## **EDITORIAL BOARD**

**Lida Anestidou, DVM, PhD** Program Officer, Institute for Laboratory Animal Medicine, The National Academies, Washington, DC

Kathryn Bayne, MS, PhD, DVM, DACLAM, CAAB

nior Director and Director of Pacific Rim Activities. AAALAC International, Waikoloa, HI

Joseph T. Bielitzki, MS, DVM Associate Director, Nanoscience Center, University of Central Florida, Orlando, FL

Cyndi Brown, DVM, DABVP (Avian Practice)
Ocean State Veterinary Specialists, Fact C

Joseph W. Carraway, DVM, MS

Director of Toxicology, NAMSA, Northwood, OH

Thomas M. Donnelly, DVM, DACLAM

The Kenneth S. Warren Institute, Ossining, NY

CEO, Biologist, Primate Products, Redwood City, CA

Robert F. Hoyt, Jr., DVM, MS, DACLAM Chief, Laboratory Animal Medicine and Surgery, National Heart Lung and Blood Institute, NIH, Bethesda, MD

Mary Lou James, BA, RLATG

Consultant, Regulatory Compliance, St. Louis, MO

Alicia Z. Karas, DVM, MS, DACVA

Assistant Professor, Anesthesia, Department of Clinical Sciences, Cummings School of Veterinary Medicine, Tufts University, North Grafton, MA

**Bruce W. Kennedy, MS, RLATG** Compliance Associate, Research and Sponsored Programs,

Cal Poly Pomona, Pomona, CA

C. Max Lang, DVM, DACLAM

Professor and Chairman, Department of Comparative Medicine, Milton S. Hershey Medical Center, Pennsylvania State University, Hershey, PA

Richard H. Latt, DVM, DACLAM Director, Animal Facilities, The Trudeau Institute, Saranac, NY

Nutritionist/Research Scientist

National Center for Toxicological Research, Jefferson, AR

Carol Cutler Linder, PhD
Assistant Professor of Biology, New Mexico Highlands University, Las Vegas, NM

John A. Maher, MS, MBA, CMAR, RLATG Senior Manager, Comparative Medicine, Pfizer, Pearl River, NY

Jörg Mayer, Dr.Med.Vet., MSc

Clinical Assistant Professor, Head of Exotics Service,

Cummings School of Veterinary Medicine, Tufts University, North Grafton, MA

Fred W. Quimby, VMD, PhD, DACLAM Director, Lab Animal Research Center, Rockefeller University, New York, NY

John Curtis Seely, DVM, DACVP

Veterinary Pathologist, Experimental Pathology Laboratories, Research Triangle Park, NC

Jo Ellen Sherow, BS, LATG

Director, Research Compliance, Ohio University, Athens, OH

Jerald Silverman, DVM, DACLAM

Professor and Director, Department of Animal Medicine, University of Massachusetts Medical School, Worcester, MA

Michael K. Stoskopf, DVM, PhD, DACZM

Professor and Director of Environmental Medicine Consortium,
College of Veterinary Medicine, North Carolina State University, Raleigh, NC

Facility Operations Manager, Boehringer Ingelheim Pharmaceuticals Inc., Ridgefield, CT

Robert H. Weichbrod, PhD, MBA, RLATG Animal Program Administrator, National Eye Institute, NIH, Bethesda, MD

Axel Wolff, MS, DVM

Director, Division of Compliance Oversight, OLAW, NIH, Bethesda, MD

## Trying out tramadol in rats

Zegre Cannon et al. evaluated the applicability of tramadol as a practical and effective analgesic in male Sprague Dawley rats. To measure the efficacy of four dosages and three routes of administration, they used the hot-plate test and the tail-flick test. The intraperitoneal route of administration was effective at dosages of 12.5 mg, 25 mg and 50 mg tramadol per kg body weight, though sedation was observed at dosages of 25 mg and 50 mg per kg body weight. There were no notable side effects after intraperitoneal administration of 12.5 mg tramadol per kg body weight. The authors plan to further study this dosage and route of administration in rodents.

See page 342

## Long-term catheter placement

Research using rats sometimes requires long-term placement of catheters in the subarachnoid space, the cavity between the arachnoid mater and the pia mater in the brain. To date, published techniques for penetrating the subarachnoid space of small experimental animals require the use of inflexible or relatively inflexible catheters, which are not ideal for long-term placement in the subarachnoid space. Ehlert and colleagues describe a reliable procedure for long-term catheterization of the subarachnoid cavity of the rat. This new approach allows personnel to repeatedly use the catheter for a period of at least 2 weeks.

See page 352

## Manipulation of light:dark cycles

Delaney and colleagues designed and built a device that can house mice or rats and allow researchers to control the light:dark cycles inside. The chamber, which (when closed) completely blocks outside light, contains two units. Each unit can hold eight small mouse cages or six rat cages and contains an optical sensor that triggers an audible and visual alarm when light is detected. Researchers have reported that this chamber is ideal for use in work involving manipulations of light:dark cycles.

See page 360