DOI: 10.1038/ncomms10863

OPEN

Correspondence: Quantitative evaluation of X-ray dark-field images for microcalcification analysis in mammography

Kai Scherer¹, Lorenz Birnbacher¹, Konstantin Willer¹, Michael Chabior¹, Julia Herzen¹ & Franz Pfeiffer¹

Nature Communications 7:10863 doi: 10.1038/ncomms10863 (2016); Published 22 Apr 2016

Wang *et al.*¹ have recently reported an X-ray grating interferometer imaging approach²⁻⁴ for mammography combining the information from the X-ray absorption and small-angle scattering signal. The authors claim that their approach can distinguish between type I (calcium oxalate dihydrate, $CaC_2O_4 \cdot 2H_2O$) and type II (calcium hydroxyapatite, $Ca_5(PO_4)_3(OH)$) microcalcifications. While such a differentiation would indeed be of great value for clinical mammography, several important deficiencies in the study put the main results and conclusions of the published article in question. The shortcomings in the published work became obvious, after we have unsuccessfully tried to reproduce the results in our own laboratory.

OMMUNICATIONS

To discriminate between type I and type II calcifications, Wang et al. use the ratio $r = \frac{\hat{s}}{\hat{\mu}} = i\frac{\hat{s}}{\hat{\mu}}$ (equation (4) in Wang et al.¹), where \overline{s} is the length-independent, effective X-ray scattering parameter and $\bar{\mu}$ is the effective X-ray attenuation coefficient $(\hat{\mu} = \int_0^L \mu(l) dl = \bar{\mu}L, \ \hat{s} = i \int_0^L s(l) dl = i\bar{s}L$, with *i* and *L* denoting a set-up-specific constant and the sample thickness, respectively). To test their hypothesis that type I and type II calcifications generally exhibit opposite absorption and scatter signals, they present (supposedly confirming) experimental results for a phantom made from calcium oxalate dehydrate and calcium hydroxyapatite powder (to mimic type I and type II calcifications, Fig. 1 in Wang *et al.*¹). However, while the obtained values may be correct for the specific powders used here, the experimental outcome cannot be generalized easily, as the small-angle scattering signal does not only depend on the chemistry and density of the sample, but also strongly on the micromorphology of the powder. Previously published theoretical and experimental results^{5–7} clearly demonstrate this strong dependence of the scatter signal (and thus the r-value) on the average size of the microstructures. Consequently, arbitrarily chosen powders (with respect to the average grain size) cannot reliably model microcalcifications in the human breast, if the actual size distribution is not taken into account (and matched to the one in a real human breast). More specifically, our calculations (based on⁶) even show that by varying the average size of the powder microstructures, one can actually obtain arbitrary r-values, regardless of the actual chemical and

density position. This is also reflected in a recent publication by Michel *et al.*⁸ which reports on a larger scattering signal in calcium oxalate dihydrate versus calcium hydroxyapatite, contradicting phantom results of Wang *et al.*¹

Second, but probably even more important, we have identified a major mistake in the analysis of the data from the real breast specimens (Figs 3 and 5 in Wang et al.¹), which render the main conclusions of the study highly questionable. In their evaluation of the r-value for various microcalcifications, Wang et al. have neglected the contribution of the underlying breast tissue. Correctly, the *r*-value has to be written as $r_{\rm mt} = \frac{\hat{s}_{\rm m} + \hat{s}_{\rm t}}{\hat{\mu}_{\rm m} + \hat{\mu}_{\rm t}}$, where the subscripts m and t denote contributions from the microcalcification and the tissue. While neglecting \hat{s}_t leads to a relatively small error in the r-value (as the scattering signal of tissue is relatively low), neglecting $\hat{\mu}_t$ leads to a large error and significantly falsifies the classification of the microcalcifications. Some exemplary results from a corresponding experiment in our lab (Fig. 1) highlight the issue. The blue and the red points represent pixels with \hat{s}_{m} and $\hat{\mu}_{m}$ values of two different microcalcifications, and they appear as a cloud with a slope corresponding to the $r_{\rm m}$ -value of this particular calcification. If now the contributions of the tissue $(\hat{s}_t \text{ and } \hat{\mu}_t)$ are neglected in the analysis, one obtains a slope (r-value) for the two clusters of $r_{1,Wang} = r_{1,mt} = 0.34 \pm 0.02$ and $r_{2,Wang} = r_{2,mt} = 0.35 \pm 0.01$, a very similar and small value in both cases (in agreement with Figs 3 and 5 in Wang et al.¹). However, when the contributions from the tissue are now correctly subtracted, the real calcification values (matching the data cloud) become $r_{1,m} = 6.63 \pm 0.18$ and $r_{2,m} = 2.48 \pm 0.07$. This means that Wang et al.'s analysis would have yielded an error of almost 2000% for r_1 and ~700% for r_2 , with the consequence of large classification errors, as demonstrated by the example above (before correction: $r_1 \approx r_2$, after tissue correction: $r_1 \gg r_2$). Because of this error in the analysis, the data presented by Wang *et al.*¹ can barely be associated with the calcifications themselves, but instead is mostly dominated by the attenuation of the breast tissue $(\hat{\mu}_t \gg \hat{\mu}_m)$, which renders a correct classification according to the hypothesis untenable. Accordingly, the presented r-values are small (0.3 < r < 1.0), whereas the real values obtained by a correct analysis show scatter dominated ratios (1.2 < r < 10).

¹Lehrstuhl für Biomedizinische Physik, Physik-Department & Institut für Medizintechnik, Technische Universität München, 85748 Garching, Germany. Correspondence and requests for materials should be addressed to F.P. (email: franz.pfeiffer@tum.de) or to K.S. (email: kai.scherer@mytum.de).



Figure 1 | Quantitative evaluation of microcalcification analysis in X-ray dark-field mammography. (**a**) Experimental absorption and (**b**) dark-field mammogram of a freshly dissected breast abladate with microcalcifications. (**c**) Scatter plots comparing the absorption $\hat{\mu}_m$ to scattering power \hat{s}_m of two exemplary microcalcifications cluster, as indicated by the blue and red frame in (**a**, **b**), respectively. An incorrect *r*-value is obtained ($r_{Wang} = r_{mt}$) if contributions of the underlying tissue are neglected in the analysis, since $\hat{\mu}_t \gg \hat{\mu}_m$.

In summary, we can conclude that the main claim of this article, namely the successful classification of different microcalcifications into type I and type II by this approach, is unjustified. Both, the experimental results of the phantom and the ones for the human breast samples, neglect major contributions to the image signal, and therefore render the main claim and specific experimental results and conclusions presented in this published study highly questionable.

Finally, we note that Wang *et al.* have neither discussed nor referenced related and partially contradicting, published results by other groups, in which detailed calculations and experimental verifications of the dependence of the scattering parameter on the sample microstructure are shown^{5–8}. Furthermore, the authors have disregarded the fact that the use of the different ratios between attenuation and scattering parameters has already been demonstrated for material⁹ or tissue discrimination¹⁰.

References

- 1. Wang, Z. et al. Non-invasive classification of microcalcifications with phasecontrast x-ray mammography. Nat. Commun. 5, 3797 (2014).
- Pfeiffer, F., Weitkamp, T., Bunk, O. & David, C. Phase retrieval and differential phase-contrast imaging with low-brilliance x-ray sources. *Nat. Phys.* 2, 258–261 (2006).
- Pfeiffer, F. et al. Hard x-ray dark-field imaging using a grating interferometer. Nat. Mater. 7, 134–137 (2008).

- Stampanoni, M. et al. The first analysis and clinical evaluation of native breast tissue using differential phase-contrast mammography. *Invest. Radiol.* 46, 801–806 (2011).
- Yashiro, W., Terui, Y., Kawabata, K. & Momose, A. On the origin of visibility contrast in x-ray Talbot interferometry. *Opt. Express* 18, 16890 (2010).
- Malecki, A., Potdevin, G. & Pfeiffer, F. Quantitative wave-optical numerical analysis of the dark-field signal in grating-based x-ray interferometry. *Europhys. Lett.* 99, 48001 (2012).
- Lynch, S. et al. Interpretation of dark-field contrast and particle-size selectivity in grating interferometers. Appl. Opt. 50, 4310–4319 (2011).
- Michel, T. *et al.* On a dark-field signal generated by micrometer-sized calcifications in phase-contrast mammography. *Phys. Med. Biol.* 58, 2713–2732 (2013).
- 9. Bech, M. et al. Quantitative x-ray dark-field computed tomography. Phys. Med. Biol. 55, 5529–5539 (2010).
- Schleede, S. *et al.* Emphysema diagnosis using x-ray dark-field imaging at a laser-driven compact synchrotron light source. *Proc. Natl Acad. Sci. USA* 109, 17880–17885 (2012).

Author contributions

K.S., M.C., J.H. and F.P. conceived the experiment. K.S., L.B. and K.W. performed the experiment. K.S., J.H. and F.P. wrote the manuscript with contributions from all co-authors.

Additional information

Competing financial interests: The authors declare no competing financial interests.

Reprints and permission information is available online at http://npg.nature.com/ reprintsandpermissions/

How to cite this article: Scherer, K. *et al.* Correspondence: Quantitative evaluation of x-ray dark-field images for microcalcification analysis in mammography. *Nat. Commun.* 7:10863 doi: 10.1038/ncomms10863 (2016).

This work is licensed under a Creative Commons Attribution 4.0 International License. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in the credit line; if the material is not included under the Creative Commons license, users will need to obtain permission from the license holder to reproduce the material. To view a copy of this license, visit http://creativecommons.org/licenses/by/4.0/