

<https://doi.org/10.1038/s44264-024-00016-2>

Towards an agroecological approach to crop health: reducing pest incidence through synergies between plant diversity and soil microbial ecology



Miguel A. Altieri¹, Clara I. Nicholls², Giovanni Dinelli³ & Lorenzo Negri³ ✉

Given environmental, economic, and social costs of unilateral chemical and biotechnological interventions to control pests, there is an urgent need to transition towards a knowledge-intensive holistic approach emphasizing agroecosystem design and management. The focus will be on what makes agroecosystems susceptible and vulnerable to insect pests, pathogens and weeds, in order to design diversified agroecosystems that prevent and suppress insect pest, pathogen and weed problems. We propose a plant health model applicable to agroecosystems that feature biodiversity enhanced designs and soils rich in organic matter and microbial life, managed with low chemical loads. In such diversified farming systems, the general protection of the plant is a consequence of mutualistic above and below ground relationships between plants, insects, and soil microbial communities. From a practical standpoint, the approach involves (a) restoring plant diversity at the landscape and field level, with spatial and temporal crop combinations that deter pests and/or enhance natural enemies and (b) increasing soil organic matter through green or animal manures, compost and other amendments, which enhance antagonists that control soilborne pathogens. Polycultures promote a complex root exudate chemistry which plays an important role in recruitment of plant-beneficial microbes, some of which enhance plants' innate immune system. Unleashing biotic interactions between plant diversity and increased microbial ecological activity generate conditions for the establishment of a diverse and active beneficial arthropod and microbial community above and below ground, essential for pest/disease regulation.

The Integrated Pest Management (IPM) concept arose in the early 1970s in response to concerns about impacts of pesticides on human health and the environment¹. By providing an alternative to the chemical control strategy, it was envisioned that ecological theory should provide a basis for predicting how specific changes in production practices and inputs might affect pest problems, and thus aid in the design of agricultural systems less vulnerable to pest outbreaks². In such systems pesticides would serve strictly as back-ups of natural regulation processes. Unfortunately, most IPM programs failed to put ecologically based theory into practice and deviated to advocate for “silver bullet” schemes to control pests, emphasizing unilateral interventions based on agrochemicals, pheromones and transgenic crops, at the

expense of more desirable cultural practices and biological control regulation processes³.

In the last two decades, calls for more holistic approaches to management of diseases⁴ insects⁵ and weeds⁶ have resurfaced in the literature, urging for a shift from a responsive, symptom-based, linear approach relying on biocides to a proactive, knowledge-intensive holistic approach emphasizing agroecosystem design and management based on cultural and biological controls. The focus is still on the pest organisms: understanding their biology and why herbivores quickly adapt and succeed in agroecosystems, in order to identify weak links in their life cycles and thus refine control measures. Despite these new perspectives, there is a need to rethink

¹Department of Environmental Science, Policy and Management, University of California, Berkeley, CA, USA. ²Global Studies, University of California, Berkeley, CA, USA. ³Department of Agricultural and Food Sciences, Alma Mater Studiorum - University of Bologna, Bologna, Italy. ✉ e-mail: lorenzo.negri4@unibo.it

the concept of crop protection for plant health using principles of the field of agroecology⁷, which focuses more on what makes agroecosystems susceptible and vulnerable to insect pests, pathogens and weeds, in order to design agroecosystems that prevent pest problems.

The old disease/pest triangle model predicts that for a disease to manifest, a susceptible host, a virulent pathogen and a favorable environment must occur simultaneously⁸. The lack of plant diversity and biologically poor soils of monocultures, plus pesticide applications create a “perfect storm” for pest infestations. A new plant health model needs to evolve applicable to agroecosystems that feature biodiversity enhanced designs managed with low chemical loads. In such diversified farming systems, the general protection of the plant is a consequence of mutualistic above and below ground relationships between plants, insects, soil microbial communities⁹.

An agroecologically based plant health approach considers that if the cause and appearance of a pest or disease is understood as imbalance, then the goal of treatments should be to restore balance and resilience of the agroecosystem. The focus of this approach is on the causes of pest and disease outbreaks—not treating symptoms (suppressing pests and diseases), which does not necessarily lead to plant health. Pest resilient agroecosystems can be achieved by restructuring and managing farm systems in ways that maximize the array of “built-in” preventive mechanisms and restore core regulatory systems, such as immune (biological pest regulation mechanisms) and metabolic (soil biological activity, organic matter dynamics and nutritionally balanced crops) functions. This preventative strategy relies on poorly explored synergies between plant diversity and the soil microbial community which are set in motion by polycultural patterns and addition of organic matter, key practices in the design of pest resilient and healthy agroecosystems¹⁰.

Why are agroecosystems vulnerable to pest and disease infestations?

Industrial agriculture has advanced at the expense of native vegetation leading to landscape simplification, by reducing habitat and food resources available to natural enemies of insect pests; this simplification reduces ecological services such as biological pest control. An emblematic example of these phenomena is biofuel-driven growth monoculture plantings in the US Midwest. The huge corn expansion has resulted in lower landscape diversity, altering the supply of aphid natural enemies, reducing biocontrol services by 24%. This soil use change cost soybean producers in these States an estimated \$58 million year⁻¹, in reduced yield and increased pesticide use¹¹.

A reduction of crop diversity was registered in the USA from 1978 to 2012, parallel to the intensification of the industrial agriculture mode, increasing possibilities for catastrophic insect pest outbreaks¹². Deploying one variety across vast agrolandscapes results in low genetic diversity, increasing the vulnerability of crops to diseases¹³. Insecticide applications create secondary insect pest outbreaks and/or resurgence due to elimination of natural enemies or creation of resistance¹⁴. More than 550 arthropod species have resistance to at least one insecticide and many pathogens have evolved resistance to fungicides within two years of use, highlighting the limits of the chemical control strategy¹⁵.

The efficacy of insect-resistant genetically modified organisms (GMO) crops, expressing δ -endotoxin genes from *Bacillus thuringiensis* (BT) has been questioned because of the potential for pest populations to evolve resistance¹⁶. There are also documented negative impacts on predators at higher trophic levels thus potentially affecting biological control processes^{17,18}. In addition, due to lower insecticide applications in BT crops, secondary pests that are not susceptible to the expressed toxin are becoming an increasing concern¹⁹. Moreover, many studies have demonstrated that the introduction of GMO crops have not reached the desired reduction of pesticides use, especially herbicide resistant crops, thereby not resulting in substantial environmental benefits^{20,21}.

Worldwide use of inorganic fertilizers significantly increased between 1990 and 2020 to about 200 million tons, representing a 46% increase since

1990. Nitrogen fertilizers contribute 56% (113 million tons) of the total, phosphorus fertilizers 24% (48 million tons) and potassium fertilizers the remaining 20% (39 million tons)²². Over-fertilizing crops can actually worsen pest problems as increasing soluble nitrogen levels in plants can decrease their resistance to pests, resulting in higher disease incidence and pest density and consequently crop damage²³. The nitrogen contents of crops grown on organic farms are often lower than those of conventional systems²⁴, suggesting that the lower foliar content of NO₃-N of organic crops may be a key factor in determining lower insect damage on crops fertilized with organic amendments. Apparently organic crops that are nitrogen-limited are often less attractive to herbivores potentially linked to the lower pest pressure often observed in organic systems²⁵. These findings have been also previously explained according to the *trophobiosis* theory^{26,27} which postulates that heavy applications of soluble nitrogen (N) fertilizers (and also certain pesticides) increase the cellular amounts of N, ammonia and amino acids, faster than the rate at which plants synthesize them for proteins. These reductions in the rate of protein synthesis result in temporary accumulation of free N, sugars and soluble amino acids in the foliage which boost growth and reproduction by insect herbivores and plant pathogens.

On the other hand, low soil organic matter content induces poor soil microbial communities and low populations of antagonists and mycorrhizal fungi, which exert suppressive effects on many soil borne pathogens²⁸. Also lack of crop diversity leads to poor root complexity and thus lower production of primary metabolites and exudates that play a key role in recruitment of plant-beneficial microbes²⁹.

Restoring plant diversity in agroecosystems

A central question in agroecology has been on how to manage biodiversity in cropping systems to influence agroecosystem function, particularly enhanced pest and disease regulation. Bolstering plant genetic and species diversity has been a cornerstone strategy of agroecosystem redesign to promote higher diversity of above and below ground-associated biota which in turn leads to more effective pest control and disease suppression³⁰.

Over the last 40 years, many studies have evaluated the effects of crop diversity on the abundance of insect pests. Results from 209 studies involving 287 pest species found that, compared with monocultures, the population of pest insects was lower in 52% of the studies, and higher in 15% of the studies. Of the 149 pest species exhibiting lower densities in intercropping systems, 60% were monophagous and 28% polyphagous species³¹. A meta-analysis involving 148 comparisons found out that farms with species-rich vegetational schemes exhibited a 44% increase in abundance of natural enemies, a 54% increase in pest mortality, and consequently a 23% reduction in crop damage when compared to monoculture farms³². Unequivocally, recent reviews and meta-analyses suggest that crop diversification strategies promote a combination of ecological mechanisms that lead to synergies and tradeoffs between natural enemy enhancement, reduction of insect pest densities, and reduced crop damage^{33–35}.

Various mechanisms have been offered to explain how intercropping leads to insect pest regulation: (1) individual plants are less apparent and are more difficult to find because they are dispersed in intercropped systems; (2) certain intercrop species will disrupt the ability of a pest to attack the main crop, or some crops might have a repellent effect on herbivores; (3) a more attractive intercrop draws the pest away from the main crop and (4) natural enemies are more abundant and efficient in diverse than in simple cropping systems^{30,33,36}. The functional push-pull system involving the intercropping of maize with a repellent plant and an attractive trap plant as a border crop, to control stemborers in Africa, adds a new dimension to the existing hypothesis³⁷. The regulation of pests was mediated by chemically mediated interactions involving release of attractant semiochemicals from the trap and repellent plants from the intercrops³⁸.

Plant pathologists have also observed that mixed crop systems can decrease pathogen incidence by slowing down the rate of disease development by host dilution, interception of propagules and spore deposition by the nonhost crop, alteration of wind velocity, direction, turbulence, or

creating environmental conditions less favorable to the spread of certain pathogens^{39–41}. Intercropping significantly reduces wind-borne fungal pathogens disease in comparison to monocultures, as it has been reported for *Uromyces appendiculatus*, the cause of bean rust, in mixed intercropping with maize (causing 51–25% reductions)⁴², or for *Alternaria solani* in tomatoes canopy, when intercropped with marigold (causing 64–73% reductions) or pigweed (causing 27–38% reductions)³⁹. Lower disease contributes to less crop damage and higher yields in mixed crops compared to corresponding monocultures.

Increasing genetic diversity leads to lower disease incidence, particularly rusts and powdery mildews of small grain crops, such as wheat⁴³. Crop variety mixtures restrict the spread of a disease relative to the mean of their components, provided that the components differ in their susceptibility, such as in the case of *Septoria nodorum*, a non-specialized and splash-dispersed pathogen and the powdery mildew pathogen, *Erysiphe graminis*, in wheat mixtures^{44,45}.

Temporal diversification through rotations decreases severity and damage of several fungal and nematodal root rot pathogens⁴⁶. Declines in pathogen abundances usually occur when non-host crops are included in the rotation. Rotation of beans with grain crops such as corn, wheat, barley, rye, or oat resulted in reduced root rot severity and increased yield. A minimum of 3-year rotation with barley was needed in fields with a history of severe potato root rot incidence⁴⁷. Similar effects were observed in Kidney beans plots that were planted to oats in the previous three seasons⁴⁸. In addition to granting enhanced nutrient provisioning to plants, improvement of soil physical properties, increases in soil C, rotations can increase soil microbial composition activity, responsible for suppressing the growth of certain plant pathogens⁴⁸.

Crop rotation strategies cut off or disturb population cycles of many low dispersal insect pests which overwinter in the host crop fields. Multi-layer rotations significantly reduced numbers of pests such as southern corn rootworm, *Diabrotica virgifera virgifera* LeConte, and the European corn borer, *Ostrinia nubilalis* Hübner, compared with no-rotations^{49,50}. Coincidentally, multi-year rotation systems showed higher populations of predators than short (2 years) rotation systems⁵¹.

Biodiversity enhancement at the landscape level is key to avoid loss of ecological service-providing organisms such as beneficial insects and soil biota components. New research shows that landscape configuration (spatial arrangement), in addition to composition, strongly affects natural enemy and pest populations. Natural enemies tend to be more abundant in fine-grained landscapes (comprised of smaller fields and habitat patches) and are influenced by the connectivity of crop fields to other habitat types⁵². Similarly soil bacterial and fungal communities, associated with sclerophyllous forests, can influence soil microbial composition of adjacent organic vineyards⁵³.

Restoring biological activity in soils

Copious additions of organic matter via green manures, compost, litter residues, etc., improve soil quality but also increase soil microbial populations including microorganisms, such as mycorrhizal fungi, species of *Pseudomonas*, *Fusarium*, *Trichoderma*, *Streptomyces*, and *Actinomyces*, known for their disease suppressiveness⁵⁴. Organic amendments, such as animal and green manure and composts can control diseases caused by soilborne pathogen species of *Rhizoctonia*, *Verticillium*, *Fusarium*, *Phytophthora*, *Pythium*, *Sclerotinia*, etc.⁵⁵. Antibiotics, antagonism via secondary metabolite production, iron-chelating siderophores, hyperparasitism, lytic enzymes, competition for space and nutrients are all likely to be responsible for disease suppression by these microbes⁵⁶. *Pseudomonas* spp. employ antibiosis and have been shown to suppress soil-borne diseases through production of antibiotic compounds such as 2,4-diacetyl phloroglucinol, phenazines, cyclic lipopeptides, and cyanides⁵⁶. Some plant-beneficial microbes suppress diseases via nutrient competition, particularly plant growth-promoting rhizobacteria (PGPR), which have evolved high-affinity siderophores that can suppress soil-borne pathogens through iron competition⁵⁷.

Another mechanism by which rhizobia, mycorrhizal fungi, and several plant growth-promoting rhizobacteria/fungi can suppress plant diseases is through enhancing the defensive capacity of the plant innate immune system, a phenomenon commonly known as induced systemic resistance (ISR)⁵⁸. Rhizobacteria of the genera *Pseudomonas* and *Bacillus* are well known for their antagonistic effects and their ability to induce resistance, mainly through the salicylic acid-dependent SAR pathway⁵⁹. Several resistance-inducing inoculants are being formulated and tested for more efficient disease biocontrol strategies⁶⁰. The application of such natural formulations represents an input substitution strategy which should be used, if necessary, to suppress certain pathogens during the agroecological transition, while assessing the risks that this may entail to agroecosystem functional biodiversity.

Root exudate chemistry (amino acids, simple organic acids and complex secondary metabolites) plays an important role in recruitment of plant-beneficial microbes. For example, benzoxazinoid metabolites in root exudates of maize are key in the recruitment of ISR-eliciting *Pseudomonas putida* bacteria⁶¹. Several researchers are elucidating ways to increase the ability of several cereal varieties to exude sufficient quantities of chemicals responsible for recruitment of ISR-eliciting PGPR⁶².

Although soil microbial effects on soil-borne pathogens are well known, soil microbial communities also play a role in depressing insect pest populations through changes in plant resistance. Root colonization by beneficial microbes can induce biosynthesis of plant defense-related compounds (flavonoids, lignin, and other secondary metabolites) against a variety of leaf-chewing as well as phloem-feeding insects through various plant hormonal signaling pathways⁶³. Organic management practices can alter soil microbial communities and plant defense potentials, for example through changes in salicylic acid (SA) levels and influence settling and performance of various insect pests. Tomatoes grown using conventional management are preferentially settled by leafhopper pests and have lower SA levels, compared to tomatoes grown using organic management⁶⁴.

Ecological weed suppression

Although weeds biologically interfere with crops in different ways than insects and pathogens, similar strategies of soil management and plant diversification are used to prevent and/or reduce their germination, growth and seed production.

Intercrops are often superior to monocultures in weed suppression, as crop combinations exploit resources more efficiently than sole crops, thus suppressing the growth of weeds more effectively through greater pre-emptive use of resources. Examples abound in the literature where various intercropping designs (fodder legumes/maize, cotton/soybean or sorghum, wheat/red clover, sorghum/cowpea, etc.) were effective to reduce densities of different narrow- and broad-leaved weed species^{65,66}.

Temporal designs also exert suppressive effects on weeds and many studies have shown that various crop × rotation combinations reduce weed plant and seed densities. The mechanisms responsible for the greater weed suppression in crop rotations is linked to the continuous soil disturbance which disrupts weed growth and through stimulation of germination, eventually depleting the soil weed seed bank⁶⁷. When cover crops or green manures are used in rotations for weed suppression, it is important to select species that exhibit high initial growth and establishment rates so that they rapidly develop a close dense canopy to cover the ground completely to smother and shade out weed species⁶⁸.

Another approach utilized in weed management systems is through the manipulation of allelopathic cover crop residues in annual and perennial cropping systems⁶⁹. Rye, sorghum, rice, sunflower, rape seed, vetch and wheat residues have been documented as having allelopathic effects affecting germination of small seeded weeds. After maturity, the residues of the mowed or rolled cover crops remain on the soil surface, and as they decompose release allelochemicals which suppress weeds⁷⁰. When crop residues like sorghum are left as a mulch, toxins are released onto the soil and diffuse only 2–3 cm in the soil profile. Small seeded crops and weeds planted into this toxic layer will not germinate due to the presence of

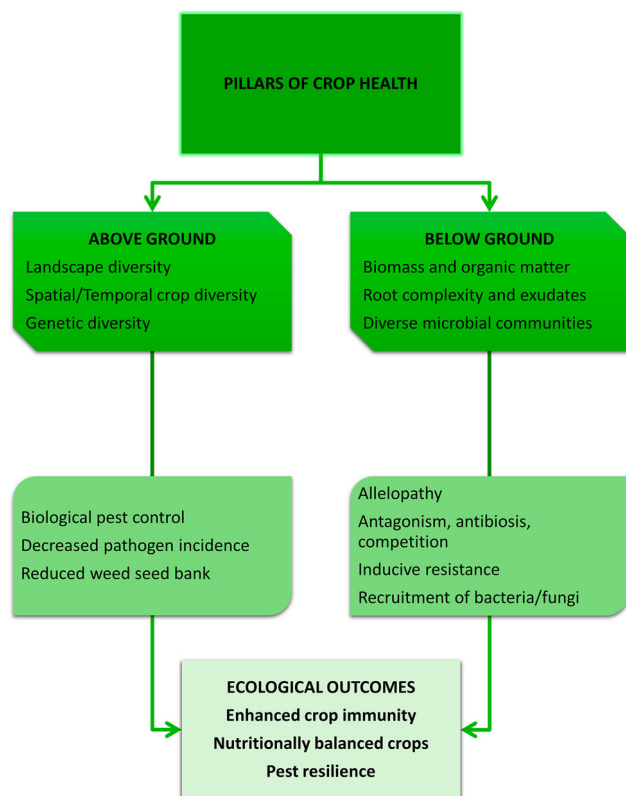


Fig. 1 | The pillars of plant health in agroecosystems. Exploiting the synergies between plant diversity and microbial community ecological activity.

allelochemical compounds such as sorgoleone, dhurrin, and other compounds⁷¹. This is why farmers traditionally plant their large-seeded crops (corn, beans, etc.) deeper and thus germination and root growth occur in a less toxic environment⁷². Many farmers in Brazil transplant vegetables such as tomatoes, onions and others through the in situ mulch. The mulch hinders weed seed germination and seedling emergence, often for several weeks, sufficiently delaying the onset of weed growth until after the crop's minimum weed-free period, making post plant cultivation, herbicides or hand weeding unnecessary, yet main crops exhibit acceptable crop yields⁷³.

Many weed species are more effective and faster than crops in capturing nutrients applied in fertilizers, and thus increases in soil fertility may benefit growth canopy development with negative effects on crop yields. Reducing applications of chemical N fertilizers may reduce populations of weeds that are responsive to fertilization⁷⁴. Organic farming systems rely upon the use of organic fertilizers and amendments that typically release nutrients (especially N) at a slower rate compared with mineral fertilizers. Slower nutrient release from organic sources usually results in decreased weed competitive ability, but effects depend on crop and weed species, plant densities, critical period of weed competition and other factors⁷⁵.

Conclusions

Agroecosystems experience multiple disturbances each growing season (plowing, irrigation, pesticide, and fertilizer applications, harvesting, etc.) creating conditions for rapid colonization by insect pests, weeds and pathogens which are opportunist species, with high dispersal ability and rapid growth. In addition, low crop diversity and soils poor in organic matter lead to an impoverished community of co-existing beneficial organisms, reducing crop plants' resilience to tolerate stress. The goal of agroecology is to create ecologically vigorous agroecosystems, which are adaptable, and diverse enough to tolerate stress. This implies agroecosystem redesign, involving two main pillars: (a) establishment of a diversified plant ecological infrastructure which leads to suppression of pests via 'top-down'

enhancement of natural enemy populations or via resource concentration and other 'bottom-up' effects acting directly on the colonization of pests and pathogens and (b) management of soils rich in organic matter that harbor diverse soil microbial communities integrally involved in antagonism, promotion of plant growth and induced resistance (Fig. 1). The approach exploits the synergies between greater plant diversity and increased microbial community biomass and ecological activity. These two pillars generate conditions for the establishment of a diverse and active beneficial arthropod and microbial community above and below ground, essential for provisioning pest/disease regulation. Despite the importance of agroecosystem biodiversity in promoting plant health, IPM programs often do not sufficiently consider its enhancement, as this approach is knowledge rather than input intensive. Agroecological design promotes interactions that set in motion ecological processes such as biological control, and this is complicated to attain via technological recipes or input prescriptions common to IPM schemes.

An agroecological approach to plant health is definitely knowledge intensive and requires in-depth, context-specific insights of each agroecosystem in question, including soil ecology, biology of target pathogens and herbivores and their antagonists, and the effects of vegetation and soil management practices on ecological processes at the farm and landscape levels.

Received: 9 January 2024; Accepted: 22 March 2024;

Published online: 29 April 2024

References

- Southwood, T. & Way, M. J. Ecological background to pest management. In *Concepts of Pest Managements* (eds Rabb, R. L. & Guthrie, F. E.) 6–28 (North Carolina State University, 1970).
- Levins, R. & Wilson, M. Ecological theory and pest management. *Annu. Rev. Entomol.* **25**, 287–308 (1980).
- Altieri, M. A., Martin, P. & Lewis, W. A quest for ecologically based pest management systems. *Environ. Manage.* **7**, 91–99 (1983).
- van Bruggen, A. H., Gamiel, A. & Finckh, M. R. Plant disease management in organic farming systems. *Pest Manage. Sci.* **72**, 30–44 (2016).
- Kogan, M. & Jepson, P. *Perspectives in Ecological Theory and Integrated Pest Management* (Cambridge University Press, 2007).
- Gallandt, E. in *Recent Advances in Weed Management* (eds Chauhan, B. S. & Mahajan, G.) 63–85 (Springer, 2014).
- Altieri, M. A. *Agroecology: the Science of Sustainable Agriculture* (CRC Press, 2018).
- Vega, D., Ibarra, S., Varela Pardo, R. A. & Poggio, S. L. Agroecological management of crop diseases: a review. *Agroecol. Sustain. Food Syst.* **47**, 919–949 (2023).
- Altieri, M. A. & Nicholls, C. I. in *Managing for Healthy Ecosystems* (eds Rapport, D. J. et al.) 999–1010 (CRC Press, 2002).
- Wyckhuys, K. A., Tang, F. H. & Hadi, B. A. Pest management science often disregards farming system complexities. *Commun. Earth Environ.* **4**, 223 (2023).
- Landis, D. A., Gardiner, M. M., van der Werf, W. & Swinton, S. M. Increasing corn for biofuel production reduces biocontrol services in agricultural landscapes. *Proc. Natl Acad. Sci. USA* **105**, 20552–20557 (2008).
- Aguilar, J. et al. Crop species diversity changes in the United States: 1978–2012. *PLoS ONE* **10**, e0136580 (2015).
- Brown, A. H. D. & Hodgkin, T. in *Genetic Diversity and Erosion in Plants: Indicators and Prevention* (eds Ahuja, M. R. & Mohan Jain, S.) 25–53 (Springer International Publishing, 2015).
- Pimentel, D. & Perkins, J. H. *Pest Control: Cultural and Environmental Aspects* (CRC Press, 2019).
- Whalon, M. E., Mota-Sanchez, D. & Hollingworth, R. in *Global Pesticide Resistance in Arthropods* (eds Whalon, M. E., Mota-Sanchez, D. & Hollingworth, R. M.) 5–31 (CABI, 2008).

16. Chandrasena, D. I. et al. Characterization of field-evolved resistance to *Bacillus thuringiensis*-derived Cry1F δ -endotoxin in *Spodoptera frugiperda* populations from Argentina. *Pest Manage. Sci.* **74**, 746–754 (2018).
17. Hilbeck, A., Meier, M. & Trtikova, M. Underlying reasons of the controversy over adverse effects of Bt toxins on lady beetle and lacewing larvae. *Environ. Sci. Eur.* **24**, 1–5 (2012).
18. Gatehouse, A., Ferry, N., Edwards, M. & Bell, H. Insect-resistant biotech crops and their impacts on beneficial arthropods. *Philos. Trans. R. Soc. B Biol. Sci.* **366**, 1438–1452 (2011).
19. Catarino, R., Ceddia, G., Areal, F. J. & Park, J. The impact of secondary pests on *Bacillus thuringiensis* (Bt) crops. *Plant Biotechnol. J.* **13**, 601–612 (2015).
20. Benbrook, C. M. Impacts of genetically engineered crops on pesticide use in the U.S.—the first sixteen years. *Environ. Sci. Eur.* **24**, 24 (2012).
21. Bonny, S. Genetically modified herbicide-tolerant crops, weeds, and herbicides: overview and impact. *Environ. Manage.* **57**, 31–48 (2016).
22. FAO. Inorganic fertilizers – 1990–2020. FAOSTAT analytical brief 47. <https://doi.org/10.4060/cc0947en> (2022).
23. Altieri, M. A. & Nicholls, C. I. Soil fertility management and insect pests: harmonizing soil and plant health in agroecosystems. *Soil Tillage Res.* **72**, 203–211 (2003).
24. Fagard, M. et al. Nitrogen metabolism meets phytopathology. *J. Exp. Bot.* **65**, 5643–5656 (2014).
25. Zehnder, G. et al. Arthropod pest management in organic crops. *Annu. Rev. Entomol.* **52**, 57–80 (2007).
26. Martinez, D. A., Loening, U. E., Graham, M. C. & Gathorne-Hardy, A. When the medicine feeds the problem; do nitrogen fertilisers and pesticides enhance the nutritional quality of crops for their pests and pathogens? *Front. Sustain. Food Syst.* **5**, 701310 (2021).
27. Chaboussou, F. *Healthy Crops: A New Agricultural Revolution* (Jon Carpenter Publishing, 2004).
28. Panth, M., Hassler, S. C. & Baysal-Gurel, F. Methods for management of soilborne diseases in crop production. *Agriculture* <https://doi.org/10.3390/agriculture10010016> (2020).
29. Stefan, L., Hartmann, M., Engbersen, N., Six, J. & Schob, C. Positive effects of crop diversity on productivity driven by changes in soil microbial composition. *Front. Microbiol.* **12**, 660749 (2021).
30. Altieri, M. & Nicholls, C. *Biodiversity and Pest Management in Agroecosystems* (CRC Press, 2018).
31. Andow, D. A. Vegetational diversity and arthropod population response. *Annu. Rev. Entomol.* **36**, 561–586 (1991).
32. Letourneau, D. K. et al. Does plant diversity benefit agroecosystems? A synthetic review. *Ecol. Appl.* **21**, 9–21 (2011).
33. Jaworski, C. C. et al. Crop diversification to promote arthropod pest management: a review. *Agric. Commun.* <https://doi.org/10.1016/j.agrcom.2023.100004> (2023).
34. van der Werf, W. & Bianchi, F. Options for diversifying agricultural systems to reduce pesticide use: can we learn from nature? *Outlook Agric.* **51**, 105–113 (2022).
35. Ratnadass, A. et al. Synergies and tradeoffs in natural regulation of crop pests and diseases under plant species diversification. *Crop Protect.* **146**, 105658 (2021).
36. Maitra, S. et al. Intercropping—a low input agricultural strategy for food and environmental security. *Agronomy* **11**, 343 (2021).
37. Ndayisaba, P. C., Kuyah, S., Midega, C. A. O., Mwangi, P. N. & Khan, Z. R. Push-pull technology improves maize grain yield and total aboveground biomass in maize-based systems in Western Kenya. *Field Crops Res.* **256**, 107911 (2020).
38. Khan, Z. R., Midega, C. A., Bruce, T. J., Hooper, A. M. & Pickett, J. A. Exploiting phytochemicals for developing a ‘push-pull’ crop protection strategy for cereal farmers in Africa. *J. Exp. Bot.* **61**, 4185–4196 (2010).
39. Boudreau, M. A. Diseases in intercropping systems. *Annu. Rev. Phytopathol.* **51**, 499–519 (2013).
40. Roese, A. D., Zielinski, E. C. & May De Mio, L. L. Plant diseases in afforested crop-livestock systems in Brazil. *Agric. Syst.* **185**, 102935 (2020).
41. Ratnadass, A., Fernandes, P., Avelino, J. & Habib, R. Plant species diversity for sustainable management of crop pests and diseases in agroecosystems: a review. *Agron. Sustain. Dev.* **32**, 273–303 (2012).
42. Fininsa, C. Effect of intercropping bean with maize on bean common bacterial blight and rust diseases. *Int. J. Pest Manage.* **42**, 51–54 (1996).
43. Browning, J. A. & Frey, K. J. Multiline cultivars as a means of disease control. *Annu. Rev. Phytopathol.* **7**, 355–382 (1969).
44. Borg, J. et al. Unfolding the potential of wheat cultivar mixtures: a meta-analysis perspective and identification of knowledge gaps. *Field Crops Res.* **221**, 298–313 (2018).
45. Mundt, C. C. Use of multiline cultivars and cultivar mixtures for disease management. *Annu. Rev. Phytopathol.* **40**, 381–410 (2002).
46. Curl, E. A. Control of plant diseases by crop rotation. *Bot. Rev.* **29**, 413–479 (1963).
47. Peters, R. D., Sturz, A. V., Carter, M. R. & Sanderson, J. B. Developing disease-suppressive soils through crop rotation and tillage management practices. *Soil Tillage Res.* **72**, 181–192 (2003).
48. Peralta, A. L., Sun, Y., McDaniel, M. D. & Lennon, J. T. Crop rotational diversity increases disease suppressive capacity of soil microbiomes. *Ecosphere* **9**, e02235 (2018).
49. Bažok, R., Lemić, D., Chiarini, F. & Furlan, L. Western corn rootworm (*Diabrotica virgifera virgifera* LeConte) in Europe: current status and sustainable pest management. *Insects* **12**, 195 (2021).
50. Brust, G. E. & King, L. R. Effects of crop rotation and reduced chemical inputs on pests and predators in maize agroecosystems. *Agric. Ecosyst. Environ.* **48**, 77–89 (1994).
51. O’Rourke, M. E., Liebman, M. & Rice, M. E. Ground beetle (Coleoptera: Carabidae) assemblages in conventional and diversified crop rotation systems. *Environ. Entomol.* **37**, 121–130 (2014).
52. Haan, N. L., Zhang, Y. & Landis, D. A. Predicting landscape configuration effects on agricultural pest suppression. *Trends Ecol. Evol.* **35**, 175–186 (2020).
53. Castaneda, L. E., Godoy, K., Manzano, M., Marquet, P. A. & Barbosa, O. Comparison of soil microbial communities inhabiting vineyards and native sclerophyllous forests in central Chile. *Ecol. Evol.* **5**, 3857–3868 (2015).
54. Abawi, G. & Widmer, T. Impact of soil health management practices on soilborne pathogens, nematodes and root diseases of vegetable crops. *Appl. Soil Ecol.* **15**, 37–47 (2000).
55. Bonanomi, G., Antignani, V., Capodilupo, M. & Scala, F. Identifying the characteristics of organic soil amendments that suppress soilborne plant diseases. *Soil Biol. Biochem.* **42**, 136–144 (2010).
56. Jayaprakashvel, M. et al. Metabolites of Plant Growth-Promoting Rhizobacteria for the Management of Soilborne Pathogenic Fungi in Crops. In *Secondary Metabolites of Plant Growth Promoting Rhizomicroorganisms* (eds Singh, H. B. et al.) 293–315 (Springer, Singapore, 2019).
57. Abdelaziz, A. M. et al. Biocontrol of soil borne diseases by plant growth promoting rhizobacteria. *Trop. Plant Pathol.* **48**, 105–127 (2023).
58. Pieterse, C. M. et al. Induced systemic resistance by beneficial microbes. *Annu. Rev. Phytopathol.* **52**, 347–375 (2014).
59. Verma, P. P., Shelake, R. M., Das, S., Sharma, P. & Kim, J.-Y. in *Microbial Interventions in Agriculture and Environment: Volume 1: Research Trends, Priorities and Prospects* (eds Singh, D. P., Gupta, v. & Prabha, R.) 281–311 (Springer, 2019).
60. Olowe, O. M., Akanmu, A. O. & Asemoloye, M. D. Exploration of microbial stimulants for induction of systemic resistance in plant disease management. *Ann. Appl. Biol.* **177**, 282–293 (2020).

61. Neal, A. & Ton, J. Systemic defense priming by *Pseudomonas putida* KT2440 in maize depends on benzoxazinoid exudation from the roots. *Plant Signal. Behav.* **8**, e22655 (2013).
62. Vejan, P., Abdullah, R., Khadiran, T., Ismail, S. & Nasrulhaq Boyce, A. Role of plant growth promoting rhizobacteria in agricultural sustainability—a review. *Molecules* **21**, 573 (2016).
63. Pineda, A., Zheng, S.-J., van Loon, J. J., Pieterse, C. M. & Dicke, M. Helping plants to deal with insects: the role of beneficial soil-borne microbes. *Trends Plant Sci.* **15**, 507–514 (2010).
64. Blundell, R. et al. Organic management promotes natural pest control through altered plant resistance to insects. *Nat. Plants* **6**, 483–491 (2020).
65. Gu, C., Bastiaans, L., Anten, N. P. R., Makowski, D. & van der Werf, W. Annual intercropping suppresses weeds: a meta-analysis. *Agric. Ecosyst. Environ.* **322**, 107658 (2021).
66. Hamzei, J. & Seyed, M. Evaluation of the effects of intercropping systems on yield performance, land equivalent ratio, and weed control efficiency. *Agric. Res.* **4**, 202–207 (2015).
67. Koocheki, A., Nassiri, M., Alimoradi, L. & Ghorbani, R. Effect of cropping systems and crop rotations on weeds. *Agron. Sustain. Dev.* **29**, 401–408 (2009).
68. Osipitan, O. A., Dille, J. A., Assefa, Y. & Knezevic, S. Z. Cover crop for early season weed suppression in crops: systematic review and meta-analysis. *Agron. J.* **110**, 2211–2221 (2018).
69. Muzell Trezzi, M., Vidal, R. A., Balbinot Junior, A. A., von Hertwig Bittencourt, H. & da Silva Souza Filho, A. P. Allelopathy: driving mechanisms governing its activity in agriculture. *J. Plant Interact.* **11**, 53–60 (2016).
70. Jabran, K., Mahajan, G., Sardana, V. & Chauhan, B. S. Allelopathy for weed control in agricultural systems. *Crop Protect.* **72**, 57–65 (2015).
71. Weston, L. A., Alsaadawi, I. S. & Baerson, S. R. Sorghum allelopathy—from ecosystem to molecule. *J. Chem. Ecol.* **39**, 142–153 (2013).
72. Lemessa, F. & Wakjira, M. Cover crops as a means of ecological weed management in agroecosystems. *J. Crop Sci. Biotechnol.* **18**, 123–135 (2015).
73. Altieri, M. A. et al. Enhancing crop productivity via weed suppression in organic no-till cropping systems in Santa Catarina, Brazil. *J. Sustain. Agric.* **35**, 855–869 (2011).
74. Kaur, S., Kaur, R. & Chauhan, B. S. Understanding crop-weed-fertilizer-water interactions and their implications for weed management in agricultural systems. *Crop Protect.* **103**, 65–72 (2018).
75. Liebman, M. & Davis, A. S. in *Organic Farming: The Ecological System*, Vol. 54, (ed. Francis, C.) 173–195 (Wiley, 2009).

Author contributions

M.A.A. and C.I.N. co-conceived the idea. M.A.A. wrote the initial draft of this manuscript. C.I.N., G.D. and L.N. brought major elements to the discussion. M.A.A. and L.N. put together the core literature for the conception of the paper. M.A.A., C.I.N., G.D. and L.N. have reviewed and commented on the final version of the manuscript.

Competing interests

The authors declare no competing interests.

Additional information

Correspondence and requests for materials should be addressed to Lorenzo Negri.

Reprints and permissions information is available at <http://www.nature.com/reprints>

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

© The Author(s) 2024