythm of these genes' expression suggests a direct link between cellular circadian rhythm mechanisms and the carcinogenic risk of night shift work [139].

Severe sleep deprivation can have even more serious consequences, leading to premature death in model organisms such as dogs, rats, cockroaches, and flies [161-164]. Insufficient sleep impairs brain function [165-167], and some propose that sleep's function is to prevent oxidative stress in the brain [168]. Studies have reported changes in antioxidant response during sleep deprivation [169–173]. Insufficient sleep can result in ROS accumulation due to increased production, decreased elimination, or both [174]. While most ROS is generated during mitochondrial oxygen-dependent ATP synthesis [175], it is also produced as a byproduct of oxidative protein folding [176]. Lack of sleep can induce endoplasmic reticulum stress, leading to oxidative stress and increased demand for protein folding, thereby increasing ROS production [177-179]. However, many studies have not detected significant oxidative damage in the brain during sleep deprivation [180–183], potentially due to effective activation of antioxidants and cellular protective mechanisms [170, 184-188]. Notably, gastrointestinal diseases are often associated with sleep restriction and abnormal ROS levels, which may contribute to tumor development through mechanisms like DNA damage and inflammation [189-196] (Fig. 3).

Factors that primarily contribute to the disruption of circadian rhythm include insufficient sleep, night shift work, and sleep deprivation. These factors can lead to disturbances in DNA damage repair and oxidative stress, increasing the risk of cancer. Sleep deficiency can interfere with the circadian rhythm, affecting multiple pathways such as cell cycle, apoptosis, cell proliferation, DNA repair, and inflammation, thereby influencing cancer outcomes. Night shift workers commonly experience circadian rhythm disruption, which increases their susceptibility to cancer. Animal studies have also shown that alterations in circadian rhythm can disrupt cell cycle, alter metabolism, promote tumor development, and induce tumor cell growth and immune cell reshaping under simulated night shift and long-term jet lag conditions. Research has also found circadian rhythm disruption in genes involved in DNA repair pathways among volunteers following a simulated night shift schedule. The effectiveness of the DNA damage repair process is compromised in white blood cells obtained during night shift work. The circadian rhythm of cell cycle and DNA repair regulatory factors in human leukocytes is also imbalanced after night shifts, and these genes are crucial for identifying and repairing DNA damage. Severe sleep deprivation may also lead to premature death in model organisms, possibly due to impaired brain function. During sleep deprivation, changes occur in the brain's antioxidant response, and insufficient sleep can result in the accumulation of reactive oxygen species (ROS), leading to oxidative stress. Considering the pathological implications associated with sleep restriction, such as gastrointestinal disorders, abnormal levels of ROS may contribute to tumorigenesis through various mechanisms.

Inflammation, cytokines, and immune cells can provide feedback to regulate sleep

Sleep and immunity have a bidirectional relationship. Sleep affects the immune system, and changes in the immune system can also affect sleep. Diseases characterized by low-level inflammation have been identified, including degenerative diseases, cardiovascular diseases, metabolic disorders, and chronic pain [197–199]. Inflammatory disorders are considered a potential mechanism linking sleep time and mortality [200, 201].

In a 7-year study of over 2500 elderly men, short sleep time (5 h) was associated with an increased risk of all-cause mortality due to

Fig. 3 The sleep-wake cycle is involved in affecting immune system function and DNA damage repair. Nighttime sleep, particularly in the early stages characterized by slow-wave sleep (SWS), promotes a shift towards Th1-mediated immune defense by inhibiting glucocorticoid release and promoting the release of growth hormone (GH) and prolactin. Sleep also inhibits Th2 cell production, further reinforcing Th1 dominance. Circadian rhythms regulate the secretion of cytokines and gut microbiota, significantly impacting the body's inflammatory state. Sleep deprivation leads to abnormal cytokine secretion, altering the inflammatory state. Experimental sleep deprivation studies have shown increased immune response and elevated levels of inflammatory markers such as IL-6, TNF- α , and C-reactive protein in circulation. Monocytes are considered a primary factor in peripheral blood inflammation. Additionally, sleep deprivation affects the expression of inflammatory cytokines, circadian clock gene PER1, signaling molecules, and growth factor-related genes. Sleep is crucial for maintaining normal immune function, particularly enhancing cell-mediated immune responses. Furthermore, sleep deprivation disrupts circadian rhythms, impairs DNA damage repair, and increases the risk of cancer occurrence. Night shift workers often experience disrupted circadian rhythms, which interfere with cellular processes involved in DNA repair, cell cycle regulation, and inflammation, thereby increasing cancer risk. Studies have shown that sleep deprivation impairs the effective repair of DNA damage from both endogenous and exogenous sources. Moreover, disruptions in the expression of key genes involved in cell cycle checkpoints and DNA repair further indicate a direct link between circadian rhythm disturbances and the carcinogenic risk associated with night shift work. Severe sleep deprivation may exacerbate these effects, potentially leading to premature death in model organisms. Insufficient sleep not only affects brain function but also disrupts the body's antioxidant response, leading to increased oxidative stress and DNA damage.

increased inflammatory burden (i.e., composite measurement of CRP, IL-6, TNF- α , sTNF-RII, IFN- γ levels); adjusting for the inflammatory burden significantly weakens this relationship, indicating that the inflammatory mechanism disrupts the relationship between sleep time and mortality [202]. Elevated CRP has been recognized as a marker for predicting cardiovascular risk [203], suggesting that inflammation may be the basis for the association between sleep deprivation and mortality, as well as the association between sleep deprivation and increased cardiovascular disease risk [204].

In most studies, inflammatory status is measured by levels of CRP or IL-6, and higher levels of CRP and IL-6 are associated with reduced sleep time [205]. The mediating effect of inflammation seems to be unique to short sleep duration, as other sleep characteristics such as sleep latency or nocturnal wakefulness are not associated with the burden of inflammation. Elevated inflammatory markers have also been reported in prolonged sleep (8 or 9 h) [206–210], although uncontrolled disease processes may

mediate this relationship. However, some studies have not found an association between inflammatory markers and sleep time [211–215], increasing the heterogeneity of research results.

Regarding inflammatory cytokines, poor subjective sleep quality has been associated with higher production of stimulating IL-1, TNF- α , and IL-6 in a cross-sectional study [213]. Recently, poor sleep quality in postmenopausal women has been associated with increased IL-6 responsiveness after psychological and social stress [216]. In terms of immune cells, an increase in white blood cell count was associated with lower sleep efficiency and higher nocturnal awakening time in women [217–220].

In summary, the bidirectional relationship between sleep and immunity is supported by research results. Inflammatory markers, cytokines, and immune cells are connected to sleep, and their dysregulation may be a potential mechanism for sleep disorders affecting disease risk [221]. Changes in sleep during illness, such as infection, may have implications for recovery [222]. EFFECT OF CIRCADIAN RHYTHM ON TUMORS AND THEIR MICROENVIRONMENT A regulatory mechanism exists between circadian rhythms and

cancer genes, which is relevant to tumor incidence and malignancy. Gene rhythms associated with metabolism and endocrine function significantly contribute to tumor development. Abnormal biological rhythm disrupts the body's innate and acquired immunity, promoting malignant tumor progression. Studies have shown that in the malignant thyroid nodule group, the expression level of CLOCK and BMAL1 was significantly higher, while the expression level of CRY2 was lower [223]. The infiltration of immune cells such as macrophages and neutrophils in renal clear cell carcinoma correlates with rhythmic changes in the expression of CLOCK-related components (BMAL1, PER, etc.) [224, 225]. CLOCK also participates in regulating the microglial content of glioblastoma stem cells (GSC) in glioblastoma (GBM) through transcriptional regulation of chemokines [226]. Single-cell transcriptomic analysis has revealed an association between circadian rhythm disturbance (CRD) and poor prognosis as well as drug resistance in lung adenocarcinoma. CRD score was significantly associated with poor prognosis and resistance to anti-cancer treatments, including chemotherapy and tyrosine kinase inhibitor (TKI) therapy, in LUAD patients [227]. Disruption of clock-related genes promotes colorectal cancer in a model of gut organoids, and loss of circadian rhythm was observed in patient-derived organoids [228].

The cell cycle and biological clock function as biological oscillators, and there is increasing evidence of molecular connections between these two oscillators using various coupling mechanisms, which are often misregulated in cancer cells [146]. The cell cycle consists of four stages: G1, S, G2, and M phases, with the G0 phase representing a resting state when cells temporarily stop dividing. Key cell cycle molecules (cyclins) regulate cell cycle progression in a sequential and unidirectional manner [229], and the kinase activity of cyclin-CDK complexes triggers specific events at defined times in the cell cycle [230]. The biological clock acts as a biological oscillator with conceptual and mechanical similarities to the mechanisms controlling cell division. Both the cell cycle and biological clock exhibit sequential stages of transcription/translation, protein modification, and degradation [146]. Their interaction jointly regulates tumor cell activity. A deeper understanding of the molecular links between the cell cycle and circadian clock may serve as the foundation for designing innovative therapeutic strategies.

The tumor microenvironment (TME) significantly influences tumor development. TME usually consists of the extracellular matrix (ECM) and various cells including congenital myeloid cells [such as tumor-associated macrophages (TAM), dendritic cells, and myeloid suppressor cells (MDSC)], lymphocytes [such as T cells and natural killer (NK) cells], and endothelial cells [231]. Understanding the molecular aspects of the circadian clock in tumorigenesis and the symbiotic interaction between cancer cells and the TME is currently a major focus of cancer research [232]. Recent evidence indicates that circadian rhythm disturbances are associated with increased cancer risk and promote metastasis in multiple tumor types. The biological clock also plays a regulatory role in tumor growth and several cancer markers. Furthermore, studies have confirmed that the circadian rhythm regulates immune function, which is of great significance in cancer treatment since the immune system plays a crucial role in eliminating and suppressing malignant tumors.

Circadian rhythm participates in the development of tumors by affecting various kinds of cells

The concentration of immune-mediated hormones and cytokines fluctuates rhythmically throughout the day [148]. Any disturbance in the balance between anti-tumor immune cells and pro-tumor immunosuppressive cells may enable cancer cells to escape immune control, resulting in disease occurrence and development [233]. T cells and myeloid-derived suppressor cells (MDSCs) play a major role. $CD8^+$ T cells are responsible for selectively killing tumor cells [234], whereas MDSCs inhibit immune response and limit inflammation, reducing the ability of T cells to kill cancer cells [235]. External disturbance of circadian rhythm in mice tumors leads to faster tumor growth due to the interruption of internal circadian rhythm of cancer cells, amplifying their proliferation [236]. Interruption of central circadian rhythm promotes tumor growth by affecting non-cancer cell signaling, while MDSC accumulation inhibits tumor-specific immune response [236]. Adrenergic signaling pathway disruption increases MDSC accumulation, regulating hematopoietic stem cells and CXCL12 expression periodic fluctuation [237], and impacting cytokine expression and cytolytic function of NK cells [238].

TAM is the most significant subgroup in TME, influencing tumor growth and metastasis [239]. BMAL1 depletion in macrophages regulates pro-inflammatory cytokine production through HIF1a upregulation, ROS accumulation, and NRF2 downregulation [240, 241]. The balance between BMAL1 and HIF1a affects macrophage metabolism and anti-tumor immunity, and BMAL1 knockout increases septic mice mortality [242, 243]. Co-injection of cancer cells and Bmal1 KO macrophages promotes tumor growth with reduced CD8⁺ T cell infiltration [240]. Activation of REV-ERBa inhibits CCL2 and its downstream signals, suppressing macrophage adhesion and migration [244]. GAMs divide into M1 and M2 subtypes with opposite functions in glioma progression [245-247]. M2 subtype promotes tumor proliferation and metastasis by reducing BMAL1 expression in TAM and regulating tumorrelated protein expression and cell apoptosis [248]. Leukocyte migration, crucial in tumor development, is regulated by the biological clock [249]. Bmal1 deletion in endothelial cells eliminates time difference in migration-promoting molecule expression [250], while CLOCK overexpression increases leukocyte adhesion to endothelium via upregulating ICAM-1, VCAM-1, and other molecules [251]. CRY1 overexpression in endothelial cells inhibits inflammatory cytokines (IL-1β, IL-6, TNF-α), adhesion molecules (VCAM-1, ICAM-1, e-selectin), and NF-κB pathway activation, impairing monocyte adhesion [135].

The circadian clock regulates cancer cell products, TME, and their symbiotic interactions. Breast cancer and melanoma mouse models show that circadian rhythm disorder enhances cancer cell proliferation, spread, stemness, and metastasis, induces immunosuppressive TME by increasing TAM and regulatory T cell (Treg) proportion, and promotes differentiation of macrophages into an anti-inflammatory phenotype resulting in reduced infiltration of active CD8⁺ T cells [160, 252]. RORy activation inhibits tumor growth, prolongs survival by enhancing Th17 cell differentiation and function, and reducing Treg levels [253]. RORa activation maintains cholesterol metabolism balance in CD8⁺ T cells, enhancing their activity through weakening the NF-kB pathway [254].

There is molecular coupling between cell cycle and biological clock, and they are jointly involved in the occurrence and development of cancer

DNA replication and cell division require a large number of bioenergy components [255]. Their synthesis is regulated by a clock-dependent mechanism, indicating that the demand for cell cycle biosynthesis is controlled by the circadian rhythm of the metabolic cycle and environmental supply [256, 257]. Mice carrying core clock gene-specific ablation have shown the importance of functional clocks for the normal mitotic rhythm and progression of progenitor cells and/or stem cells in various cell types [258-262]. The circadian clock also plays a role in coordinating the regeneration process after inducing damage to the skin [263] and intestinal epithelium [261].

The circadian clock is not a mandatory component of the cell cycle but contributes to the timing of cellular events and adaptive mechanisms needed for optimal tissue function [146]. There is a significant relationship between the circadian clock and the cell cycle, with evidence of circadian rhythm changes in cyclin abundance and clock gene expression at specific cell cycle stages [264, 265]. The circadian clock controls the expression of CDK and cyclins such as CDK2, CDK4, and cyclin D1, as well as cell cycle inhibitors including Wee1, p21, p27, and p57 [266]. CLOCK-BMAL1 regulates the circadian expression of the G2/M inhibitor WEE1 [267], while the expression of p21 is regulated by antagonistic effects of ROR and REV-ERB proteins [268]. NONO, in association with PER protein, regulates the circadian rhythm regulation of p16 expression [263]. P53 directly binds to the Per2 promoter and inhibits CLOCK-BMAL1-mediated transcription [269].

Cell cycle checkpoints detect replication stress or DNA damage and delay cell cycle progression to repair or eliminate damaged cells. Circadian clock proteins interact with checkpoint reactive protein complexes to mediate DNA damage responses [146]. This coupling may protect cells from DNA damage caused by ultraviolet radiation by limiting replication during dark periods and optimizing repair mechanisms during the day [259, 270]. DNA damage can also regulate circadian rhythms and affect cell cycle progression [271].

Cell cycle disruption is a key event in carcinogenesis, and there is a correlation between cancer incidence rate and circadian rhythm disorder [272]. Both cell cycle genes and biological clock genes are incorrectly regulated in cancer cells [266]. Exploring the interaction between circadian rhythm and the cell cycle in cancer is of great significance. However, the causal relationship between sleep circadian rhythm disorders and cancer emergence and progression, the effectiveness of controlling cancer with the circadian clock, and the impact of cancer-related therapies on the biological clock remain unresolved [146].

Metabolism and multiple factors in tumor microenvironment are regulated by biological rhythm

Apart from that, tumor-induced signals such as granulocytemacrophage colony-stimulating factor (GM-CSF), granulocyte-CSF (G-CSF), macrophage-CSF (M-CSF), stem cell factor (SCF) and vascular endothelial growth factor (VEGF), along with inflammatory signals including IFN- γ , IL-1 β , IL-6, and TNF- α , are rhythmically expressed and released by specific cells [236]. Central circadian rhythm changes can increase the production of these signal factors in tumor cells. Knockout of clock proteins Cry1 and Cry2 leads to a constitutive increase in pro-inflammatory cytokines via NF- κ B and PKA signals, indicating direct regulation of cytokine expression by circadian genes [75]. Deletion of myeloid-specific circadian rhythm gene disrupts chemokine expression rhythm and Ly6Chi monocyte tissue recruitment [242]. Adhesion molecules and chemokines in endothelial cells oscillate in a circadian rhythm, affecting leukocyte adhesion and exosmosis [273].

Clock components regulate the expression of angiogenesis factors in cancer cells, such as HIF1a, ARNT, and VEGF, promoting tumor growth and metastasis [56, 274–276]. CLOCK knockout decreases angiogenesis factor expression in human colorectal cancer (CRC) cells, while CLOCK overexpression increases it [274]. Xenotransplantation tumors show fluctuating VEGF levels controlled by PER2 and CRY1 transcription in a circadian rhythm [277]. Clock genes not only regulate the circadian rhythm and transcription factor activity of cancer cells but also affect macrophage, neutrophil, dendritic cell, and lymphocyte infiltration [224, 225]. Clock gene mutations in cancer patients lead to genomic instability, T cell depletion, upregulation of immunosuppressive molecules, and immune escape [97, 278].

In a study on breast cancer, chronic jet lag (JL) was shown to impact its development [252]. JL mice had higher blood lipid levels and increased leptin and insulin resistance, which are related to weight gain, obesity, and type 2 diabetes [252, 279, 280]. Expression of Per2 and Cry2 clock genes decreased in primary tumors of JL mice, with significant changes in genes involved in light transduction in the tumors and mononuclear myeloma cells [252]. CRD-related enhancement of tumor genesis and metastasis involved the CXCL5-CXCR2 axis. CRD increased Cxcl5 expression, leading to increased infiltration of CXCR2⁺ myeloid cells (such as MDSCs) [252]. This promoted the accumulation of MDSCs, TAM, and TAN, creating an immunosuppressive microenvironment [281]. These cells directly inhibit T-cell response and suppress CD8 T-cell infiltration, impairing anti-tumor activity [281-284]. Inhibition of CXCR2 significantly reduced lung metastasis and bone diffusion in JL mice [252]. The CXCL12-CXCR4 axis also plays a crucial role in JL mouse cancer cell metastasis and

Table 1. Clock genes participate in the development of tumors.			
Clock gene	Cell	Effects	Ref.
BMAL1↓	Macrophage	↑ HIF1α, ROS; ↓ NRF2→ ↑ pro-inflammatory cytokines→ ↑ the mortality of septic mice	[71, 72]
	GAM (M2 subtype)	↑ tumor-related proteins; ↓ cell apoptosis→ ↑ tumor proliferation and metastasis	[81]
	Endothelial cell	Time difference in the expression of migration-promoting molecules was eliminated	[<mark>83</mark>]
CLOCK↑	Endothelial cell	↑ ICAM-1, VCAM-1→ ↑ the adhesion of leukocytes to endothelium	[84]
	CRC cells	↑ angiogenesis factors	[<mark>91</mark>]
CRY↓	Endothelial cell	↑ inflammatory cytokines, adhesion molecules, NF-κB pathway→ ↑ monocyte adhesion	[85]
	Tumor cell	↑ NF-κB, PKA→ ↑ pro-inflammatory cytokines	[32]
RORα↓	CD8 ⁺ T cell	\uparrow NF-κB pathway → ↓ CD8 ⁺ T cell activity	[89]
RORγ↓	Th17 cell, Treg	↓ differentiation and function of Th17 cells; ↑ the level of Tregs \rightarrow ↑ tumor growth	[88]
REV-ERBα↑	Macrophage	\downarrow CCL2 $\rightarrow \downarrow$ the adhesion and migration	[75]

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diffusion [252]. Its control of circadian rhythm and its involvement in immunosuppression and breast cancer metastasis have been extensively described [237, 250, 285–288] (Table 1).

SUMMARY

Circadian rhythm is a widespread physiological phenomenon in nature. It has been increasingly recognized for its impact on the body's physiology and pathology. Maintaining circadian rhythm is crucial for normal cellular function and metabolic pathways [1]. Disruption of circadian rhythms, such as through shift work, can alter clock gene expression and increase disease risk [63].

In pathological conditions like cancer and allergic diseases, abnormal clock gene expression affects the phenotype and function of immune cells, including macrophages [63], dendritic cells [81, 289], and lymphocytes [1, 90, 98, 102, 103]. It also influences cytokine and chemokine levels, contributing to the development of various diseases.

Circadian rhythm controls Toll-like receptor expression, polarization, and cytokine secretion in macrophages through BMAL1, REV-ERBa, and PER/CRY complex [63]. Disruption of Bmal1 causes migration defects in dendritic cells and affects CD80 expression timing, impacting CD8⁺ T cell proliferation [81, 289]. BMAL1 also regulates chemokine receptor secretion in B cells, while CLOCK negatively correlates with IL-10 levels in B10 cells, influencing CD4⁺ T cell and Tr1 cell production [1, 90]. Circadian oscillation of proliferation and migration has been observed in CD8⁺ T cells, indicating their endogenous circadian rhythm [91]. Furthermore, Th17-related genes Nfil3 and Rorc play a role in disease regulation through immune response mediation [102].

The sleep-wake cycle significantly influences mammalian physiological activities and functions. It affects cytokines, inflammation, DNA damage repair, and oxidative stress [115]. Sleep can promote the transition to Th1-mediated immune defense and inhibit cortisol release, leading to increased melatonin levels [115]. Lack of sleep alters pro-inflammatory cytokine expression and disrupts signal transduction pathways [128]. Clock genes also regulate intestinal inflammation by affecting the abundance of specific gut bacteria through BMAL1 downregulation [138]. Shift work-induced sleep-wake cycle disorders impact circadian rhythms in white blood cell cycle and DNA repair pathwayrelated genes [139]. This disruption induces endoplasmic reticulum stress, oxidative stress, and increased ROS production, potentially causing premature biological death [174]. Inflammation, cytokines, and immune cells reciprocally regulate sleep. Inflammatory mechanisms disrupt the relationship between sleep duration and mortality, potentially contributing to increased cardiovascular disease risk [221].

Circadian rhythm is associated with cancer incidence and malignancy. Rhythmic genes influence the expression of cancer genes, metabolism, and endocrine functions, contributing to the development of glioblastoma, breast cancer, and other tumors [243]. Clock gene mutations affect various cells in the tumor microenvironment, promoting immune escape, tumor growth, and altering the balance between anti-tumor immune cells and pro-tumor immunosuppressive cells. They also regulate TAM cell subsets' proliferation, migration, infiltration ability, Th17 and Treg levels, and CD8⁺ T cell numbers [243]. Molecular coupling exists between the cell cycle and the biological clock. Clock genes are expressed at specific stages of the cell cycle, resulting in rhythmic changes in cyclin abundance. Proteins encoded by rhythmic genes, such as BMAL1, CLOCK, PER, regulate cell cycle-related genes like Wee1, p21, p16, and p53. Circadian clock proteins directly interact with checkpoint reactive protein complexes, mediate DNA damage responses, and impact cancer development [146]. In addition, clock genes regulate cytokines, chemokines, and migration-promoting molecules produced by tumors, playing a role in immunosuppression and cancer metastasis [243]. Targeting

clock genes offers new avenues for cancer treatment, and various targets (e.g., CLK8 [290], mTOR [291]) and drugs (e.g., MLN4924 [292]) are under development with ongoing clinical trials.

Overall, circadian rhythms play a vital role in the development and functioning of immune cells. Current research has confirmed that various immune cell types are regulated by their own circadian rhythm genes as well as environmental factors during development. The distribution and functional status of these cells are also influenced by the expression of circadian rhythm-related products in the microenvironment. For the immune system as a whole, circadian rhythm acts like a control panel, leading to different responses to external stimuli (such as bacterial or viral infections, vaccination, etc.) at different times of the day. For example, receiving the COVID-19 vaccine in the morning may be more beneficial for patients' health [293]. In recent years, the relationship between energy metabolism and circadian rhythm has also been explored. Hormones closely linked to energy metabolism, such as insulin and glucocorticoids, exhibit circadian rhythm characteristics. These hormones significantly impact immune system function, suggesting that the connection between the immune system and circadian rhythm is not simply a direct regulation. In addition, the liver, as the metabolic hub of the human body, demonstrates distinct circadian rhythm patterns in glucose and lipid metabolism [294]. Another intriguing observation is the loss of control of circadian rhythmrelated genes in cancer, which has been demonstrated to have varying effects across different types of cancer [295]. Considering the unique cell cycle regulation and metabolic properties of tumor cells, as well as the intricate relationship among the immune system, energy metabolism, and circadian rhythm, we hypothesize that tumor cells actively modify their rhythmic characteristics to adapt to their specific metabolism and proliferation requirements. By inducing dysregulated rhythms through downstream molecular changes, they can manipulate relevant immune responses to evade attacks from the immune system. Therefore, restoring the circadian rhythm in tumor cells while protecting the immune system from the detrimental effects of circadian rhythm disruptions could potentially serve as a novel therapeutic strategy. Furthermore, monitoring circadian rhythmrelated molecules may prove to be an effective approach for cancer prediction and immune function evaluation.

DATA AVAILABILITY

Data sharing is not applicable to this article as no datasets were generated or analyzed during the current study. All data generated or analyzed during this study are included in this published article or available from the corresponding author on reasonable request.

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AUTHOR CONTRIBUTIONS

MW and YZ: Investigation, visualization, writing—original draft. ZG and FC: Validation, funding acquisition, supervision, writing—review & editing. LC: Conceptualization, project administration, funding acquisition, supervision, writing—review & editing.

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COMPETING INTERESTS

The authors declare no competing interests.

ADDITIONAL INFORMATION

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