S-100 Protein in Embryonic Chick Retinae

EMBRYONIC differentiation presumably reflects differ-ential gene expression¹. It has been pointed out that the retina is particularly well suited for studies of these phenomena^{2,3} because of its ease of isolation, lack of vascularization, discrete origin and well studied pattern of morphogenesis4,5.

Gene expression in the differentiating cells of the nervous system has been most conveniently investigated by analysis of enzyme activity^{6,7}. For systems in which the enzyme pathways have not been worked out immunochemical methods can be used. Like the enzyme assays, immunochemical methods are specific and quantifiable, and are applicable to non-catalytic substances as well as enzymes; the availability of purified antigens and highly specific antisera makes them easy to use. These conditions were met by the isolation and purification of a highly acidic protein, termed S-100, unique to the nervous system⁸, and by the subsequent production of antibodies against this protein.

We have traced the pattern of accumulation in the embryonic chick retina of this well characterized "endproduct" protein of no known enzyme activity, and related this pattern to metabolic changes which occur during differentiation.



Fig. 1. Accumulation of S-100 protein in embryonic chick retinae. Vertical lines indicate range of values obtained. Figures beneath these indicate number of retinae tested. H-II=IIamburger Hamilton stage. There was 2 μ g total protein per tube.

Antisera to purified beef S-100 were prepared in rabbits as previously described⁹. Embryos from New Hampshire Red eggs incubated at 38°-39° C in a forced draft incubator were removed at various times, floated in saline to remove adhering yolk and staged according to the Hamburger and Hamilton stage series¹⁰. The lower temporal quadrant of the retina of each eye was excised under a stereoscopic microscope using watchmaker's forceps and iridectomy scissors. The pigment and neural layers were separated and the neural retinae were immediately homogenized in micro-glass homogenizers using 130 μ l. of cold, triple distilled water.

The homogenates were divided into small aliquots, stored at -80° C and each thawed only once for determination of protein¹¹ and assay of S-100. The relative concentration of S-100 in the embryonic chick retinae was determined by a micro-complement fixation assay⁹, and the degree of lysis was determined by spectrophotometric measurement of released haemoglobin. Because absolute concentrations of S-100 could not be determined in this assay we expressed results as amount of fixation

(decrement in optical density of haemoglobin from lysed erythrocytes) relative to total retinal protein.

After dropping from an initial peak at 7.75 days, retinal S-100 concentration, relative to total protein, increases by a factor of about five between 15 days of incubation, when it was minimal, and 18 days of incubation (Fig. 1). The increase in relative S-100 concentration continues until at least 2 weeks after hatching. To obtain an approximation of the absolute concentration of S-100 in chick retinae, known concentrations of purified beef S-100 were tested by complement fixation against the same fixed concentration of antiserum used in the retinal assays⁹. Chick retinal homogenates contain S-100 in amounts of the order of 2×10^{-6} g/ml. of homogenate at early embryonic stages and 1.5×10^{-5} g/ml. of homogenate at 18 days of incubation. The assumption has been made that brain and retinal S-100 are similar in immunological reactivity. These data establish that S-100 constitutes less than 0.1 per cent of the total retinal protein at 6.5 days and rises to about 0.5 per cent of the total by 19 days of incubation.

Statistical analysis of the peak between 15 and 16 days shows it to be significant at a level of P = 0.014 using the Mann-Whitney U test. The same test, applied to the decrease occurring between 7.75 and 15 days, yields a value of P = 0.067 which is the greatest level of significance obtainable for the sample sizes involved.

Although the present data are not directly comparable with values obtained by Friedman and Wenger⁹ on whole brain, the S-100 content of the retina at about 7 days relative to the eventual level (about 66 per cent) corresponds to the value of 7.75 day brain compared with the adult level. This suggests that the retina follows a pattern of S-100 accumulation similar to that of whole brain up to this time. From 7.75 to 15 days, however, the decrease in relative S-100 concentration suggests a more rapid accumulation of other proteins.

The rapid increase between 15 and 18 days coincides with increased glutamine synthetase³ and alkaline phosphatase¹² and occurs concurrently with the functional maturation of the eye². Cicero, Cowan and Moore (personal communication) have found a similar increase in S-100 content of the chick optic tectum between 16 and 18 days of incubation. This period seems critical for the differentiation of the visual system in the chick.

This work was supported in part by a grant from the University of Missouri, St Louis, to H. F. We thank Miss Mary Ruscha for technical assistance.

B. WENGER

Department of Anatomy, University of Saskatchewan. Saskatoon, Saskatchewan, Canada.

H. FRIEDMAN

Department of Biology, University of Missouri-St Louis, 8001 Natural Bridge Road, St Louis, Missouri

Received April 24, 1970.

- ¹ Stern, C., in *Analysis of Development* (edit. by Willier, B., Weiss, P., and Hamburger, V.), 151 (Saunders, Philadelphia, 1955).
- ² Coulombre, A. J., Amer. J. Anal., 96, 153 (1955).
- ³ Moscona, A. A., and Hubby, J. L., Develop. Biol., 7, 192 (1963).
- 4 Coulombre, A. J., Intern. Rev. Cytol., 11, 161 (1961).

- ⁵ Kirk, D. L., and Moscona, A., Develop, Biol., 8, 841 (1963).
 ⁶ Herrmann, H., and Tootle, M. L., Physiol. Rev., 44, 280 (1964).
 ⁷ Scarano, E., and Augusti-Tocco, G., Comp. Biochem., 28, 55 (1967).
 ⁸ Moore, B. W., Biochem. Biophys. Res. Commun., 19, 739 (1965). Friedman, H. P., and Wenger, B. S., J. Embryol. Exp. Morphol., 23, 289 (1970).
- ¹⁰ Hamburger, V., and Hamilton, H. L., J. Morphol., 88, 49 (1951).
- ¹¹ Lowry, O. H., Rosebrough, N. J., Farr, A. L., and Randall, R. J., J. Biol. Chem., 193, 265 (1951).
- 12 Moog, F., Ann. NY Acad. Sci., 55, 57 (1952).