

COMMENT OPEN



BclX_L (*Bcl2l1*) gene therapy lessens retinal ganglion cell soma loss but not axonal degeneration after acute axonal injury

Olivia J. Marola^{1,2,3}, Sarah E. R. Yablonski^{1,3,4}, Peter G. Shrager⁴, Robert W. Nickells⁵ and Richard T. Libby^{1,3,6}✉

© The Author(s) 2022

Cell Death Discovery (2022)8:331; <https://doi.org/10.1038/s41420-022-01111-4>

Glaucoma, a leading cause of irreversible blindness, is characterized by loss of retinal ganglion cells (RGCs). In glaucoma, RGCs are thought to sustain axonal injury at the glial lamina [1]. This injury triggers molecularly distinct cell death pathways governing degeneration of the RGC soma and the distal axon. Much work has elucidated the mechanisms controlling degenerative processes in both RGC compartments [2]. In ocular hypertensive DBA/2J mice and after acute mechanical RGC axonal injury (controlled optic nerve crush, CONC), the apoptotic molecule BAX was shown to be required for degeneration of the soma, but not distal Wallerian degeneration of the axon [3]. In contrast, manipulation of molecules important for axonal degeneration (e.g. expression of *Wld^Δ*) lessened death of the entire RGC in DBA/2J glaucoma [1]. Of note, after CONC (which allows independent analysis of the RGC somal and axonal compartments), *Wld^Δ* expression significantly delayed axonal degeneration but did not lessen RGC somal degeneration [4]—suggesting *WLD^Δ*'s activity is restricted to the RGC axon. Taken together, these data suggest axon-localized degenerative pathways ultimately drive degeneration of both RGC compartments in glaucoma. In contrast, there is evidence that effectors originating from the soma are important in initiating axonal degeneration after neurodegenerative injury [5], suggesting that the factor(s) governing both somal and axonal degeneration in glaucoma may be initially triggered in the soma. Elucidating the inciting mechanism(s) driving both somal and axonal degeneration after glaucoma-relevant injury will be important in the development of neuroprotective therapies.

Recently, it was shown that overexpression of *BclX_L* protected the entire RGC in DBA/2J glaucoma [6]. *BCLX_L* inhibits BAX induction and is the principal pro-survival family member of the *Bcl2* gene family expressed in RGCs [7]. *BclX_L* deletion significantly increased RGC death after CONC, suggesting *BCLX_L* activity protects RGCs after glaucoma-relevant injury [8]. *BCLX_L* was shown to localize to both somas and axons in dorsal root ganglion neurons [5]. Given this, it is possible that loss of *BCLX_L* activity from the RGC soma, axon, or from both compartments, drives RGC degeneration after glaucoma-relevant injury. Locating *BCLX_L*'s protective effect will aid in understanding the role of somal and axonal contributions to RGC degeneration in glaucoma. Here, we utilize CONC to investigate the protective effect of *BclX_L* overexpression in the RGC soma and axon compartments independently.

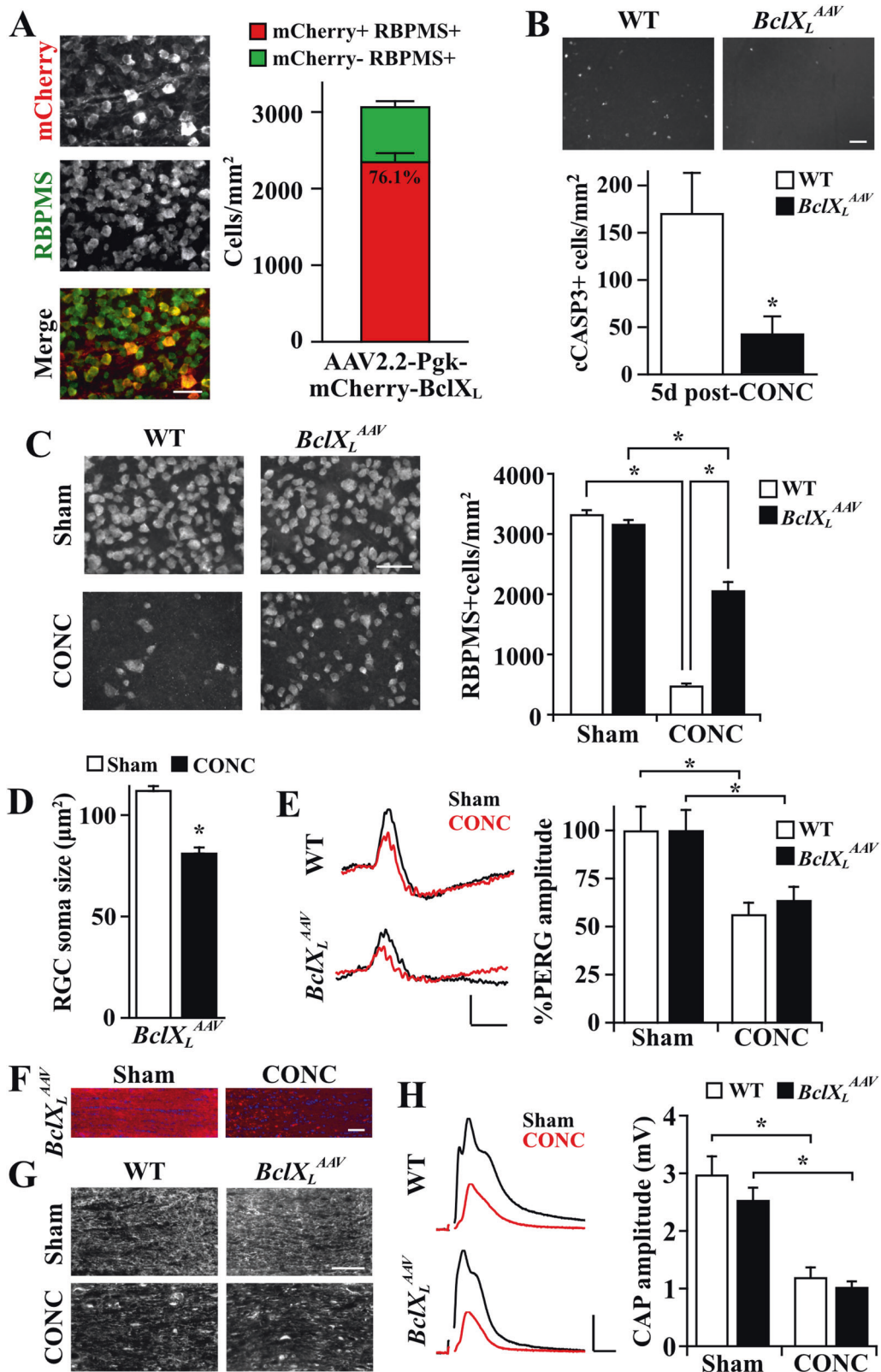
To study the compartment-specific effects of *BclX_L* overexpression after CONC, *BclX_L* was overexpressed (*BclX_L^{AAV}*) in the retinas of C57BL/6J mice (aged 3–7 months) by bilateral intravitreal delivery of AAV2.2-Pgk-mCherry-*BclX_L* vector, performed as previously described [6]. Control animals (WT) were bilaterally intravitreally injected with volume-matched PBS. Mice were randomly selected to receive intravitreal AAV2.2-Pgk-mCherry-*BclX_L* or PBS. Mice were fed chow and water *ad libitum* and housed on a 12-hour light-to-dark cycle. All experiments were conducted in adherence with the Association for Research in Vision and Ophthalmology's statement on the use of animals in ophthalmic and vision research and were approved by the University of Rochester's University Committee on Animal Resources. A priori exclusionary criteria included abnormal eye phenotypes (e.g. shrunken eye, cataracts, displaced pupil, lens damage). CONC (performed as previously described [9]) was done no earlier than 28 days after intravitreal injection to allow for sufficient transduction. To determine gross physiological function of RGC somas, pattern electroretinography (PERG) was performed using the Celeris Diagnosys system according to manufacturer's instructions. To assess physiological function of RGC axons, compound action potentials (CAPs) were recorded as previously described [4, 9] with peak amplitudes measured at 37 °C. Immunohistochemistry and imaging for retinal flat mounts and optic nerve longitudinal sections were performed as previously described [9] using antibodies against RBPMS (Genetex, GTX118619, 1:250), RFP (Chromotek, 5f8-100, 1:1000), cCASP3 (R&D, AF835, 1:1000), and Neurofilament (Millipore, AB5539SP, 1:1000). RBPMS+ cell counts and soma size measurements were performed using Image J. In all cases, experimenters were masked to experimental group and condition. Experimental groups had roughly equal numbers of males and females, were sex- and age-matched, and littermates were used wherever possible. Power analyses were performed a priori to determine appropriate sample sizes. Data are reported as mean ± standard error of the mean, and in all cases, data sets being compared had similar variances and met the assumptions of each statistical test used.

To determine the compartment-specific effect of *BclX_L* overexpression after mechanical axonal injury, CONC was performed on *BclX_L^{AAV}* and WT control mice. Of note, as assessed by the percentage of mCherry+ RBPMS+ cells, AAV2.2-Pgk-mCherry-*BclX_L* transduced ~76% of RGCs (Fig. 1A), consistent with previously

¹Department of Ophthalmology, Flaum Eye Institute, University of Rochester Medical Center, Rochester, NY, USA. ²Cell Biology of Disease Graduate Program, University of Rochester Medical Center, Rochester, NY, USA. ³The Center for Visual Sciences, University of Rochester, Rochester, NY, USA. ⁴Department of Neuroscience, University of Rochester, Rochester, NY, USA. ⁵Department of Ophthalmology and Visual Sciences, University of Wisconsin-Madison, Madison, WI, USA. ⁶Department of Biomedical Genetics, University of Rochester Medical Center, Rochester, NY, USA. ✉email: richard_libby@urmc.rochester.edu

Received: 13 May 2022 Revised: 28 June 2022 Accepted: 30 June 2022

Published online: 22 July 2022



published results [6]. Five days post-CONC, *BclX_L^{AAV}* retinas had significantly fewer dying (cCASP3+) RGCs (Fig. 1B), and 14 days post-CONC, had significantly improved RGC survival compared to WT controls (Fig. 1C). Therefore, consistent with previous reports [6, 10], *BclX_L* overexpression improved RGC somal survival after

axonal injury. These data suggest loss of BCLX_L activity in the soma contributes to RGC somal degeneration in glaucoma and could also possibly contribute to degeneration of the axonal compartment.

Strikingly, despite improved somal survival in *BclX_L^{AAV}* retinas, surviving *BclX_L^{AAV}* RGC somas were significantly shrunken 14 days

Fig. 1 *BclX_L* overexpression improved RGC somal survival but not axonal degeneration after CONC. **A** Transduction efficiency of AAV2.2-Pgk-mCherry-*BclX_L* in RGCs as assessed by the percentage of mCherry+ (red) RGCs (RBPMS + cells, green) depicted in retinal flat mounts. On average, $76.1 \pm 1.2\%$ of RGCs were colabeled with mCherry. $n = 4$. Scale bar, 50 μm . **B** WT ($n = 5$) and *BclX_L^{AAV}* ($n = 6$) retinal flat mounts and quantification of cleaved caspase 3 (cCASP3)+ cells 5 days post-CONC. *BclX_L^{AAV}* retinas had $74.9 \pm 10.5\%$ fewer cCASP3+ cells compared to WT controls. $*P = 0.030$, two-tailed t -test. Scale bar, 50 μm . **C** WT and *BclX_L^{AAV}* retinal flat mounts and quantification of RGCs (RBPMS + cells) 14 days post-CONC. Both WT and *BclX_L^{AAV}* retinas had significant RBPMS + cell loss after CONC compared to Sham controls ($85.4 \pm 0.8\%$ and $34.8 \pm 4.2\%$ loss respectively, $*P < 0.001$). However, *BclX_L^{AAV}* retinas had $59.2 \pm 4.9\%$ improved RGC survival after CONC compared to WT controls ($*P < 0.001$). $n = 5$, two-way ANOVA, Holm-Sidak's post hoc test. Scale bar, 50 μm . **D** Quantification of RBPMS + RGC soma size from *BclX_L^{AAV}* retinas 14 days after Sham and CONC. After CONC, surviving RGCs from *BclX_L^{AAV}* retinas were $27.5 \pm 2.2\%$ smaller compared to Sham controls. $n = 5$, $*P < 0.001$, two-tailed t -test. **E** Representative PERG traces and quantification of PERG amplitudes from WT and *BclX_L^{AAV}* eyes 14 days post-Sham ($n = 17, 18$, respectively) and CONC ($n = 18, 17$, respectively). WT and *BclX_L^{AAV}* eyes had significant reductions in PERG amplitude after CONC relative to Sham ($43.5 \pm 5.8\%$ and $36.2 \pm 6.6\%$ reductions respectively, $*P < 0.05$). *BclX_L^{AAV}* eyes did not have improved PERG amplitudes after CONC compared to WT controls ($P = 0.816$). Two-way ANOVA, Holm-Sidak's post hoc test. Scale bar: X: 100 ms, Y: 5 μV . **F** Longitudinal *BclX_L^{AAV}* optic nerve sections 5 days post-Sham and CONC. Sham *BclX_L^{AAV}* optic nerves had notable axonal mCherry labeling, which was markedly "beaded" and lost post-CONC. $n = 4$. Scale bar, 50 μm . **G** Longitudinal WT and *BclX_L^{AAV}* optic nerve sections 5 days post-Sham and CONC immunostained for neurofilament-H. *BclX_L^{AAV}* optic nerves had similar histological signs of degeneration after CONC compared to WT controls. $n = 4$. Scale bar, 50 μm . **H** Representative CAP traces and quantification of CAP amplitudes from WT and *BclX_L^{AAV}* optic nerves 5 days post-Sham and CONC. Both WT and *BclX_L^{AAV}* optic nerves had significantly decreased CAP amplitudes after CONC compared to Sham controls ($59.7 \pm 5.6\%$ and $59.3 \pm 3.7\%$ amplitude reductions respectively, $*P < 0.001$). After CONC, *BclX_L^{AAV}* optic nerves did not have improved CAP amplitudes compared to WT controls ($P = 0.582$). $n = 5$, two-way ANOVA, Holm-Sidak's post hoc test. Scale bar: X: 1 ms, Y: 1 mV. All numerical data are reported as mean \pm standard error of the mean. For graphs, bars represent the mean, and error bars represent standard error of the mean.

post-CONC compared to Sham controls (Fig. 1D), suggesting injury or metabolic stress [11, 12]. This somal shrinkage was also observed in *Bax* deficient RGCs after CONC [13]. In addition, *BclX_L* overexpression was not sufficient to prevent a decrease in PERG amplitude (which is thought to be reflective of RGC activity [14]) 14 days after CONC (Fig. 1E). Thus, while *BclX_L* overexpression improved RGC soma survival after CONC, RGC somas did not appear to retain normal function. These data imply the separable nature of the mechanisms governing RGC somal survival and retention of physiological function.

Given that *BclX_L* overexpression protected RGC axons and somas in a model of ocular hypertension [6], it remained important to distinguish whether somal BCLX_L confers protection to the RGC axon, or if axonal BCLX_L affords this protection. To investigate this, axonal degeneration of *BclX_L^{AAV}* and WT optic nerves was assessed after CONC. Of note, the BCLX_L fusion protein (mCherry) prominently co-localized to RGC axons in the optic nerve (Fig. 1F), as was shown previously [6]. Axonal health was assessed histologically (labeling for neurofilament-H) and electrophysiologically by measuring CAPs. *BclX_L* overexpression did not lessen histological hallmarks of RGC axonal degeneration (Fig. 1G), nor prevent CAP amplitude decline after CONC (Fig. 1H). Thus, *BclX_L* overexpression did not appear to elicit neuroprotective effects by acting in the RGC axon after glaucoma-relevant injury.

Taken together, these data suggest that the detrimental effect of BCLX_L loss may be localized to the soma in the context of glaucomatous injury. This implicates the importance of degenerative mechanisms initiated in the RGC soma in ultimately driving death of the entire RGC. Future work should elucidate the mechanisms by which loss of somal BCLX_L activity initiates axonal degenerative activity to further uncover the earliest drivers of glaucomatous neurodegeneration.

DATA AVAILABILITY

The datasets used in the current study are available from the corresponding author on reasonable request.

REFERENCES

- Howell GR, Libby RT, Jakobs TC, Smith RS, Phalan FC, Barter JW, et al. Axons of retinal ganglion cells are insulted in the optic nerve early in DBA/2J glaucoma. *J Cell Biol*. 2007;179:1523–37.
- Syc-Mazurek SB, Libby RT. Axon injury signaling and compartmentalized injury response in glaucoma. *Prog Retin Eye Res*. 2019;73:100769.
- Libby RT, Li Y, Savinova OV, Barter J, Smith RS, Nickells RW, et al. Susceptibility to neurodegeneration in a glaucoma is modified by *Bax* gene dosage. *PLoS Genet*. 2005;1:17–26.

- Fernandes KA, Mitchell KL, Patel A, Marola OJ, Shrager P, Zack DJ, et al. Role of SARM1 and DR6 in retinal ganglion cell axonal and somal degeneration following axonal injury. *Exp Eye Res*. 2018;171:54–61.
- Simon DJ, Pitts J, Hertz NT, Yang J, Yamagishi Y, Olsen O, et al. Axon degeneration gated by retrograde activation of somatic pro-apoptotic signaling. *Cell*. 2016;164:1031–45.
- Donahue RJ, Fehrman RL, Gustafson JR, Nickells RW. BCLX_L gene therapy moderates neuropathology in the DBA/2J mouse model of inherited glaucoma. *Cell Death Dis*. 2021;12:781.
- Levin LA, Schlamp CL, Spiedoch RL, Geszvain KM, Nickells RW. Identification of the BCL-2 family of genes in the rat retina. *Invest Ophthalmol Vis Sci*. 1997;38:2545–53.
- Harder JM, Ding Q, Fernandes KA, Cherry JD, Gan L, Libby RT. BCL2L1 (BCL-X) promotes survival of adult and developing retinal ganglion cells. *Mol Cell Neurosci*. 2012;51:53–9.
- Fernandes KA, Harder JM, John SW, Shrager P, Libby RT. DLK-dependent signaling is important for somal but not axonal degeneration of retinal ganglion cells following axonal injury. *Neurobiol Dis*. 2014;69:108–16.
- Malik JMI, Shevtsova Z, Bähr M, Kügler S. Long-term in vivo inhibition of CNS neurodegeneration by Bcl-X_L gene transfer. *Mol Ther*. 2005;11:373–81.
- Harris CA, Deshmukh M, Tsui-Pierchala B, Maroney AC, Johnson EM Jr. Inhibition of the c-Jun N-terminal kinase signaling pathway by the mixed lineage kinase inhibitor CEP-1347 (KT7515) preserves metabolism and growth of trophic factor-deprived neurons. *J Neurosci*. 2002;22:103–13.
- Deckwerth TL, Easton RM, Knudson CM, Korsmeyer SJ, Johnson EM. Placement of the BCL2 family member BAX in the death pathway of sympathetic neurons activated by trophic factor deprivation. *Exp Neurol*. 1998;152:150–62.
- Janssen KT, Mac Nair CE, Dietz JA, Schlamp CL, Nickells RW. Nuclear atrophy of retinal ganglion cells precedes the *bax*-dependent stage of apoptosis. *Investigative Ophthalmol Vis Sci*. 2013;54:1805–15.
- Porciatti V. Electrophysiological assessment of retinal ganglion cell function. *Exp Eye Res*. 2015;141:164–70.

ACKNOWLEDGEMENTS

This work was supported by National Institutes of Health (NIH) grants R01 EY018606 (RTL), F31 EY030739 (OJM), R01 EY030123 (RWN), and P30 EY016665 (RWN). RTL and RWN are supported by Research to Prevent Blindness, unrestricted grants to the Department of Ophthalmology at the University of Rochester Medical Center, and the Department of Ophthalmology and Visual Sciences at the University of Wisconsin-Madison, respectively. PGS is supported by the Schmitt Program in Integrative Neuroscience. The authors would like to acknowledge Alyssa Parker for her excellent technical assistance.

AUTHOR CONTRIBUTIONS

OJM, RWN, and RTL designed the experiments. OJM, SERY, and PGS performed the experiments and analyzed the data. OJM prepared the figure and wrote the manuscript. OJM, SERY, RWN, PGS, and RTL reviewed and edited the manuscript. All authors read and approved the final version.

COMPETING INTERESTS

The authors declare no competing interests.

ADDITIONAL INFORMATION

Supplementary information The online version contains supplementary material available at <https://doi.org/10.1038/s41420-022-01111-4>.

Correspondence and requests for materials should be addressed to Richard T. Libby.

Reprints and permission information is available at <http://www.nature.com/reprints>

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons license and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>.

© The Author(s) 2022