

Table 2 Transfer of RP41 from *A. vinelandii* to *E. coli* K12

<i>A. vinelandii</i> donor strains	<i>E. coli</i> recipients	Selection	Transfer frequency	Exconjugant phenotype
UW100(RP41)	JC5466	His ⁺	3.2 × 10 ⁻⁷	7/8 Km ^R Tc ^R Nif ⁺
		Tc ^R	2.0 × 10 ⁻⁷	1/7 His ⁺ Nif ⁺ , 7/7 Km ^R
		Km ^R	1.4 × 10 ⁻⁷	0/9 His ⁺ Nif ⁺ , 9/9 Tc ^R
UW100(RP41)	SB1801	His ⁺	3.0 × 10 ⁻⁴	28/28 Nif ⁺ Tc ^R
		Km ^R	3.0 × 10 ⁻²	28/28 Tc ^R , 0/28 His ⁺ Nif ⁺
UW91(RP41)	SB1801	His ⁺	5.0 × 10 ⁻⁶	8/8 Km ^R Nif ⁺
		Km ^R	1.0 × 10 ⁻⁵	8/8 Tc ^R , 0/8 Nif ⁺ His ⁺

Plate matings as described in text. Genotype of JC5466 is *trp his recA56 spc*; Genotype of SB1801 is *his750* (deletion extending through *his* and *RHA-2A*) *str* λ^R *ara gal malA xyl mtl* (λ⁻). Symbols as in legend to Table 1.

was pleiotropically negative for both component activities. The presence of RP41 *nif* genes in one of each class of Nif⁺ exconjugants, UW91(RP41) and UW100(RP41), was confirmed by using these strains as donors and observing transfer of *his*, *nif* and drug resistance markers back to *E. coli* (Table 2). Some segregation of markers occurred in these matings: exconjugants selected directly for drug resistance were mainly His⁻ and Nif⁻ while all but one of the His⁺ exconjugants were Nif⁺ and drug resistant. Such segregation has been observed in other transfers involving *nif* plasmids^{3,6}.

Loss of RP41 *nif* genes ought to restore the Nif⁻ phenotype in these exconjugants. One *Azotobacter* strain UW91 (RP41) was cultured in non-selective conditions and spontaneous loss of *nif* genes occurred: After 10 subcultures in Burk's medium supplemented with 2 mg ml⁻¹ ammonium acetate, all clones among 52 tested were Nif⁻ and 28, screened for drug resistance, were resistant to Tc, Km and Carb. These drug-resistance markers, which originated on RP4, the parent plasmid of RP41 (ref. 2), were stable in *A. vinelandii*.

One *Azotobacter* strain, UW100(RP41) was examined for immunological evidence for the expression of *Klebsiella nif* structural genes. Crude extracts of this strain, grown in Burk's medium, contained material which cross reacted antigenetically with antiserum prepared against purified *K. pneumoniae* nitrogenase molybdoprotein (KpI). This antigen was absent from extracts of the parental strain, UW100, grown in ammonium-limited conditions (100 μg N (as ammonium acetate) ml⁻¹ Burk's medium). Material cross reacting with antiserum to purified *Azotobacter* nitrogenase molybdoprotein (AvI) was, however, present in both extracts. When these strains were grown with excess NH₄⁺ ion, no such cross-reacting materials were detected in extracts prepared similarly.

We conclude from these experiments that the Nif⁺ *Azotobacter* exconjugants carried RP41 and expressed its *nif* genes, which originated in *Klebsiella pneumoniae* strain M5a1 (ref. 2). The ability of *Azotobacter* to transcribe and translate *Klebsiella nif* genes and, in contrast to *Agrobacterium*³, actually to make use of their products has several important consequences:

(1) The regulatory apparatus, whereby ammonia repressed nitrogenase synthesis in parents and exconjugants, is common to *Klebsiella* and *Azotobacter*, even at a molecular level. For example, if glutamine synthetase (GS) is accepted as a positive activator of *nif* genes⁷⁻⁹, then *Azotobacter* GS can activate *Klebsiella nif* genes.

(2) *Klebsiella nif* genes normally function only in an anaerobic environment, yet transfer to the obligate aerobe *Azotobacter* permitted their expression in air. Clearly they must have shared the oxygen exclusion processes¹⁰ of *Azotobacter*.

(3) The possibility arises that hybrid nitrogenases, such as the active mixtures of AcI+KpII or KpI+AcII which can be studied *in vitro*¹¹, are functioning *in vivo*, but comparable experiments with characterised mutants of RP41 would be necessary to establish this point conclusively.

RP41 is thus potentially useful for a genetic complementation analysis of *nif* mutants in *Azotobacter*. It could also be used for the isolation of selected mutants in *Azotobacter* by introducing RP41 with characterised *nif* mutations into *Azotobacter* and preparing homogenates.

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Errata

In the article "Cell differentiation by 3',5'-cyclic AMP in a lower plant" by A. K. Handa and M. M. Johri (*Nature*, **259**, 480; 1976) there is an error in Table 3. The entries under the heading 'Medium' should read . . . MM, MM+1% glucose, MM+1 μM IAA, MM+1% glucose . . . and not as printed.

In the paper "Dopamine-like renal and mesenteric vasodilation caused by apomorphine, 6-propylnorapomorphine and 2-amino-6,7-dihydroxy-1,2,3,4-tetrahydronaphthalene", by H. J. Crumley, R. M. Pinder, W. B. Hinshaw and L. I. Goldberg (*Nature*, **259**, 584; 1976) Dr Pinder's address should be: . . . Birkenhead, Auckland, New Zealand.

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